

# Effects of Dietary Probiotic (*Lactobacillus acidophilus*) on Hematology, Blood Biochemistry and Lipid Profile of Carbon Tetrachloride-Induced Toxicity in Rats

M.G.A.EL.Sayed<sup>1</sup>, Enas. A. H. Farag<sup>2</sup>, Heba. M. Nasr<sup>3</sup>

<sup>1</sup>Department of Pharmacology, Faculty of Veterinary Medicine Benha University Egypt

<sup>2</sup>Professor of Pharmacology at Animal Health Research Institute Benha branch, Egypt

<sup>3</sup>Hospital of Benha University, Faculty of Medicine Benha University Egypt

**Abstract:** Exposure to carbon tetrachloride induces acute and chronic hepatic injuries as well as renal injuries in rats. Therefore, the current study aimed to evaluate the effect of probiotic (*Lactobacillus acidophilus*) on hematology, blood biochemistry and lipid profile of healthy rat, also the protective role against carbon tetrachloride -induced toxicity in albino rat. Four groups with ten rats each group were used for this purpose; these groups included the control vehicle group that received saline daily for 30 days, probiotic group ( $0.009 \times 10^6$  CFU/ gram of rat body weight orally) daily for 30 days, carbon tetrachloride group (2.5ml/kg intraperitoneally twice per week for three week; the probiotic –carbon tetrachloride group. The results revealed that carbon tetrachloride significantly increased serum levels of alanine aminotransferase, aspartate aminotransferase, alkaline phosphatase, total bilirubin, cholesterol, triglyceride, urea and creatinine. In addition, there were substantial increase in lipid peroxidation (malondialdehyde), white blood cell, packed cell volume and level of glucose with significant decreases in albumin, total protein, creatinine kinase, hemoglobin and red blood cells. Carbon tetrachloride also caused histological changes in liver, kidney and spleen tissues. However, administration of probiotic ameliorated the carbon tetrachloride induced liver and kidney damage with improved hematological, lipid profile and glucose level.

## 1. Introduction

Carbon tetrachloride is a toxic substance and commonly used as a chemical intermediate, degreasing agent and dry cleaning fluid. It has been proven to have a highly hepatotoxic effect, as well as inducing nephrotoxicity. Carbon tetrachloride can lead to acute tubular necrosis in the kidney and damage to the liver, which leads to cirrhosis [1]. Its harmful effect on the liver and kidney occurs due to the carbon tetrachloride metabolites, toxic trichloromethyl, and trichloromethylperoxy radicals inherent in the cytochrome P450 system [2]. CCL4 induced hepatocellular damage as fatty degeneration, fibrosis and impairment of liver function causing leakage of aspartate aminotransferase and alanine aminotransferase into circulation and released into the blood stream [3] also rise level of alkaline phosphatase [4]. High in the level of total bilirubin than normal must have been due to liver damage and fibrosis [5]. Carbon tetrachloride treatment significantly decreased serum total protein, albumin and globulin level compared to control and these may result in decreased hepatic ability to synthesize protein. Albumin significantly decreased that associated with active cirrhosis and biliary liver damages [5].

The term "probiotics" is used to describe the kind of non-pathogenic microorganisms that are good bacteria that are either the same as or very similar to the bacteria that are already in your body. Probiotic is a live microbial feed supplement which beneficially affects the host animal by improving its intestinal microbial balance [6]. Probiotics containing food has the ability to influence body functions in order to reduce the risk of many diseases [7]. Some strains of *Lactobacillus acidophilus* may be considered to have

probiotic characteristics [8]. *Lactobacillus acidophilus* is species of gram positive bacteria in the genus *Lactobacillus* [9]. Probiotic are used medicinally as *Lactobacillus acidophilus* [10-11]. *Lactobacillus acidophilus* selected as a probiotic bacteria because criteria that suitable and are originate from human and other mammals. The dietary supplement with *Lactobacillus acidophilus* modulates immunity, release antioxidants and improve gastrointestinal functions [12]. *Lactobacillus acidophilus* on hepatocellular, preventing hepatocellular damage in experimentally induced colitis [13]. The supplementation of *Lactobacillus acidophilus* is found to be beneficial in altering the levels of lipid profile in hypercholesterolemic subjects [14]. Administration *Lactobacillus acidophilus* increase immunity as increase total white cells blood and thrombocytes in mice [15].

## 2. Materials and Methods

### 2.1 Chemicals

Probiotic is available as capsules, each capsule contains over 100 million active *Lactobacillus acidophilus* (including the naturally occurring metabolic product produced by lactobacilli) at the time of manufacture and it marked by Puritan's Pride as Probiotic Acidophilus®. In the present study probiotic group was treated with oral dose  $0.009 \times 10^6$  per gram of rat body weight [16] which equivalent to 37.50 mg/kg b.wt [17] in drinking water. Carbon tetrachloride obtained from the laboratory of toxicology faculty of veterinary, medicine Benha University, Pharmaceuticals industries (Cairo, Egypt). Total bilirubin, serum transaminases activities (AST and ALT), alkaline

phosphatase (ALP), total protein, albumin, blood creatinine, blood urea, creatinine kinase, blood cholesterol, blood triglyceride, L-malondialdehyde (MDA), blood glucose erythrocytic count, total leucocytic count, haemoglobin concentration and packed cell volume were kits were purchased from Diamond Company (Cairo, Egypt).

## 2.2 Animals and experimental design

40 male Wister albino rats weighting 200-280 g (age of rat 50~60 days) were obtained from animal house of Faculty of Veterinary Medicine, Benha University, Egypt. The animals were housed in 49x35 cm stainless steel wire mesh cages with bedding of ground wood chips at 21 °C. They were fed fresh-pelleted food and their water as placed in glass bottles of 500 ml. Rats were kept at a constant environmental and nutritional condition throughout the period of experiment. The animals were left for 15 days for acclimatization before the beginning of the experiment. The rats were randomly divided into main 6 groups. **Group (1):** Ten rats were administered saline only 0.2ml daily for 30 days. **Group (2):** Ten rats were administered probiotic (*Lactobacillus acidophilus*) orally at dose  $0.009 \times 10^6$  CFU/ gram of rat body weight for 30 days daily. **Group (3):** Ten rats were administered carbon tetrachloride 25% (dissolved in olive oil) intraperitoneally injection 2.5ml/kg b.wt twice per week for three week to induce hepatic and renal toxicity. **Group (4):** Ten rats were administered probiotic (*Lactobacillus acidophilus*) as group 2 followed by carbon tetrachloride intraperitoneally injection 2.5 ml/kg b.wt twice per week for three week to induce hepatic and renal toxicity.

## 2.3 Blood collection

Blood samples were taken at first, seventh and fourteenth day post-treatment in all groups after the end of administration. Two blood samples were taken from each rat in the group for both biochemical and hematological studies from median canthus of the eye. **The first blood sample** was collected without anticoagulant for separation of clear serum for biochemical analysis. **The second sample** of blood was collected in the test tube mixed with sodium Citrate 3.8% as anticoagulant, the sample was shake several times to ensure mixing of blood with anticoagulant. These blood samples were used for hematological studies to determine erythrocytic count and hemoglobin concentration.

## 2.4 Serum biochemical analyses

The collected sera were used for biochemical analysis to determine serum total bilirubin, serum transaminases activities (AST and ALT), alkaline phosphatase (ALP), total protein, albumin, blood creatinine, blood urea, creatinine kinase, blood cholesterol, blood triglyceride, L-malondialdehyde (MDA) and blood glucose.

## 2.5 Evaluation of oxidative stress markers

Lipid peroxidation was measured by determination of MDA content in liver and kidney homogenates.

## 2.6 Histopathology

The treated rats were sacrificed at first day, seventh day and fourteenth day. Specimens were collected from liver, kidney and spleen from each sacrificed tested rats and fixed directly in formalin 10% for at least 24 hours, then the sample were washed under running tap water followed by immersion in serial dilutions of ethyl alcohol. Specimens were cleared in xylene and embedded in paraffin at 56 degree in hot air oven for twenty four hours. Paraffin bees wax tissue block were prepared for sectioning at 4 microns thickness by slide microtome. The obtained tissue sections were collected on glass slides, deparaffinized and stained by hematoxylin and eosin stain for routine examination through the light microscope [18].

## 2.7 Statistical analysis

Statistical analysis was conducted with the Statistical Package for Social Science [19] to determine if variables differed between groups, according to [20]. The Shapiro-Wilk test was used to test the normal distribution of the data before statistical analysis was performed. Compare between means were conducted by one-way ANOVA and subsequent Duncan's multiple range test [21]. Probability values of less than 5% ( $P < 0.05$ ) were considered significant.

## 3. Results

Effect of probiotic ( $0.009 \times 10^6$  CFU/ gram of rat body weight) orally for 30 days and carbon tetrachloride 25% (2.5ml/kg body weight twice per week) intraperitoneally for three week on serum total bilirubin, aspartate aminotransferase level (AST), alanine aminotransferase level (ALT), alkaline phosphatase (ALP), total protein, albumin, globulin, blood creatinine, blood urea, creatinine kinase, blood cholesterol, blood triglyceride, L-malondialdehyde (MDA), blood glucose, erythrocytic count (RBCs), haemoglobin (Hb), total leukocytic (WBCs) and packed cell volume.

Values with different letters within different row differed significantly at ( $P \leq 0.05$ ).

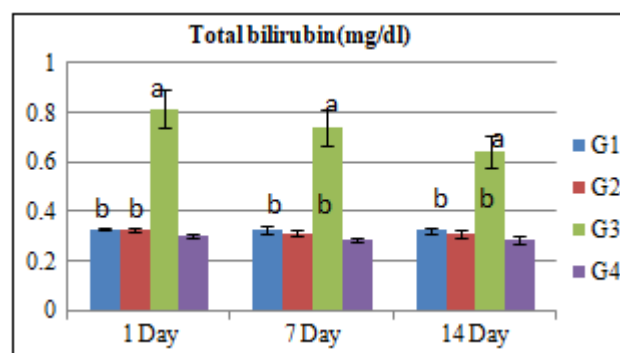


Figure 1: G (1): Control, G (2): Probiotic, G(3): CCL<sub>4</sub> G(4): Probiotic+CCL<sub>4</sub>

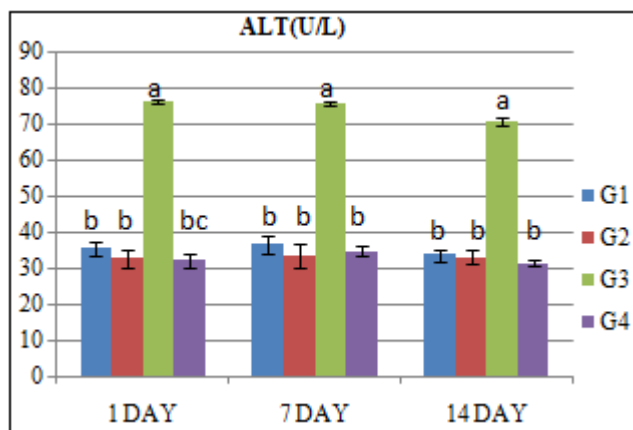


Figure 2: G (1): Control G (2): Probiotic G (3): CCL<sub>4</sub> G (4): Prbiotic+CCL<sub>4</sub>

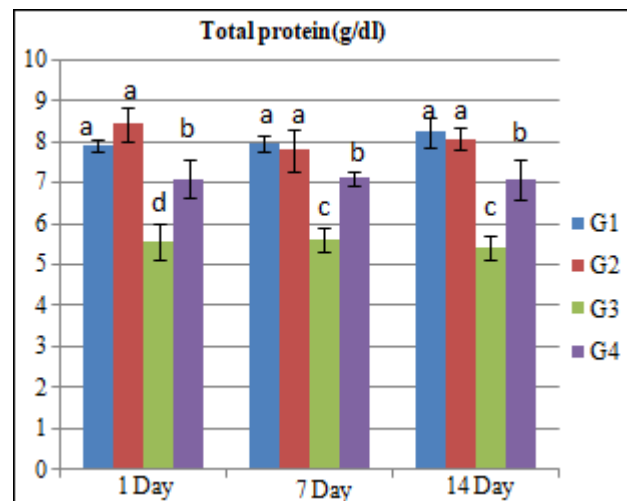


Figure 6: G (1): Control G (2): Probiotic G (3): CCL<sub>4</sub> G (4): Prbiotic+CCL<sub>4</sub>

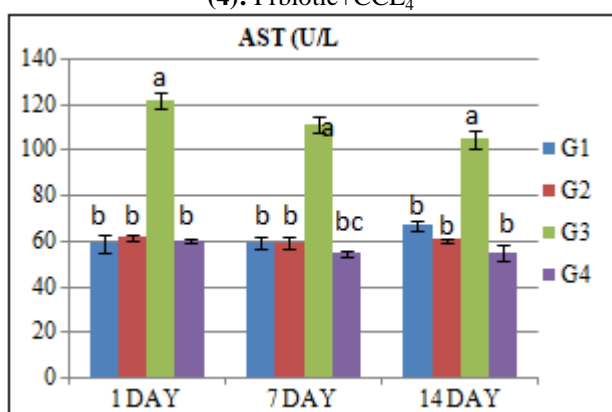


Figure 3: G (1): Control G (2): Probiotic G (3): CCL<sub>4</sub> G (4): Prbiotic+CCL<sub>4</sub>

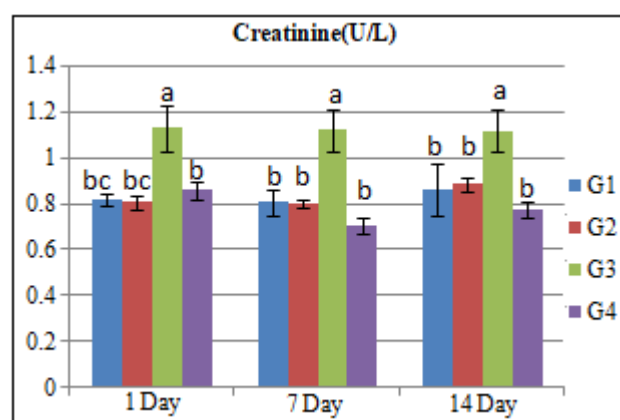


Figure 7: G (1): Control G (2): Probiotic G (3): CCL<sub>4</sub> G (4): Prbiotic+CCL<sub>4</sub>

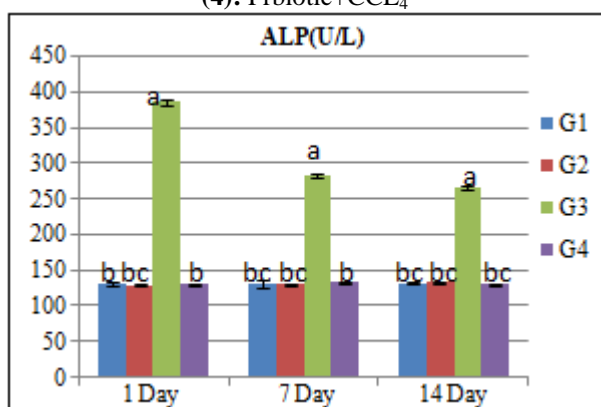


Figure 4: G (1): Control G (2): Probiotic G (3): CCL<sub>4</sub> G (4): Prbiotic+CCL<sub>4</sub>

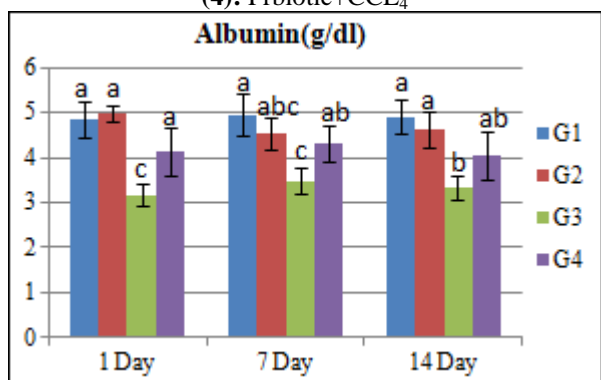


Figure 5: G (1): Control G (2): Probiotic G (3): CCL<sub>4</sub> G (4): Prbiotic+CCL<sub>4</sub>

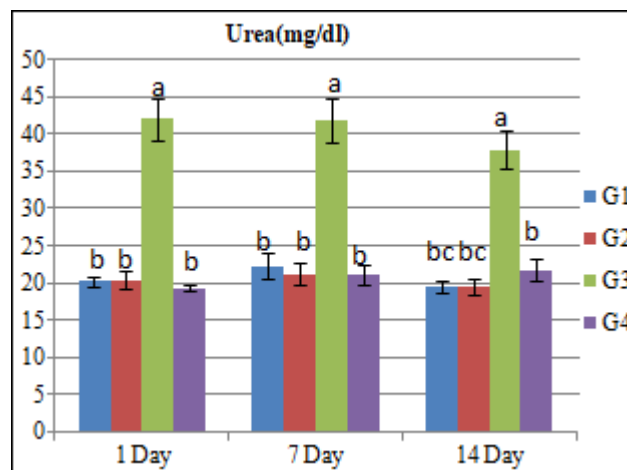


Figure 8: G (1): Control G(2): Probiotic G (3): CCL<sub>4</sub> G (4): Prbiotic+CCL<sub>4</sub>

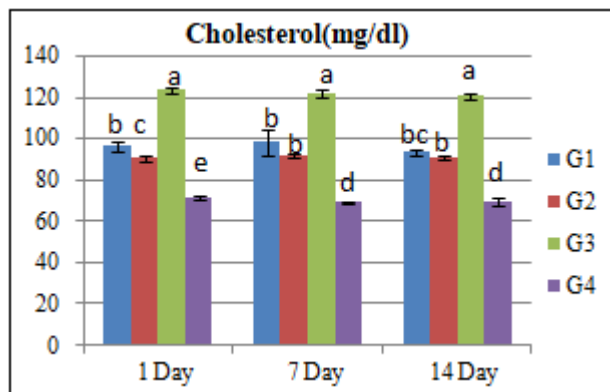


Figure 9: G (1): Control G(2): Probiotic G(3): CCL<sub>4</sub>G(4):Prbiotic+CCL<sub>4</sub>

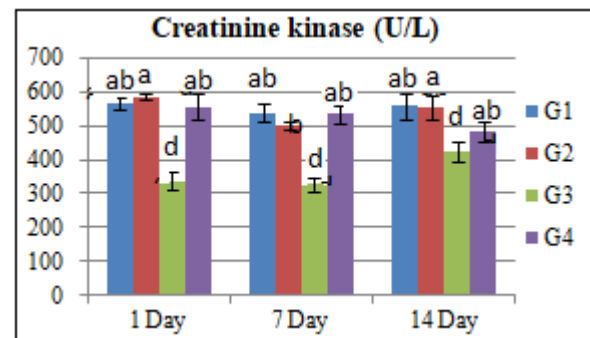


Figure 13: G (1): Control G(2): Probiotic G(3): CCL<sub>4</sub> G(4):Prbiotic+CCL<sub>4</sub>

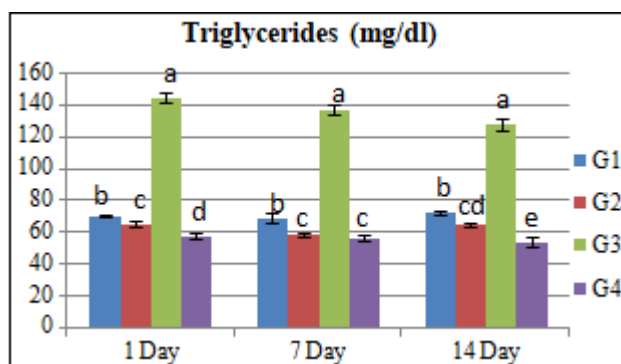


Figure 10: G (1): Control G(2): Probiotic G(3): CCL<sub>4</sub> G(4):Prbiotic+CCL<sub>4</sub>

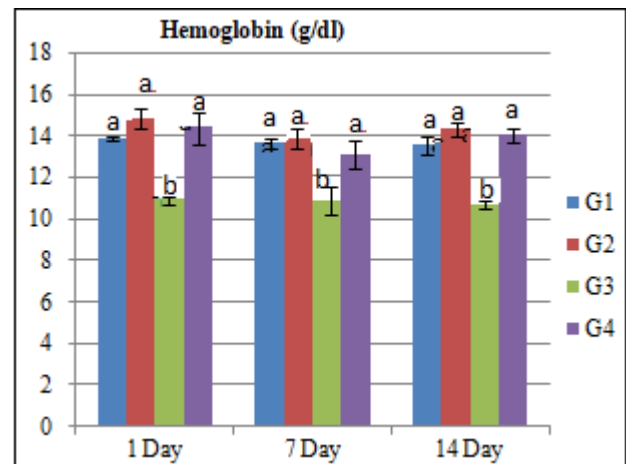


Figure 14: G (1): Control G(2): Probiotic G(3): CCL<sub>4</sub> G(4):Prbiotic+CCL<sub>4</sub>

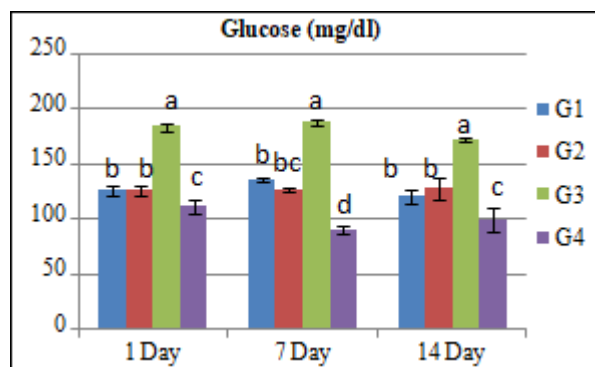


Figure 11: G (1): Control G(2): Probiotic G(3): CCL<sub>4</sub> G(4): Prbiotic+CCL<sub>4</sub>

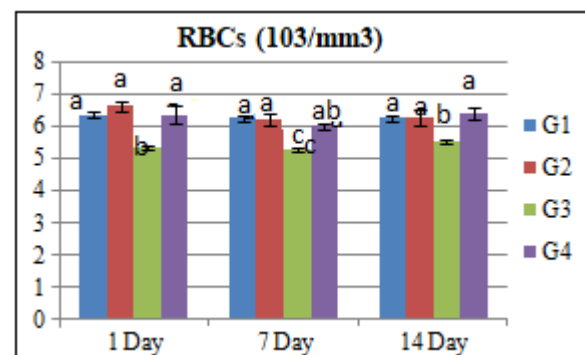


Figure 15: G (1): Control G(2):Probiotic G(3): CCL<sub>4</sub> G(4):Prbiotic+CCL<sub>4</sub>

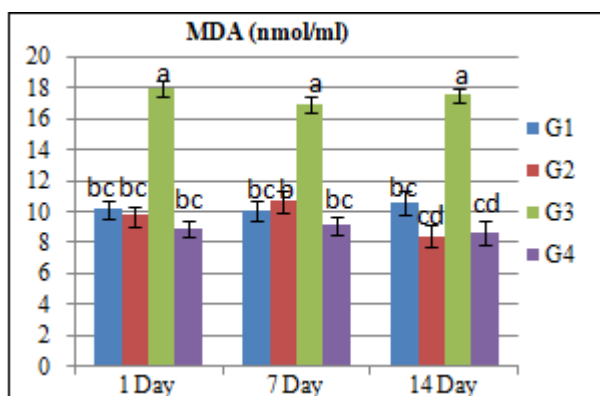


Figure 12: G (1): Control G(2): Probiotic G(3): CCL<sub>4</sub> G(4): Prbiotic+CCL<sub>4</sub>

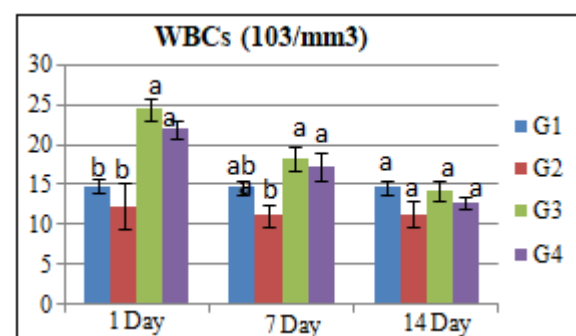


Figure 16: G (1): Control G(2): Probiotic G(3): CCL<sub>4</sub> G(4):Prbiotic+CCL<sub>4</sub>



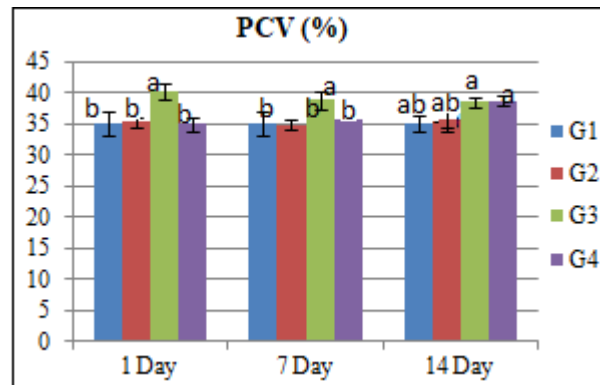
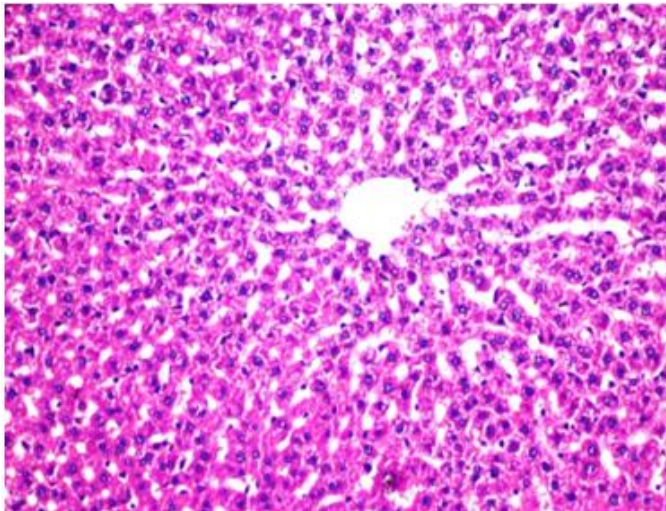
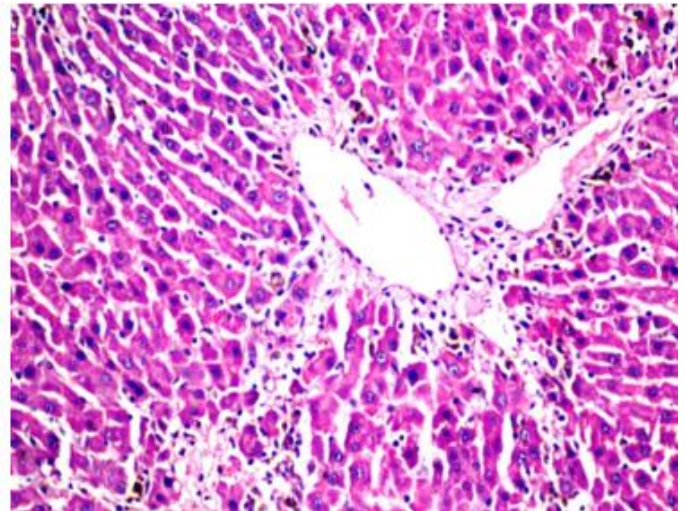


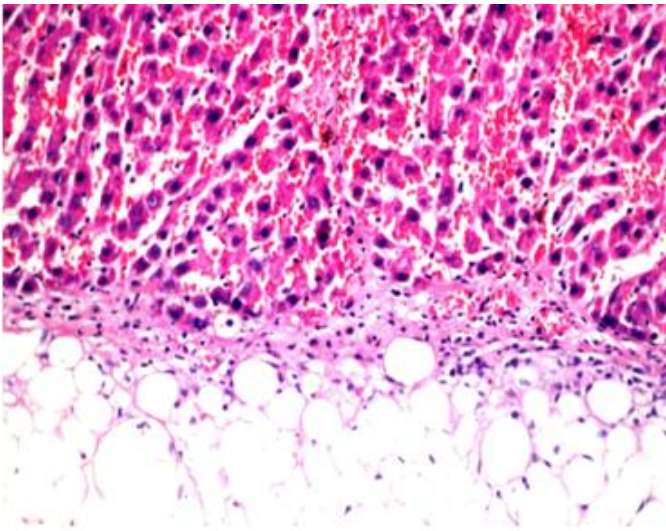
Figure 17: G (1): Control G (2): Probiotic G (3): CCL<sub>4</sub> G(4): Prbiotic+CCL<sub>4</sub>



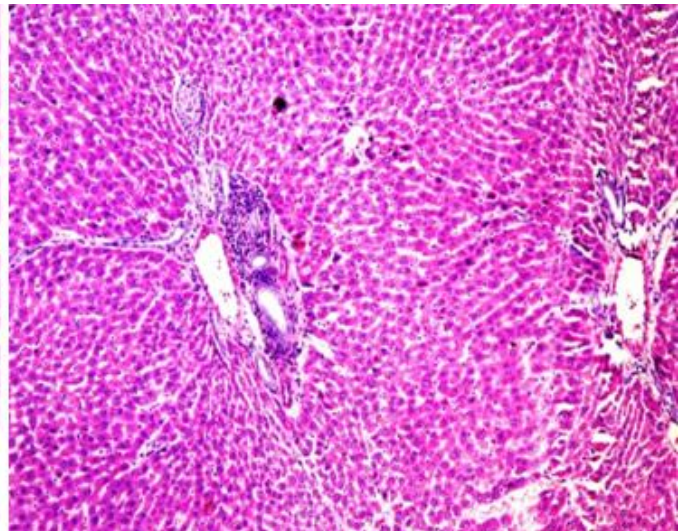
(A)



(B)

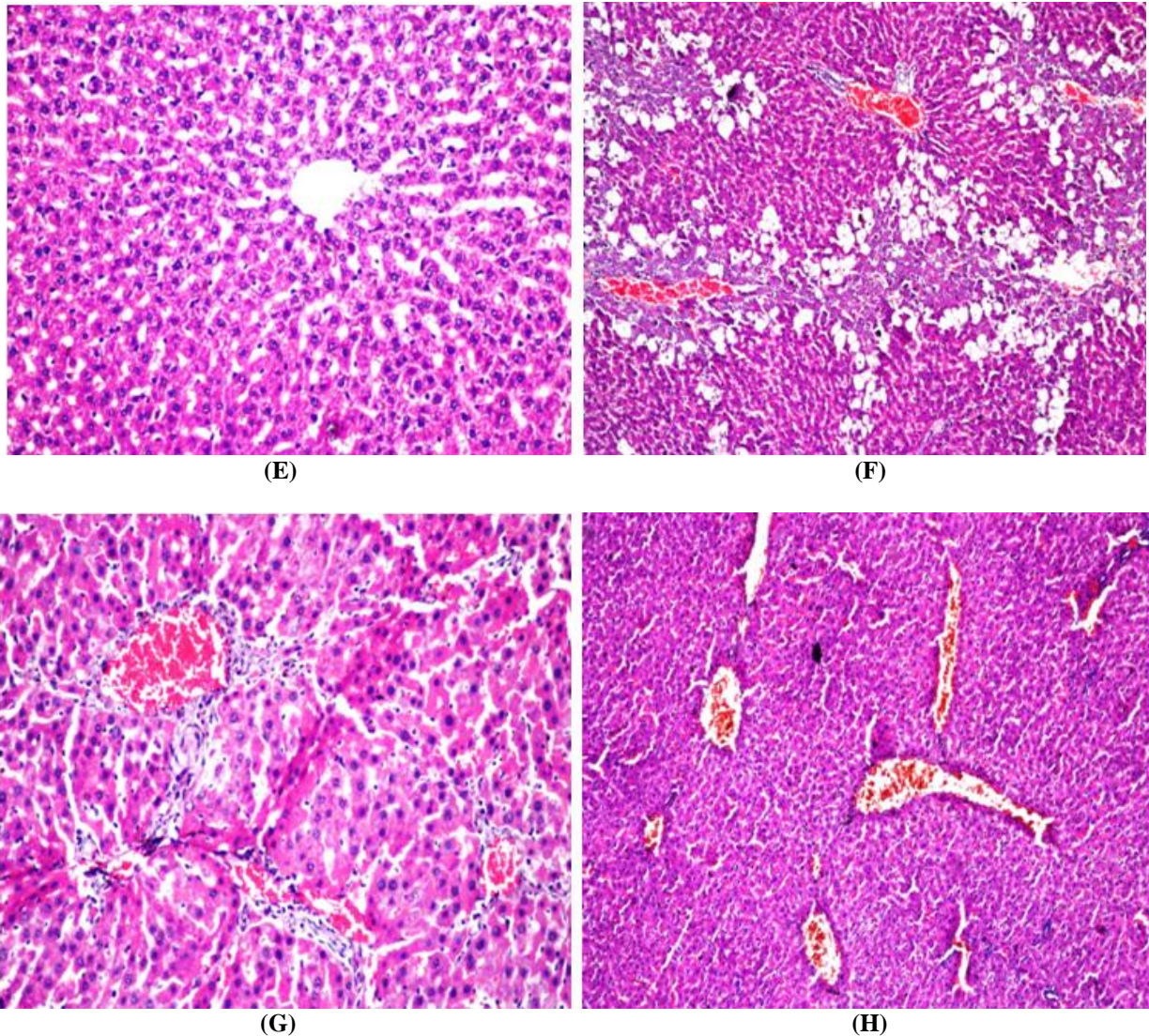


(C)

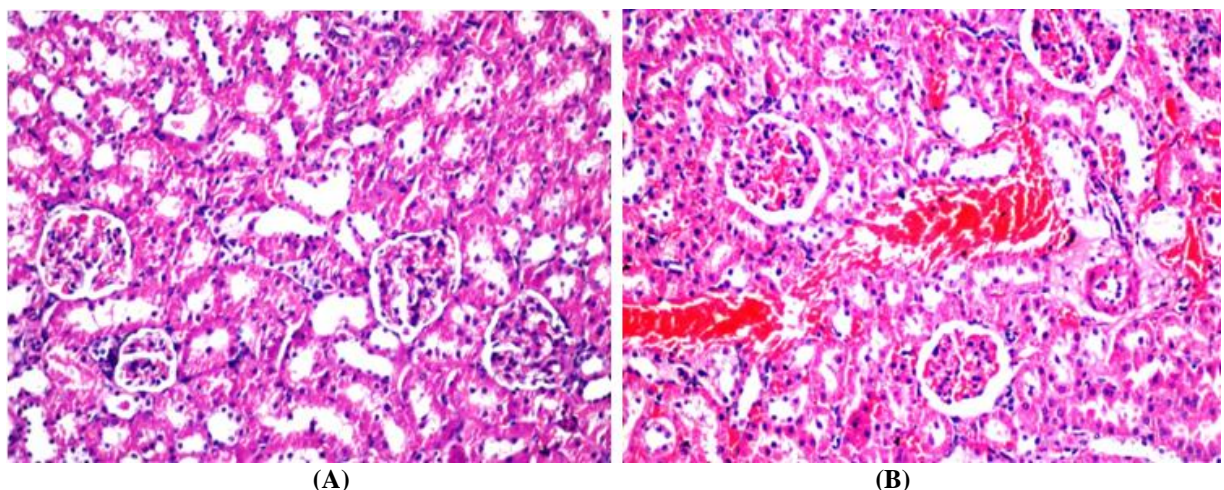


(D)

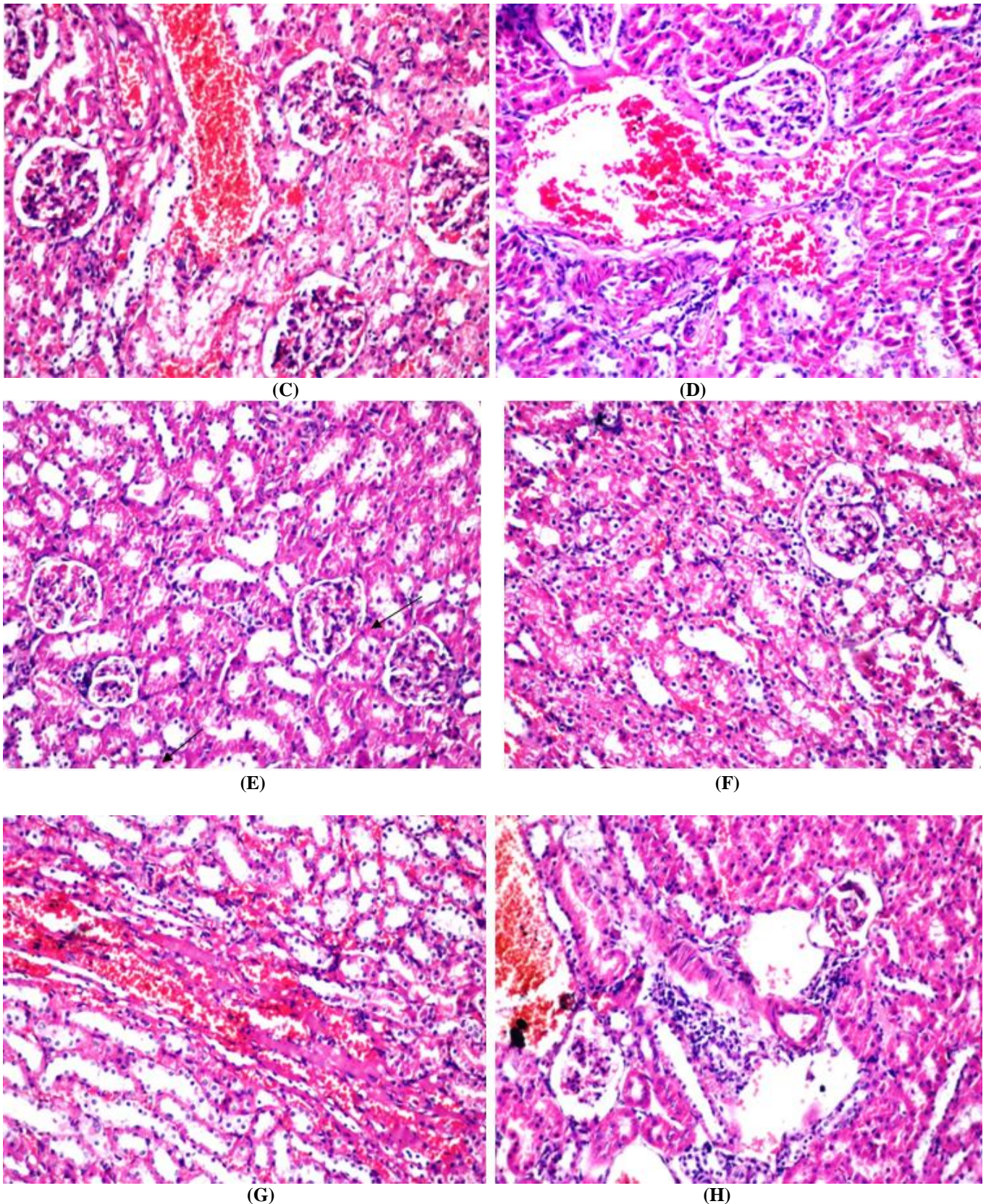




**Figure 18: Histopathological changes in liver sections of control normal ,carbon tetrachloride, probiotic and probiotic with carbon tetrachloride****A:**Liver sections from a control rat show normal histological structure of central vein and surrounding hepatocytes in the parenchyma **B,:** CCL4 rat shows steatosis with inflammatory cells infiltration in the hepatic capsule at first day. **C:**CCL4 rat shows steatosis with inflammatory cells infiltration in hepatic capsule with congestion in sinusoids of underlying parenchyma at seventh day. **D:** CCL4 rat shows massive inflammatory cells infiltration in the portal area with dilatation in portal vein at fourteenth day. **E:**probiotic shows normal histological structure of central vein and surrounding hepatocytes in the parenchyma **F:**probiotic-CCL4 rat shows centrilobular hepatic vacuolization and necrosis in the hepatocytes associated with congestion in the portal veins at first day **G:**probiotic-CCL4 rat shows fine fibrosis with inflammatory cells infiltration dividing the parenchyma into lobules associated with congestion in the portal vein at seventh day. **H:**probiotic-CCL4 rat shows congestion in the central and portal veins at fourteenth day.

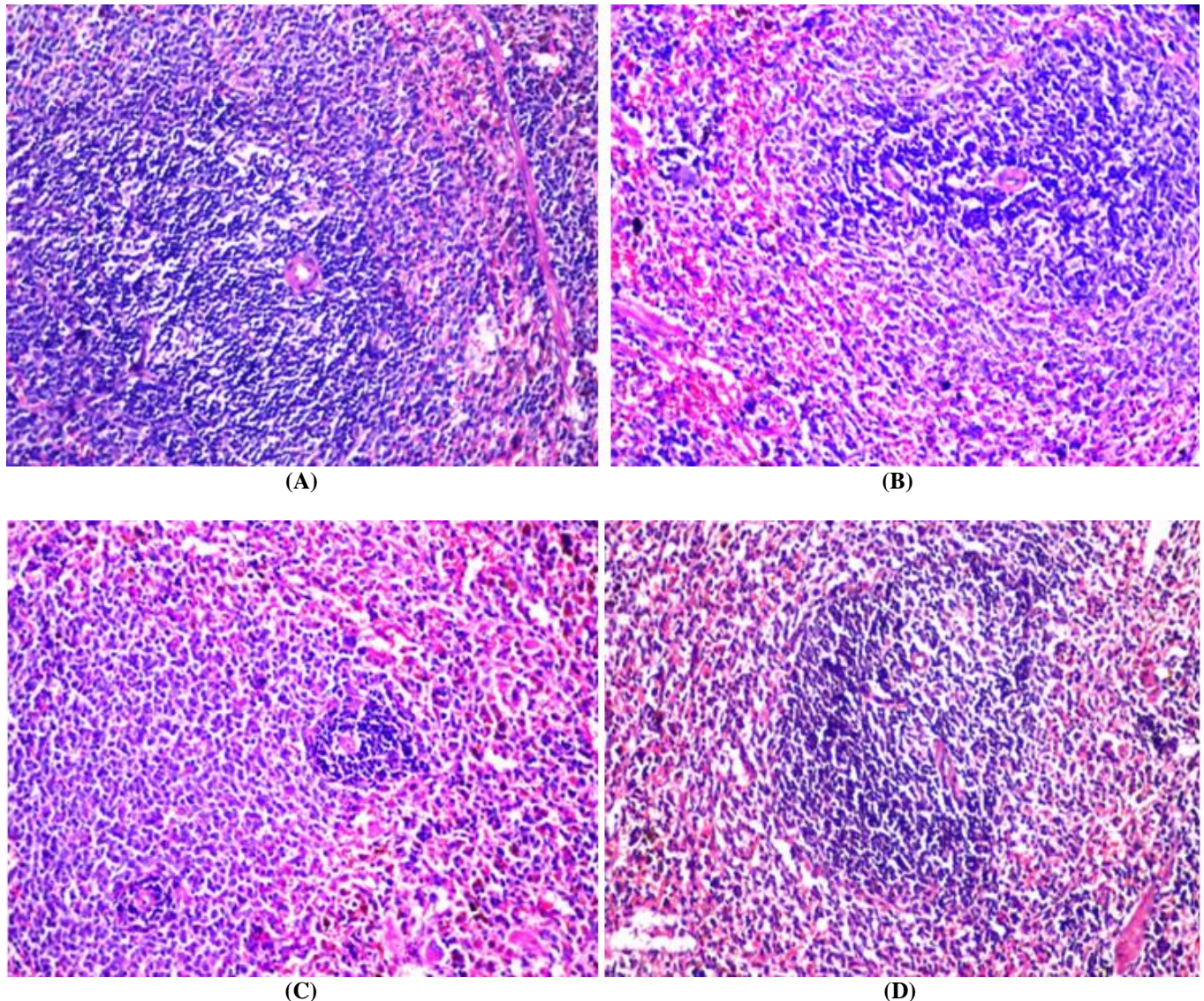






**Figure 19: Histopathological changes in kidney sections of control, carbon tetrachloride, probiotic and probiotic with carbon tetrachloride** **A:** Kidney sections from a control rat show normal histological structure of glomeruli and tubules. **B:** CCL4 rat shows congestion in cortical blood vessels at first day. **C:** CCL4 rat focal haemorrhagic in between the tubules at cortex at seventh day. **D:** CCL4 rat shows focal haemorrhagic in between the tubules at cortex at fourteenth day. **E:** probiotic normal histological structure of glomeruli and tubules **F:** probiotic-CCL4 rat shows swelling and degeneration in the tubular lining epithelium of the cortex at first day. **G:** probiotic-CCL4 rat shows focal haemorrhage between the tubules at the corticomedullary at seventh day **H:** probiotic-CCL4 rat shows focal inflammatory cells infiltration in between the tubules with congestion in the blood vessels at the cortex at fourteenth day.





**Figure 20: Histopathological changes in spleen sections of control, carbon tetrachloride, probiotic and probiotic with carbon tetrachloride** **A:** spleen sections from a control and probiotic rat show normal histological structure of the white and red pulps with surrounding sinusoids at first, seventh and fourteenth day. **B:** CCL4 rat shows lymphoid depletion in white pulps at first and seventh day. **C:** CCL4 rat shows severe depletion in lymphoid cells with pulps at fourteenth day **D:** probiotic-CCL4 rat shows lymphoid depletion in the white at first seventh and fourteenth day.

## 4. Results

### 4.1 Serum biochemical analysis

As shown in Figures (1-8)

These data suggested that when probiotic compared with control have insignificant change on AST, ALT, ALP, total bilirubin, urea and creatinine. CCL4 induced hepatotoxicity and nephrotoxicity as demonstrated by the elevation of serum liver and kidney biomarker. The AST, ALT, ALP, total bilirubin, urea and creatinine levels were substantially increased ( $p < 0.05$ ) in response to CCL4 treatment. CCL4 caused significant decrease total protein, albumin and creatinine kinase (Fig 13) compared to those of control rats. In contrast, these parameters were significantly reduced ( $p < 0.05$ ) when CCL4 treated-rats were administrated probiotic compared to the CCL4 group.

### 4.2. Lipid profile

As shown in Figures (9-10)

There was a significance decrease of probiotic group on serum cholesterol at first day but no remarkable change at seventh day and fourteenth day compared to control group (1). Probiotic followed by carbon tetrachloride group (7) had a significant decrease on serum cholesterol at first, seventh and fourteenth day compared to carbon tetrachloride toxicity. Probiotic caused remarkable decrease on serum triglyceride at first, seventh and fourteenth day compared to control group. There was a significant decrease of probiotic group followed by carbon tetrachloride on serum triglyceride at (1,7 and 14) day compared to carbon tetrachloride.



### 4.3 Hepatic oxidative damage parameter

Carbon tetrachloride intoxication and treatment with probiotic, on lipid peroxidation and liver oxidative parameters are shown in Figure 12. Carbon tetrachloride intoxicated rats showed significant increases ( $P \leq 0.05$ ) in MDA compared to those of control rats. However, the toxic effects of carbon tetrachloride on hepatic MDA were significantly ( $P \leq 0.05$ ) reduced by administration of probiotic.

### 4.4. Glucose level

As shown in Figure (11) carbon tetrachloride caused significant increase in glucose level, while probiotic reduced the elevation compared to Carbon tetrachloride.

### 4.5. Some hematological aspects (Hb, RBCs, WBCs and PCV)

As shown in Figures (14-17). The effects of carbon tetrachloride intoxication and treatment with probiotic, on hemoglobin concentration, RBCs, WBCs and PCV are shown in Figure (14-17). Carbon tetrachloride intoxicated rats showed significant decreases ( $P \leq 0.05$ ) in hemoglobin concentration and RBCs but there was increase in PCV compared to those of control rats. However, the toxic effects of CCL4 on Hb concentration, RBCs and PCV were significantly improved ( $P \leq 0.05$ ) by administration of probiotic.

### 4.6. Histopathological findings

As shown in Fig (18) Liver sections of control saline-treated rats had uniform polyhedral hepatocytes with normal sinusoids and central veins. In contrast, we observed portal vein congestion, severe hydropic degeneration, necrosis, nuclear condensation, and lymphocytic infiltration around the portal vein in CCL4-treated rats at first, seventh and fourteenth day after end of administration. Probiotic restored the normal hepatic architecture.

As shown in Fig. (19) Control rats had normal glomeruli and renal tubular epithelia. In contrast, CCL4-intoxicated rats showed a severe loss of the brush border, tubular necrosis, and tubular vacuolization. In addition, CCL4-treated rats exhibited moderate tubular dilatation and inflammatory cell infiltration at first, seventh and fourteenth day. Treatment with probiotic caused a notable recovery of the histopathological appearance after CCL4-induced renal injury.

As shown in Fig. (20) Spleen showed no histopathological alterations on control and probiotic group. Spleen of carbon tetrachloride showed severe depletion in lymphoid cells with pulps while probiotic followed carbon tetrachloride reduced lymphoid depletion in the white pulps.

## 5. Discussion

### 5.1 Effect on liver function:

Carbon tetrachloride induced hepato cellular damage as fatty degeneration, fibrosis and impairment of liver function [22-23-24 -25] and liver examination proved that

histopathological findings showed steatosis with inflammatory cells infiltration. The obtained results came in agreement with [26] proved that histopathological examination of livers reduced fatty degeneration, cytoplasmic vacuolization and necrosis in carbon tetrachloride treated rats. Carbon tetrachloride caused devaluation of liver tissues and diffuse hepatosteatosis in rats [27]. Histopathological injuries were recorded in liver of rat with carbon tetrachloride treatment [28].

Probiotic (*Lactobacillus acidophilus*) had no adverse effect compared to normal control rats. *Lactobacillus acidophilus* occurs naturally in the human and animal gastrointestinal tract and mouth [29-30] and histological examination proved that data [31] stated that no histopathological abnormalities or changes were observed in liver of both male and female rats of probiotic group, while probiotic and carbon tetrachloride induced marked reduction in liver function enzymes compared to hepatotoxicity of carbon tetrachloride. Our results came consistent with [32] evaluated that probiotics simultaneously with cadmium chloride restored the altered values of alanine aminotransferase and aspartate aminotransferase to the normal level in rat male Wistar rats *Rattus norvegicus*. [13] revealed that colitis caused significant ( $p < 0.05$ ) decrease in liver function enzymes. While *Lactobacillus acidophilus* recipient succeeded in keeping alkaline phosphatase and plasma total protein values within normal, but decreased alanine aminotransferase and aspartate aminotransferase in coparasm with control group. The *lactobacillus plantarum* CCFM8246 recovered the alanine aminotransferase and aspartate aminotransferase in serum against copper intoxication in mice [33]. [34] stated the probiotic *Clostridium butyricum* decrease the carbon tetrachloride induced levels of alanine aminotransferase and aspartate aminotransferase in the serum of these mice and liver examination showed significant improvement hepatocellular damage of probiotic and carbon tetrachloride. The obtained results came in agreement with [35] evaluated the efficiency of *Lactobacillus acidophilus* to protect liver from the toxicity of zearalenone in vital systems of albino white rats. The protective effects of probiotic consumed simultaneously with cadmium attenuated histomorphological changes in the liver [36]. Probiotics improved the histological feature of hepatocytes reduced apoptosis and stimulated proliferation on thioacetamide induced hepatotoxicity rats [37].

### 5.2 Effect on kidney function

Carbon tetrachloride caused nephrotoxicity which manifested by a significant increase serum level of creatinine and urea. Renal damage was represented by significant increase in serum urea and creatinine as recorded by [38]. Creatinine on the other hand is mostly derived from endogenous source by tissue creatinine breakdown. Altitude of urea and creatinine levels in the blood was taken as the index of nephrotoxicity [39]. Our findings are in accordance with [40-41-42-43-44-45] and kidney examination proved that histological findings of kidney showed renal degeneration, necrosis congestion in cortical blood vessels and focal haemorrhagic in between the tubules at cortex [46-41]



Probiotic did not have effect on serum level of creatinine and urea compared to control. This results came agreement with [47] suggested that the probiotic on blood urea nitrogen and creatinine concentrations no significance difference in large felids. Also [48] stated that *Lactobacillus plantarum* AD3 had no significant visible changes in respect to control rat, while probiotic and carbon tetrachloride induced a significant decrease on serum level of creatinine and urea compared to carbon tetrachloride. There is a direct correlation between the structure of microflora in gastrointestinal tract and uremic condition. Restructuring the gastrointestinal microbial community may be beneficial reducing uremic load in the gut and its escape in system circulation. Modification of intestinal flora to refrain generation of toxins by probiotic [49]. Probiotic evaluate the impact on solute concentration in plasma or on their fecal or urinary excretion [50]. It has been shown to have very potent antioxidant effects in preventing endothelial apoptosis caused by oxidants [51]. Our findings are in accordance with [52] investigated that probiotic strains supplemented to aflatoxins treated group revealed a significantly depletion in serum creatinine and urea levels. *Lactobacillus* played a protective role against the progression of chronic kidney disease [39]. *Lactobacillus plantarum* AD3 as an effective probiotic strain for acetaminophen induced uremic patient the level of plasma urea and creatinine in tested rats were significantly lower in comparison to uremic control [48] and kidney examination of probiotic and carbon tetrachloride showed good improvement of the histological picture. There were focal degenerative changes in proximal tubules without vacuoles and apoptosis, while glomeruli were not affected. These findings are in agreement with [39] investigated the protective effect of probiotic supplementation against cadmium-induced toxicity in rat male Wistar rats *Rattus norvegicus* by histopathological changes in kidneys.

### 5.3 Effect on lipid profile

Carbon tetrachloride induced a significant increase on lipid profile (cholesterol and triglyceride) compared to control. Rise in serum total cholesterol might be attributed to its reduced catabolic rate and/or reduced activity of hepatic cholesterol-7-alpha-hydroxylase, the rate limiting enzyme in bile acid synthesis from cholesterol [53-54-55-56].

Probiotic caused a significant decrease on lipid profile (cholesterol and triglyceride) compared to control. The obtained results came in agreement with that obtained by [57-58] showed that the hypocholesterolemic effect of local *Lactobacillus* strains was attributed to its ability to lower serum and liver total cholesterol levels. *Lactobacillus acidophilus* significantly reduced in cholesterol and triglycerides in rats [59-60-61-62].

Probiotic and carbon tetrachloride induced a significant decrease on serum level of cholesterol and triglyceride compared to carbon tetrachloride rats. The obtained results came consistent with that obtained by [63] concluded that *Lactobacillus acidophilus* supplementation exert hypotriglyceridemic in diabetic rats. *Lactobacillus acidophilus* La5 and *Bifidobacterium lactis* improved total cholesterol and low density lipoprotein cholesterol

concentrations in type 2 diabetic people [64]. *Lactobacillus casei* 01 improved the total cholesterol, low-density lipoprotein-cholesterol, high-density lipoprotein-cholesterol and triacylglycerides of the hypertensive overweighted women [65].

### 5.4 Effect on serum glucose level:

Carbon tetrachloride showed a significant increase in serum glucose level compared to control rats [66-67]. Probiotic had insignificant difference on glucose level compared to control. The obtained results came consistent with [68] found that probiotic treatment had no effect on blood glucose levels in healthy rats and [69] said that *Lactobacillus casei* did not cause a significant change in blood glucose levels in control mice receiving the probiotic.

The effect of probiotic and carbon tetrachloride restored the increase of glucose of carbon tetrachloride. The obtained results came in agreement with that attained by [70] stated that *Lactobacillus acidophilus* had the effect of the decreasing blood glucose levels and in alloxan-induced diabetic rats. [71] suggested that probiotic consumption improved the glycemic control in Type 2 diabetes.

### 5.5 Effect on serum L-malondialdehyde (MDA) level:

Carbon tetrachloride caused a significant increase in serum L-malondialdehyde in rats compared to control [43-72-73]. Probiotic showed insignificant change on malondialdehyde level. The obtained results came in agreement with [74] said that *Lactobacillus rhamnosus* feeding to mice ( $10^7$ ,  $10^9$ ,  $10^{11}$  and  $10^{13}$  cfu/animal/d) repetitively for 28 days revealed no adverse effects on malondialdehyde in rat. Probiotic and carbon tetrachloride restored the increase malondialdehyde level induced by carbon tetrachloride. The obtained results came in agreement with that attained by [63] concluded that *Lactobacillus acidophilus* significantly decrease the elevated malondialdehyde in diabetic rats. *Lactobacillus plantarum* CCFM8661 offered a significant protective effect against lead toxicity by preventing alterations in the levels of malondialdehyde caused by lead exposure [75]. Inconsistent with [76] evaluated the effect of probiotics indicated a non-significant declining trend in the level of malondialdehyde.

### 5.6 Effect on serum creatinine kinase level:

Carbon tetrachloride induced a significant decrease on creatinine kinase level compared to control [77-78]. Probiotic and carbon tetrachloride caused significant increase on creatinine kinase level compared to carbon tetrachloride group. The findings agreement with [79] revealed that *Lactobacillus plantarum* TWK10 (LP10) supplementation increase creatine kinase of exercise performance, physical fatigue, and gut microbial profile mice. Long-term supplementation with probiotic may increase muscle mass, enhance energy harvesting, and have health-promotion, performance-improvement, and anti-fatigue effects.

### 5.7 Effect on hematological picture:

Carbon tetrachloride induced hematological toxicity in rat which manifested by significant increase on white blood



cells and packed cell volume but significantly decreased in red blood cell and hemoglobin. This result was similar to that obtained by [80] illustrated that carbon tetrachloride caused significant reduction in hemoglobin content and haematocrit percentage accompanied by leukocytosis, granulocytosis, monocytosis and lymphocytopenia. Probiotic had no remarkable change on hemoglobin, red blood cell, white blood cell packed cell volume compared to normal control and the results were consistent with [81] stated that *Lactobacillus plantarum* and *Lactobacillus casei* or their mixed culture improved health performance of rats in terms of hematological and these increases were only of statistical significance without any clinical relevance as most of these parameters were within the normal physiological values for rats. Inconsistent with [82] investigated that the effect of oral administration of different doses of probiotic, *Lactobacillus plantarum*. There was a significant increase in hemoglobin and red blood cell when compared with the control in rat

Probiotic and carbon tetrachloride induced improvement on level of hemoglobin, and red blood cell count but there was no effect on white blood cells and packed cell volume compared to carbon tetrachloride. The results were agreement with [35] evaluated that *Lactobacillus acidophilus* had the ability to raise hemoglobin treating rats with zearalenone.

## 6. Conclusion

Available evidence indicates that probiotics (*Lactobacillus acidophilus*) supplements is not toxic and have benefit effect on lipid profile (cholesterol and triglyceride) of healthy rat. Probiotics (*Lactobacillus acidophilus*) may play a vital role in management toxicity of carbon tetrachloride by alleviating the carbon tetrachloride toxicity-induced on liver and kidney damage with improvement on hematological, lipid profile and glucose level.

## References

- [1] Demirenen, K.; Dogan, Y.; Kocamaz, H.; Ozercan, I.H.; Ilhan, S.; Ustundag, B. "Protective effects of L-carnitine, N-acetylcysteine and genistein in an experimental model of liver fibrosis". *Clin Res Hepatol Gastroenterol*. 2014; **38**(1): 63-72
- [2] Sheweita, S.A.; El-Gabar, M.A. and Bastawy M. "Carbon tetrachloride changes the activity of cytochrome P450 system in the liver of male rats: role of antioxidants". *Toxicology*. 2001; **169**(2): 83-92.
- [3] Fujii T. "Toxicological correlation between changes in blood biochemical parameters and liver histopathological findings". *Journal of Toxicological Science*; 1997. 22: 161-83.
- [4] Sanmugopriya, E. and Venkataraman, S. "Studies on hepatoprotective and antioxidant actions of *Strychnos potatorum* Linn. seeds on CCl<sub>4</sub>-induced acute hepatic injury in experimental rats". *J. Ethnopharmacol*. 2006, 105: 154-160.
- [5] Shukla, A. and Bhatia, S.J. "Outcome of patients with primary hepatic venous obstruction treated with anticoagulants alone". *Indian J Gastroenterol*. 2010, 29(1): 14-17.
- [6] Fuller, R. "Probiotic in man and animal". *J Appl Bacteriol* 1989, 66(5): 365-378.
- [7] Sobko, T.; Norman, M.; Noria, E.; Gustafsson, L.E.; Lundberg, J.O. "Birth-related increase in intracolonic hydrogen gas and nitric oxide as indicator of host microbial interactions". *Allergy* 2005 60: 396-400.
- [8] Ljungh, A. and Wadstrom, T. "Lactic acid bacteria as probiotics". *Curr Issues Intest Microbiol*, 2006, 7: pp. 73-89.
- [9] Hasen, P.A. and Mocquot, G. "Lactobacillus acidophilus (Moro) comb. nov". *Int. J. Syst. Bacteriol*. 1970, 20: 325-327.
- [10] Duggan, C.; Gannon, J. and Walker, W.A. "Protective nutrients and functional foods for the gastrointestinal tract". *Am. J. Clin. Nutr* 2002, 75, 789-808.
- [11] Bengmark, S. "Biological control of the gastrointestinal tract: the role of flora and supplemented probiotics and synbiotics". *Gastroenterol Clin North Am*. 2005, 34: 413-436.
- [12] Madsen, K.L.; Doyle, J.S.; Jewell, L.D.; Tavernini, M.M. and Fedorak, R.N. "Lactobacillus species prevents colitis in interleukin 10 gene-deficient mice". *Gastroenterology* 1999, 116: 1107-1114.
- [13] Alqayim, M.A.J. and Abass, D.E. "Effects of probiotics (*Lactobacillus acidophilus*) on liver functions in experimental colitis in rats". *Iraqi J. Vet. Med*. 2014, 38(2): 48-58.
- [14] Vani, M.; Prakash, M.S. and Devi, P.Y. "Hypocholesterolemic effects of *Lactobacillus acidophilus* as a dietary supplement". *Indian Journal of Clinical Practice*, 2012, Vol. 23, No. 4.
- [15] Kang, S.S.; Byeon, H.S.; and Kim, J.T. (2011): Effects of *Lactobacillus acidophilus* on innate immunity. *Korean J. Vet. Serv* 34(3): 235-243.
- [16] Sadeghzadeh, J.; Vakili, A.; Sameni, H.R.; Shadnough, M.; Bandegi, A.R. and Khorasani, M.Z. "The effect of oral consumption of probiotics in prevention of heart injury in a rat myocardial infarction model: A histopathological, hemodynamic and biochemical evaluation". *Iran. Biomed*. 2017, J. 21, 174-181.
- [17] Velayudham, A.; Dolganiuc, A.; Ellis, M.; Petrasek, J.; Kodys, K.; Mandrekar, P. and Szabo, G. "VSL#3 probiotic treatment attenuates fibrosis without changes in steatohepatitis in a diet-induced nonalcoholic steatohepatitis model in mice". *Hepatology*, 2009, 49(3): 989-97.
- [18] Bancroft, J.D.; Stevens, A. and Turner, D.R. "Theory and practice of histological techniques Fourth ED". Churchill Livingstone, New York, London, San Francisco, Tokyo. 1996.
- [19] SPSS Inc. Released "PASW Statistics for Windows, Version 18.0. Chicago: SPSS Inc. SPSS Inc. Released (2009): PASW Statistics for Windows, Version 18.0. Chicago: SPSS Inc.
- [20] Snedecor, G.W. and Cochran, W.C. "Statistical methods". The eighth. Edition, Iowa University Press, Ames, Iowa, USA. 1989
- [21] Duncan, D.B. "Multiple range and multiple F tests". *Biometrics* 11: 142. 1955
- [22] Gnanaprakash, K.; Madhusudhana, C.C.; Ramkanth, S.; Alagusundaram, M.; Tiruvengadarajan,



- V.S.;Angala, P.S and Saleem, M. "Aqueous extract of *Flacourtiaindica* prevents carbon tetrachloride induced hepatotoxicity in rat". Int J Biol Life Sci.;6:51–55.2010
- [23] **Bolanle, J.D; Adetoro,K.O; Balarabe,S.A and Adeyemi,O.O.**Hepatocurative potential of *Vitexdoniana* root bark, stem bark and leaves extracts against CCl<sub>4</sub>-induced liver damage in rats.Asian Pac J Trop Biomed. Jun; 4(6): 480–485.2014.
- [24] **Batool, R; Khan, M.R.andMajid,M**"Euphorbia dracunculoides L. abrogates carbon tetrachloride induced liver and DNA damage in rats".BMC Complement Altern Med. Apr 20;17(1):223.2017
- [25] **Otrubová, O; Turecký, L; Uličná, O; Janega, P; Luha, J, and Muchová J.**"Therapeutic effects of N-acetyl-L-cysteine on liver damage induced by long-term CCl<sub>4</sub> administration".Gen PhysiolBiophys.Jan;37(1):23-31
- [26] **Deng, J.S; Chang, Y.C; Wen, C.L; Liao, J.C; Hou, W.C; Amagaya, S; Huang, S.S and Huang, G.J.**"Hepatoprotective effect of the ethanol extract of *Vitisthunbergii* on carbon tetrachloride-induced acute hepatotoxicity in rats through anti-oxidative activities".Journal Ethnopharmacol. Aug 1;142(3):795-803.2012
- [27] **Cetinkaya, A; Kantarceken, B; Bulbuloglu, E; Kurutas, E.B; Ciralik, H and Atli, Y**"The effects of L-carnitine and N-acetylcysteine on carbontetrachloride induced acute liver damage in rats".BratisLek Listy.;114(12):682-8.2013
- [28] **Laouar,A;Fahima,K;Bourogaa,E and AmelBenamara,A.** Potential antioxidant properties and hepatoprotective effects of *Juniperusphoenicea* berries against CCl<sub>4</sub> induced hepatic damage in rats.Asian Pacific Journal of Tropical Medicine 10(3).2017
- [29] **Asemi, Z and Esmailzadeh,A**"Effect of daily consumption of probiotic yoghurt on serum levels of calcium, iron and liver enzymes in pregnant women".Int J Prev Med. 4(8): 949–955.2013
- [30] **Eze,J.I and Ubochioma,E.D.**"Effect of dietary probiotics (*Saccharomyces cerevisiae*) supplementation on serum biochemicals of *Trypanosomabrucei* infected rats". Current Bioactive Compounds. 14(3).2018
- [31] **Skrypnik,K; Bogdański,P; Loniewski,I ;Regula,J andSuliburska,J.** "Effect of probiotic supplementation on liver function and lipid status in rats".ActaSci Pol Technol Aliment Apr-Jun;17(2):185-192.2018
- [32] **Jama,A.M.;Đurasevic, S. andJasnic, N.** "Protective effect of probiotic in the model of cadmium toxicity in rats". Archives of Biological Sciences 64(3):1197-1206.2013
- [33] **Tian, F.W.; Xiao, Y.; Li, X.X.;Zhai, Q.X.; Wang, G.; Zhang, Q.X.; Zhang, H . andChen, W.** "Protective effects of *Lactobacillus plantarum* CCFM8246 against copper Toxicity in Mice" .PLoSOne.10(11):e0143318. 2015
- [34] **Majlesi, M.;Shekarforoush, S.S.; Ghaisari, H.R.;Nazifi, S.;Sajedianfard, J. andEskandari, M.H.** "Effect of probiotic *Bacillus coagulans* and *Lactobacillus Plantarum* on alleviation of mercury toxicity in rat" .Probiotics and Antimicrobial Proteins 9(3).2017
- [35] **Ali, S.A.; Ibrahim, H.J.; Ali, M.N.; Ali, K.M.;Shahad, H.H.;Rusul, N.K.;Zahraa, J.R. and Meyameen, H.M.**"Bioremediation of Zearalenone by using *Lactobacillus acidophilus* in albino rats bodies (in vivo)".J Cont Med Sci| Vol. 1, No. 1.;21–25.2015
- [36] **Djurasevic,S.;Jama, A.;Jasnic N.;Vujovic, P.;Jovanovic, M.;Mitic-Culafic, D.;Knezevic-Vukcevic, J.; Cakic-Milosevic, M.;Ilijevic, K andDjordjevic, J**"The Protective Effects of Probiotic Bacteria on Cadmium Toxicity in Rats". J Med Food. Feb;20(2):189-196.2017
- [37] **Emam, M. A.; Farouk, S. M. and Abdo, M.** "The ameliorative potential of probiotics and/or silymarin on thioacetamide induced hepatotoxicity in rats: histological and immunohistochemical study". Int. J. Morphol., 36(2):661-669.2018.
- [38] **Kaneko JJ.** *Clinical biochemistry of domestic animals*. 5th ed. San Diego (CA): AcademicPress, 1997.
- [39] **Alatriste,P.M; Arronte,R.U; Espinosa,C.G and Cuevas,M.D.**Effect of probiotics on human blood urea levels in patients with chronic renal failure. Nutr Hosp.;29(3):582-590.2014
- [40] **Olagunju, J.A;Adeneye, A.A;Fagbohunka, B.S;Bisuga, N.A;Ketiku, A.O;Benebo, A.S;Olufowobi, O.M;Adeoye, A.G;Alimi, M.A andAdeleke, A.G.** "Nephroprotective activities of the aqueous seed extract of *Carica papaya* Linn. In carbon tetrachloride induced renal injured Wistar rats: a dose- and time-dependent study". Biol Med, 1: 11-19.2009.
- [41] **Venkatanarayana, G;Sudhakara, G;Sivajyothi, P and Indira, P.** Protective effects of curcumin and vitamin E on carbon tetrachloride-induced nephrotoxicity in rats. EXCLI J, 11: 641-650.2012.
- [42] **Yacout,G.A; Elguindy,N.M and El Azab,E.F.**"Hepatoprotective effect of basil (*OcimumBasilicum* L.) on CCl<sub>4</sub>-induced liver fibrosis in rats.Afr J Biotechnol;11(90):15702–15711.2012
- [43] **Haghi,M.Es;Dehghan,G;Banihabib,N;Zare,S;Mikail i,Pand Panahi,F** "Protective effects of *Cornus mas* fruit extract on carbon tetrachloride induced nephrotoxicity in rats".Indian J Nephrol. Sep-Oct; 24(5): 291–296.2014
- [44] **Kamal, S.** "The protective effect of curcumin on nephrotoxicity induced by carbon tetrachloride in rats". Journal of advances in life and Natural sciences, Vol. 1 (1), 22-29,2015
- [45] **Mazani,M; Mahmoodzadeh,Y; Chinifroush-Asl,M.M; Banaei,S; Rezagholizadeh,L; Mohammadnia,A.**"Renoprotective effects of the methanolic extract of *Tanacetumparthenium* against carbon tetrachloride-induced renal injury in rats". AvicennaJournalPhytomedicien, Vol. 8, No. 4, Jul-Aug. 2018
- [46] **Dogukan, A; Celiker,H;Akpolat,N and Ilhan,N.(2003):** Protective effect of interferon on carbon tetrachloride-induced nephrotoxicity Journal of nephrology 16(1):81-4
- [47] **McCain ,S; Schumacher ,J; Allender ,M.C and Ramsay ,E**"The effects of a probiotic on blood urea nitrogen and creatinine concentrations in large felids". Journal of Zoo and Wildlife Medicine 42(3):426-9.2011
- [48] **Patra,A; Mandal,S; Samanta,A;Mondal,K; and Nandi, D** "Therapeutic potential of probiotic

- Lactobacillus Plantarum AD3 on acetaminophen induced uremia in experimental rats". Clin Nutr Exp Volume 19, Pages 12-22.2018
- [49] **Mandal, a; Roy, S; Das, K; Mondal, K and Nandi, D.K.** "In vivo assessment of bacteriotherapy on acetaminophen-induced uremic rats". J Nephrol, 26pp 228-236.2013
- [50] **Chow, K.M; Kiu, Z.C; Prakash, S and Chang, T.M** "Free and microencapsulated Lactobacillus and effects of induction on urea removal. Artif Blood Substit. Immobil Biotechnol, 31, pp 425-434.2003
- [51] **Di Cerbo, A; Pezzuto, F; Pamieri, L; Rottigni, V; Lannitti, T and Pamieri, B** "Clinical and experimental use of probiotic formulation for management of end stage renal disease: an update". Int Urol Nephrol, 45, pp. 1569-1576.2010
- [52] **Salim, A. B.; Zohair, A.; Hegazy, A. E. S. and Said, A.** "Effect of some Strains of Probiotic Bacteria against Toxicity Induced by Aflatoxins in vivo". J. Am. Sci., 7(1). 2011.
- [53] **Marimuthu S; Adluri RS; Rajagopalan R; Menon VP.** "Protective role of ferulic acid on carbon tetrachloride induced hyperlipidemia and histological experimental rats". J Basic Clin Physiol Pharmacol. 24(1):59-66.2013
- [54] **Agatemor, U.M; Nwodo, O.F.C; Adejoh, I.P and Anosike, C.A.** "Protective effect of cucumissativus on carbon tetrachloride CCl<sub>4</sub>-induced liver damage in rats. Asian Journal of Biochemistry, 13: 22-29.2018.
- [55] **Abdul-Lattif, R.F** "Effect of sesame oil on lipid profile and liver enzymes in male albino rats treated with carbene tetrachloride (CCl<sub>4</sub>)". Ibn Al-Haitham Journal for Pure and Applied science. IHSCICONF. 1769.2018
- [56] **Shahwan, M and Zain Al Abdin, S.M** "Antioxidant, hepatoprotective and lipid lowering activity of Sarcopoterium Spinosum on carbon tetrachloride (CCl<sub>4</sub>)- induced hepatic damage in rats". J. Pharm. Sci. & Res. Vol. 10(11), 2800-2804.2018
- [57] **Chiu, C.H.; Lu, T.Y.; Tseng, Y.Y. and Pan, T.M.** "The effects of Lactobacillus-fermented milk on lipid metabolism in hamsters fed on high-cholesterol diet". Appl Microbiol Biotechnol. Jun; 71(2): 238-45.2006
- [58] **Kapila, S.; Vibha, and Sinha, P.R. (2006):** Antioxidative and hypocholesterolemic effect of *Lactobacillus casei* sp. *casei* (biodefensive properties of lactobacilli). Indian J Med Sci. Sep; 60(9): 361-70.
- [59] **Huang, Y.; Wang, J.; Cheng, Y. and Zheng, Y.** "The hypocholesterolaemic effects of Lactobacillus acidophilus American type culture collection 4356 in rats are mediated by the down-regulation of Niemann-Pick C1-like 1". Br J Nutr. Sep; 104(6): 807-12.2010.
- [60] **Vani, M.; Prakash, M.S. and Devi, P.Y.** "Hypocholesterolemic effects of Lactobacillus acidophilus as a dietary supplement". Indian Journal of Clinical Practice, Vol. 23, No. 4.2012
- [61] **Shrivastava, A.; Chaturvedi, U. and Bhatia, G.** "Hypolipidemic and antioxidant effect of *Lactobacillus acidophilus* bacteria in hyperlipidemic rats". Asian Journal of Pharmaceutical and Clinical Research 6(2): 84-87.2013
- [62] **Jabbar, M.K.** "The Effect of probiotic (*Lactobacillus acidophilus*) on the absorption of calcium and cholesterol in the intestines of rats". International Journal of ChemTech Research Vol. 10 No. 6, pp 20-24,
- [63] **Harisa, G.I.; Taha, E.I.; Khalil, A.F. and Msalem, M.** "Oral administration of *Lactobacillus Acidophilus* restores nitric oxide level in diabetic rats". Australian Journal of Basic and Applied Sciences, 3(3): 2963-2969, ISSN 1991-8178.2009
- [64] **Ejtahed, H.S.; Mohtadi-Nia, J.; Homayouni-Rad, A.; Niafar, M.; Asghari-Jafarabadi, M.; Mofid, V and Akbarian-Moghari, A** "Effect of probiotic yogurt containing *Lactobacillus acidophilus* and *Bifidobacterium lactis* on lipid profile in individuals with type 2 diabetes mellitus". J Dairy Sci ; 94: 3288-3294.2011
- [65] **Sperry, M.F.; Silva, H.L.; Balthazar, C.F.; Esmerino, E.A.; Verruck, S. and Prudencio, E.S** "Probiotic Minas Frescal cheese added with L. casei 01: Physicochemical and bioactivity characterization and effects on hematological/biochemical parameters of hypertensive overweighted women-A randomized double-blind pilot trial". Journal of Functional Foods, 45, pp. 435-443.2018.
- [66] **Adeneye, A.A; Awodele, O; Aiyeola, S.A and Benebo, A.S.** "Modulatory potentials of the aqueous stem bark extract of *Mangifera indica* on carbon tetrachloride-induced hepatotoxicity in rats. Journal of Traditional and Complementary Medicine. 5: 106-115.2015
- [67] **Ozturk, M; Akdogan, M; Keskin, I; Kisioglu, A.N; Oztas, S and Yildiz, K.** "Effect of silybummarianum on acute hepatic damage caused by carbon tetrachloride in rats". Biomedical research Volume 23, Issue 2.2012
- [68] **Al-Salami, H.; Butt, G.; Fawcett, J.P.; Tucker, I.G.; Golocorbin-Kon, S. and Mikov** "Probiotic treatment reduces blood glucose levels and increases systemic absorption of gliclazide in diabetic rats". Eur J Drug Metab Pharmacokinet. Apr-Jun; 33(2): 101-6.2008
- [69] **Asgharzadeh, F.; Tanomand, A.; Ashoori, M.R.; Asgharzadeh, A. and Zarghami, N.** "Investigating the effects of *Lactobacillus casei* on some biochemical parameters in diabetic mice". Journal of Endocrinology, Metabolism and Diabetes of South Africa; 22(3): 47-50.2017
- [70] **Nawangsih, E N.; Paryati, S Y.; Yoga, L. B. and Yuslianti, E. R.** "Effect of Munghurt *Lactobacillus acidophilus* from Green Beans to Blood Glucose Levels in Alloxan-induced Diabetic Rats". Reaearch journal of medicinal plants. Volume: 11 | Issue: 2, P; 41-47.2017
- [71] **Tonucci, L.B.; Olbrich Dos Santos, K.M.; Licursi de Oliveira, L.; Rocha Ribeiro, S.M. and Duarte Martino, H.S** "Clinical application of probiotics in type 2 diabetes mellitus: A randomized, double-blind, placebo-controlled study". Clin Nutr. Feb; 36(1): 85-92.2015
- [72] **Shen B, Chen H, Shen C, Xu P, Li J, Shen, G; Yuan, H and Han, J.** "Hepatoprotective effects of lignans extract from *Herpetospermum caudigerum* against CCl<sub>4</sub>-induced acute liver injury in mice". J Ethnopharmacol, 164: 46-52.2015
- [73] **Nie, Y; Ren, D; Lu, X; Sun, Y and Yang, X.** "Differential protective effects of polyphenol extracts from apple peels and flesh against acute CCl<sub>4</sub>-induced liver damage in mice". Food Funct. Feb; 6(2): 513-24.2015



- [74] **Bhat, M.I;Singh, V.K;Sharma, D;Kapila, S;Kapila, R.**“Adherence capability and safety assessment of an indigenous probiotic strain *Lactobacillus rhamnosus* MTCC-5897”.*MicrobPathog.* 2019 May;130:120-130.
- [75] **Tian, F;Zha,O; Zhao, J and Liu, X.(2012):** *Lactobacillus plantarum* CCFM8661 Alleviates Lead Toxicity in Mice.*Biological trace element research* 150(1-3)
- [76] **Mazloom,Z.;Yousefinejad, A. andDabbaghmanesh, M.H**“Effect of probiotics on lipid profile, glycemic control, insulin action, oxidative stress, and inflammatory markers in patients with type 2 diabetes: a clinical trial”. *Iran J Med Sci.* Mar;38(1):38-43.2013.
- [77] **Pacheco,G.S; Panatto, J.P.** “Brain creatine kinase activity is inhibited after hepatic failure induced by carbon tetrachloride or acetaminophen” .*Metabolic Brain disease* 24(3):383-94.2009
- [78] **Gabardo, T and Funchal, C.** “Assessment of changes in energy metabolism parametrs provoked by carbon tetrachloride in Wistar rats and the protective effect of white juice.*Toxicology Volume 2* Pages 645-653.2015
- [79] **Huang, Y.;Wang, X.;Wang, J.;Wu, F.;Sui, Y.;Yang, L. andWang, Z.**“*Lactobacillus plantarum* strains as potential probiotic cultures with cholesterol-lowering activity”. *Journal of Dairy Science* 96(5).2018
- [80] **El Bialy, B.E; El-Boraey, N.G; Hamouda, R.A and Abdel-Daim,M.M.**“Comparative protective effects of *Spirulina* and *Spirulina* supplemented with thiamine against sub-acute carbon tetrachloride toxicity in rats”. *Biomedical & Pharmacology Journal*, June Vol. 12(2), p. 511-525.2019
- [81] **Singh, P.;Pandey, R.K.; Paswan, V.K.;Yadav, S.P.;Bhinchhar, B.K. and Singh, C.S.**“Effect of supplementation of *L. plantarum* and *L. casei* based probiotic milk powder on hematology, blood biochemistry and lipid profile of Charles Foster rats.*Indian J. Animal Research.*2018.
- [82] **Aboderin,F.I. andOyetayo, V.O**“Haematological studies of rats fed different doses of probiotic, *Lactobacillus plantarum*, isolated from fermenting corn slurry”. *Volume: 5, Issue: 2, Page 102-105.*2006