International Journal of Science and Research (IJSR)

ISSN: 2319-7064

ResearchGate Impact Factor (2018): 0.28 | SJIF (2018): 7.426

Contact Action of Spinosad on *Callosobruchus chinensis* (L.) in Three Successive Generations

Tahera Tanjin Naher¹, W Islam²

Institute of Biological Sciences, University of Rajshahi, Bangladesh

Abstract: Stored product insect pest's causes substantial economic and quality losses to stored pulses or cereals. Callosobruchus chinensis (L.) is a major insect pest of stored pulses. The beetle infested lentil (Lens esculentus L.) seeds in stored conditions and causes severe damages. Indiscriminate uses of insecticides or fumigants to control the beetle can cause health hazards to human being. This study explores development technology for use in control of C. chinensis which is eco-friendly and biodegradable. Spinosad influenced the progeny production, fertility, adult mortality and sex-ratio of C. chinensis in three successive generations. In the first generation, the concentration was increased (6.2×10⁻⁵ppm/µml), the progeny production rate was decreased (male 32.67 and female30.33). In the second and third generations the same was gradually decreased.

Keywords: Contact action, spinosad, Callosobruchus chinensis

1. Introduction

Stored pulses are primarily attacks by major insect pest particularly the pulse beetle, Callosobruchus chinensis. chemicals provide effective Callosobruchus sp., but the efficiency of the chemicals mostly depends on the mode of exposure. The maximum effectiveness is given when the insecticides are provided by gustatory method, but majority of these are toxic to human (Bekele et al. 1995). Under such circumstances, alternative methods are needed to be considered (Isman 2008). The alternative solution currently recommended, is the judicious exploitation of our phylogenetic resources and microbial organisms. These two methods are often termed as biological control. Within the framework of the biological fight against such insect attacks, several works have been done by number of researchers (Thiam and Ducommun 1993, Isman 2006, Chiasson et al. 2008, Regnault-Roger et al. 2008, Allahvaisi et al. 2011 and AbdElhady 2012).

Spinosad is a biologically derived insecticide produced via fermentation culture of the actinomycete *Saccharopolyspora spinosa*, a soil bacteria. It is composed of a two members of the chemical class of 12-membered macrocyclic lactones in a unique tetracyclic ring. Spinosyn A and D are identical in structure except for an additional methyl group on the core macrolide of spinosyn D. Commercial formulations contain a spinisyn A to spinosad D ratio of approximately 85:15.

It is commonly known as "reduced-risk" insecticide. The contact action of spinosad has low toxicity (Bert *et al.* 1997). Reports on the effects of contact action of spinosad against the development stages of *Callosobruchus* sp. are scanty. So, the present experiment was designed to determine the contact activities of spinosad on the oviposition, fertility, adult emergence and sex ratio of *C. chinensis* for three successive generations.

2. Materials and Methods

Collection of the insects

C. chinensis was collected from a rearing culture maintained at the Controlled Temperature (30±0.5°C) room, Entomology and Insect Biotechnology Laboratory, Institute

of Biological Sciences, Rajshahi University. Newly emerged adult beetles were collected for sub-culture of the experiments.

Concentrations used

Pilot experiments were conducted with *C. chinensis* released on filter papers treated with different concentrations of spinosad. The concentration for each spinosad doses were determined where adult mortality rate was zero percent. Depending on concentrations, a series of three concentrations for each compound were selected. The doses were 6.2×10^{-5} , 3.1×10^{-5} and 1.5×10^{-5} ppm/ μ ml.

Experimentations

Filter papers (9 cm) were soaked in different concentrations of spinosad solutions and dried at room temperature for an hour. Previously sterilized uninfested 2gm lentil seeds placed in different petri dishes. After that five mated females of C. chinensis released separately. The petri dishes were covered and kept until the eggs hatched and develop as adults. The emerged adults were counted and recorded. It was treated as F₁ generation. Again a treated filter paper with same concentrations of spinosad was placed in the petri dish. Previously five mated females of C. chinensis from F₁ generation were released in the prepared petri dishes. The experiments have done three successive generations. A similar set of experiment was carried on filter paper soaked with distilled water only, as a control. All the experiments were conducted at $30\pm1^{\circ}$ C in the CT room and replicated three times.

After 1st generation the present experiments were divided into three reproductive combinations or crosses, which are as follows:

- Treated males were paired with untreated females (TU)
- Untreated males were paired with treated females (UT)
- Treated males were paired with treated females (TT)

Each pair of the concentration, spinosad was kept separately in petri dish which was checked. The experiment was conducted and the parameters recorded of the second and third generations, similarly as first generation.

ResearchGate Impact Factor (2018): 0.28 | SJIF (2018): 7.426

The eggs laid by females on the surface of the seeds were counted after 24 hours with the help of magnifying glass and their numbers were counted. The hatching was confirmed when the frass was observed by the magnifying glass. The emerged adults were counted and their sexes were recorded. The mortality beetles were observed after 24-, 48- and 72 hours. Adults were considered to be dead when probing with a hot needle failed to produce a response (Yousefinezhad-Irani and Asghar 2007).

Data analysis: The mortality data were corrected by Abbott's formula:

Parameters observed

The following parameters are observed

- a) Effects on progeny production
- b) Effects on fertility
- c) Effects on adult mortality and
- d) Effects on the sex ratio

Statistical analysis

The effect of spinosad on the progeny products of various concentrations were tested by ANOVA and Tukey's test. The percent reduction of fertility rate in treatment compared to control (PRC) were calculated by the formula provided by Mian and Mulla (1982).

3. Results and Observation

Effect on progeny products of C. chinensis

The progeny productions rate of F_1 generation of C. *chinensis* was found that concentration was increased progeny productions rate were decreased (Table 1). In the highest concentration 6.2×10^{-5} ppm/ μ ml, the male production was 32.67 and the female was 30.33 among the eggs of 72.33. In the lowest concentration 1.5×10^{-5} ppm/ μ ml, the male (40.33) and the female (39.0) progeny produced from the deposited eggs 86.33 whereas in control male progeny production was 41.33 and the female production was 39.67 among the eggs of 90.67.

In the second and third generations the experiment were divided in three pair groups which was described earlier. In the second generation three concentrations of spinosad in TU group the male progeny productions rate respectively were 27.67, 30.00, 31.00 and the female rate 21.67, 24.67, 24.00 among the eggs rate 52.67, 59.67, 61.33. In UT group the progeny productions rate of male 21.00, 22.00, 30.33 and the female rate 16.33, 21.00, 21.33 among the eggs rate 48.33, 52.67, 60.00. Similarly in TT group, the progeny productions rate of male 21.00, 22.00, 21.33 and the female rate 16.00, 18.33, 21.00 among the eggs rate 39.00, 45.67, 51.33 respectively (Table 1).

In the third generation, three concentrations of spinosad in TU group produced male 18.67, 22.33, 20.00 and the female rate 16.67, 18.33, 19.67 respectively among the eggs rate 46.67, 50.00, 55.00. The UT group the progeny products rate of male 16.33, 20.67, 24.67 and the female rate 14.00, 16.33, 18.33 among the eggs rate 36.33, 44.00, 49.67. The TT group the progeny products rate of male 13.67, 19.00,

19.67 and the female rate 11.00, 12.00, 11.67 among the eggs rate 29.33, 35.00, 40.00 respectively (Table 1).

The present experiment noticed that progeny productions rate were reduced drastically in TT group than UT and TU groups. Female progeny productions rate was less than male progeny productions rate. The F_3 generation were showed better result than F_2 and F_1 generations.

Table 1: Effect of pair types with spinosad where treated male or female or both, on the fertility (%) of the egg hatched by *C. chinensis* in three successive generations

Gene-	e- Concentration		Number of	Total	Fertility	PRC
rations	ppm/µml	type	eggs	Hatch	%	
	Control	-	90.67±.0.66	81.00±0.88	89.33	-
1 st	6.2×10^{-5}	-	72.33±1.20	63.00±0.60	87.10	22.22
1	3.1×10 ⁻⁵	-	79.67±0.88	72.33±1.13	90.78	10.70
	1.5×10 ⁻⁵	-	86.33±0.88	78.67±0.33	91.12	2.87
	_	TU	52.67±1.20	49.34±1.05	93.63	39.08
	6.2×10^{-5}	UT	48.33±0.88	37.33±0.57	77.23	53.91
		TT	39.00±1.00	37.00±0.57	94.87	54.32
		TU	59.67±0.88	54.67±0.74	91.62	32.50
2^{nd}	3.1×10^{-5}	UT	52.67±1.20	43.00±0.57	81.64	46.91
		TT	45.67±1.20	40.33±0.72	88.30	50.20
	1.5×10 ⁻⁵	TU	61.33±0.88	55.00±0.57	89.67	32.09
		UT	49.67±1.20	43.00±1.64	86.57	46.91
		TT	40.00±1.73	31.34±1.04	78.35	61.30
		TU	46.67±0.88	35.34 ± 0.88	75.72	56.34
	6.2×10^{-5}	UT	36.33±0.88	30.33±0.90	83.48	62.55
		TT	29.33±0.66	24.67±0.45	84.11	69.54
		TU	50.00±1,15	40.66±0.60	81.32	49.80
$3^{\rm rd}$	3.1×10^{-5}	UT	44.00 ± 1.73	37.00±0.67	84.09	54.32
		TT	37.00±1.15	31.00±1.52	83.57	61.72
		TU	55.00±1.15	41.67±0.94	75.76	48.62
	1.5×10^{-5}	UT	49.67±1.20	43.00±1.64	86.57	46.91
		TT	40.00±1.73	31.34±1.04	78.35	61.30

Effect on fertility on C. chinensis: All concentrations of spinosad reduced fertility of the deposited eggs in a concentration dependent manner in three successive generations (Table 1). The percentage of fertile eggs in control was found as 89.33. In 1st generation the highest fertility PRC value 22.22 at the concentration of 6.2×10⁻⁵ but lowest PRC value was 2.87 of 1.5×10⁻⁵ concentration. In second generation highest PRC value 54.32 in TT pair type where the concentration was 6.2×10⁻⁵ but lowest PRC value 32.09 in TU pair type in 1.5×10^{-5} concentration. In the same concentration, the highest PRC value was 69.54 in TT pair type in same highest concentration and lowest PRC value was 48.62 in the same lowest concentration in 3rd generation. The present experiment explained the treated and untreated pair types effected egg laying of the female beetles, but showed no significant effect on their viability. Only both treated male and treated female paired showed the fertility of egg significant results.

Though, compared $2^{\rm nd}$ and $3^{\rm rd}$ generations among different concentrations and different pair types 3.1×10^{-5} and 1.5×10^{-5} concentrations showed non-significant results. Only 6.2×10^{-5} dose was showed significant result. Moreover, $3^{\rm rd}$ generation was showed better results than $2^{\rm nd}$ and $1^{\rm st}$ generations.

Table 2: Mortality (Mean±SE) of *C. chinensis* at different concentrations of spinosad after three generations time

Volume 8 Issue 6, June 2019

www.ijsr.net

Licensed Under Creative Commons Attribution CC BY

ResearchGate Impact Factor (2018): 0.28 | SJIF (2018): 7.426

Gene-	Concentrations	Pair	Number of	T . 1 II . 1	Mortality	DDC
rations	ppm/µml	type	eggs	Total Hatch	$(Mean \pm SE)$	PRC
	Control	-	90.67±.0.66c	81.00±0.88	10.66±0.23a	-
	6.2×10 ⁻⁵	-	72.33±1.20	63.00±0.60	14.19±0.45b	33.11
1 st	3.1×10 ⁻⁵	-	79.67±0.88	72.33±1.13	13.41±0.27b	25.79
	1.5×10 ⁻⁵	-	86.33±0.88	78.67±0.33	13.27±0.24b	15.72
		TU	52.67±1.20	49.34±1.05	16.55±0.47d	55.25
	6.2×10 ⁻⁵	UT	48.33±0.88	37.33±0.57	16.65±0.49d	56.19
		TT	39.00±1.00	37.00±0.57	17.33±0.28de	62.57
		TU	59.67±0.88	54.67±0.74	15.87±0.33cd	48.03
2 nd	3.1×10^{-5}	UT	52.67±1.20	43.00±0.57	16.01±0.30d	50.18
۷		TT	45.67±1.20	40.33±0.72	16.37±0.13d	53.56
		TU	61.33±0.88	55.00±0.57	14.82±0.24c	39.02
	1.5×10 ⁻⁵	UT	49.67±1.20	43.00±1.64	15.56±0.54c	45.96
		TT	40.00±1.73	31.34±1.04	15.62±0.03cd	46.52
		TU	46.67±0.88	35.34±0.88	18.23±0.29ef	71.01
	6.2×10 ⁻⁵	UT	36.33±0.88	30.33±0.90	18.72±0.10g	75.60
		TT	29.33±0.66	24.67±0.45	19.67±0.40h	84.52
		TU	50.00±1,15	40.66±0.60	18.27±0.49ef	71.38
3 rd	3.1×10 ⁻⁵	UT	44.00±1.73	37.00±0.67	18.43±0.26ef	72.88
		TT	35.00±1.15	31.00±1.52	18.68±0.43g	75.23
		TU	55.00±1.15	41.67±0.94	17.45±0.33e	63.69
	1.5×10 ⁻⁵	UT	49.67±1.20	43.00±1.64	17.88±0.36ef	67.72
		TT	40.00±1.73	31.34±1.04	18.00±0.15ef	68.85

Effect on mortality on C chinensis

The tested concentrations of spinosad revealed that mortality of *C. chinensis* was higher in $3^{\rm rd}$ generations than $2^{\rm nd}$ and $1^{\rm st}$ generations (Table 2). The concentrations were found to be potential causing mortality based on the contact compare to control. The average mortality was highest 19.67 ± 0.40 at 6.2×10^{-5} ppm in third generation in treated male and treated female pair type group but lowest was 14.82 ± 0.24 at $1.5\times10^{-}$

⁵ ppm in second generation in treated male and untreated female pair groups. After 1st generation the experiments were divided in three pair groups. Results shows that the first generation was not significant. There were significant differences in mean mortality pair type groups between exposure generation and between concentrations. In addition, the interaction between exposure generation and concentration were significant (Table 2).

Table 3: Effects of spinosad on progeny productions of *C. chinensis* of the F_1 generation (Mean \pm SE)

Concentrations ppm/µml	No. of eggs	Hatch					
Concentrations ppin/µmi	No. of eggs	Male	Female				
Cont.	90.67±. 0.66c	41.33±.088b	39.67±0.88c				
6.2×10 ⁻⁵	72.33±1.20a	32.67±0.88a	30.33±0.33a				
3.1×10^{-5}	79.67±0.88b	38.33±1.76b	34.00±0.57b				
1.5×10 ⁻⁵	86.33±0.88c	40.33±1.20b	39.00±0.57c				
In a column means followed by same letter (s) are not significantly different at p < 0.05 according to Tukey HSD Test.							

Table 4: Effects of spinosad on progeny productions of *C. chinensis* of the F₂ generation

Concentrations	Treated male	and untreate	ed female	Untreated ma	le and treat	ed female	Treated male and treated female		
	No. of egg	Hatch		No of ago	Hatch		No of ago	Hatch	
ppm/µml	No. of egg	Male	Female	No. of egg	Male	Female	No. of egg	Male	Female
Control	90.67	41.33	39.67	90.67	41.33	39.67	90.67	41.33	39.67
Control	$\pm 0.66c$	±0.88b	±0.88c	$\pm 0.66c$	±0.88c	±0.88c	±0.66d	±0.88b	±0.88c
6.2×10- 5	52.67	27.67	21.67	48.33	21.00	16.33	39.00	21.00	16.00
0.2×10- 3	±1.20a	±1.45a	±0.66a	±0.88a	±0.57a	±0.57a	±1.00a	±0.57a	±0.57a
3.1×10-5	59.67	30.00	24.67	52.67	22.00	21.00	45.67	22.00	18.33
3.1×10-3	±0.88b	±1.15a	±0.33b	±1.20a	±0.57a	±0.57b	±1.20b	±0.57a	±0.88ab
1.5×10-5	61.33	31.00	24.00	60.00	30.33	21.33	51.33	24.33	21.00
1.5×10-5	±0.88b	±0.57a	±0.57b	±1.15b	±0.33b	±1.20b	±0.88c	±1.76a	±0.57b
In a column means followed by same letter (s) are not significantly different at p < 0.05 according to Tukey HSD Test.									

Table 5: Effect of spinosad on progeny productions of *C. chinensis* of the F₃ generation

Concentrations	Treated male and untreated female			Untreated male and treated female			Treated male and treated female		
	No. of egg	Hatch		No of ogo	Hatch		N6	Hatch	
ppm/µml		Male	Female	No. of egg	Male	Female	No. of egg	Male	Female
Control	90.67	41.33	39.67	90.67	41.33	39.67	90.67	41.33	39.67
Control	± 0.66c	±0.88b	±0.88b	±0.66d	±.088c	±0.88c	±0.66c	±0.88c	±0.88b
6.2×10- 5	46.67	18.67	16.67	36.33	16.33	14.00	29.33	13.67	11.00
0.2×10- 3	±0.88a	±0.88a	±0.88a	±0.88a	±0.66a	±1.15a	±0.66a	±0.33a	±0.57a

Volume 8 Issue 6, June 2019

www.ijsr.net

Licensed Under Creative Commons Attribution CC BY

ResearchGate Impact Factor (2018): 0.28 | SJIF (2018): 7.426

	3.1×10-5	50.00 ±1,15a	22.33 ±0.33a	18.3 ±30.8a	44.00 ±1.73b	20.67 ±1.20ab	16.33 ±0.33ab	35.00 ±1.15b	19.00 ±1.52b	12.00 ±1.52a
	1.5×10-5	55.00	22.00	19.67	49.67	24.67	18.33	40.00	19.67	11.67
		±1.15b	±1.00a	±0.88a	±1.20c	±2.40b	±0.88b	±1.73b	±0.88b	±1.20a
In a column many followed by some letter (a) are not significantly different at n < 0.05 according to Tylkay IICD Test								Cost		

Effect of sex ratio of *C. chinensis* in three successive generations:

The results of sex ratio of 1^{st} generation of *C. chinensis* in mated condition with different concentrations of spinosad. These ratio represented from female to male. It is striking to note that the number of progeny production is always lower in higher concentration 6.2×10^{-5} compared to the number of progeny produced in lower concentrations 3.1×10^{-5} and 1.5×10^{-5} .

It is found that, in 1st generation of C. chinensis progeny production of male and females were very same ratio. It was also showed very similar results in the control concentration. After 1st generation, the experiments were divided into three pair groups. It is also obvious that the TT pair type of C. chinensis always higher than other two pairs in the tested concentrations. In second (Table 3) and third generations (Table 5), female progeny production was lower than male progeny production. After generations to generations different concentrations of spinosad and different pair types of both sexes was decreased remarkably. The highest concentration 6.2×10⁻⁵, in the TT pair type there was very low female progeny production of C. chinensis and it also noticed that other pair types of sex ratio were very remarkable. These results were so significant than the control.

In 1st generation, the female or male ratio was recorded as 1.21 to 1 of C. chinensis in the highest concentration of spinosad. The ratios were 1.11 to 1 and 1.05 to 1 respectively in other two 3.1×10^{-5} and 1.5×10^{-5} concentrations. In 2nd generation, the highest concentrations of TU, UT and TT pair groups, the sex ratio were 1.27 to 1, 128 to 1 and 1.31 to 1 respectively. In 3.1×10^{-5} dose, the sex ratio of C. chinensis is 1.21 to 1, 1.04 to 1 and 1.20 to 1 respectively TU, UT and TT pair groups. In 1.5×10⁻⁵ dose, the sex ratio of the same beetles were 1.29 to 1, 1.34 to 1 and 1.68 to 1 respectively TU, UT and TT pair groups. Finally in 3rd generation the highest doses and similar pair groups the sex ratio were 1.11to 1, 1.16 to 1 and 1.24 to 1 respectively. These showed very significant results because the progeny produced were reduced generation to generations. These sex ratio was not much difference from among the concentrations and among pair groups.

4. Discussion

The present experiments revealed that there was significant impacts of the contact activity of different concentrations of spinosad and exposure generations against egg hatchability of *C. chinensis*. Egg hatchability, fertility and mortality were found to be dependent on the concentrations and successive generations. Similar effects of spinosad and other extraction have been reported on other stored product insects (Tabu *et al.* 2012, Ratnasekera and Rajapakse 2012 and Fatiha *et al.* 2014).

The biopesticide Spinosad controls many insect pests of stored-food products reported Niango et al. (2010). Laboratory and field trials were carried out to determine the efficacy of this pesticide against Callosobruchus maculatus, the main storage pest of cowpea, Vigna unguiculata, Walp, in West Africa. Spinosad caused high mortality of adult C. maculatus and decreased the number of eggs laid by females. Spinosad, however, was less toxic in the 24 h treatment to C. maculatus than deltamethrin, an insecticide commonly used in Burkina Faso to control this insect. In "on-farm" experiments, Spinosad was effective in controlling C. maculatus. After 6 months of storage, the number of insects emerging from cowpeas seeds was reduced by >80% by coating seeds with Spinosad but only by 43% by coating with deltamethrin. Less than 20% of the seeds were perforated in the Spinosad treatment compared with 29% for deltamethrin. Spinosad controlled C. maculatus throughout the 6 mo of cowpea storage whereas deltamethrin failed to control C. maculatus after 3 mo of storage. Spinosad has the potential to be more effective in controlling C. maculatus than deltamethrin. The present results are similar to the above findings.

Spinosad's performance was evaluated on four classes of wheat by Fang et al. (2002) (hard red winter, hard red spring, soft red winter, and durum wheats) against adults Rhyzopertha dominica (F.), Sitophilus oryzae (L.), Oryzaephilus surinamensis (L.), Tribolium castaneum (Herbst) and larvae of Plodia interpunctella (Hübner). Beetle adults (25) or P. interpunctella eggs (50) were exposed to untreated wheat and wheat treated with spinosad at 0.1 and 1 mg (AI)/kg of grain. On all untreated wheat classes, adult beetle mortality ranged from 0 to 6%, and P. interpunctella larval mortality ranged from 10 to 19%. The effects of spinosad on R. dominica and P. interpunctella were consistent across all wheat classes. Spinosad killed all exposed R. dominica adults and significantly suppressed progeny production (84-100%) and kernel damage (66-100%) at both rates compared with untreated wheat. Spinosad was extremely effective against P. interpunctella on all wheat classes at 1 mg/kg, based on larval mortality (97.6-99.6%), suppression of egg-to-adult emergence (93-100%), and kernel damage (95-100%), relative to similar effects on untreated wheats. The effects of spinosad on S. oryzae varied among wheat classes and between spinosad rates. Spinosad was effective against S. oryzae, O. surinamensis and T. castaneum only on durum wheat at 1 mg/kg. Our results suggest spinosad to be a potential grain protectant for R. dominica and P. interpunctella management in stored wheat.

In another study Spinosad performance were recorded against *Plodia interpunctella* (Hübner), *Tribolium castaneum* (Herbst), *Rhyzopertha dominica* (F.), and *Cryptolestes ferrugineus* (Stephens), winter wheat stored on farms in northeastern Kansas by Huang *et al.* (2004). Fifty eggs of *P. interpunctella* and 25 beetle adults of each species

Volume 8 Issue 6, June 2019

www.ijsr.net

Licensed Under Creative Commons Attribution CC BY

ResearchGate Impact Factor (2018): 0.28 | SJIF (2018): 7.426

were exposed to 100 g of untreated wheat or wheat treated with various rates of spinosad, to determine susceptibility of field and corresponding insecticide-susceptible laboratory strains. Mortality of beetle adults and P. interpunctella larvae was assessed after 7 and 21 d post infestation, respectively. Field strains of P. interpunctella, C. ferrugineus, and T. castaneum were less susceptible to spinosad than the corresponding laboratory strains. The LD₅₀ and LD₉₅ values for P. interpunctella and C. ferrugineus field strains were 1.7-2.5 times greater than values for corresponding laboratory strains. Adults of both laboratory and field strains of T. castaneum were tolerant to spinosad, resulting in <88% mortality at 8 mg/kg. The LD₅₀ and LD₉₅ values for the field strains of T. castaneum were 2.0-7.5 times greater compared with similar values for the laboratory strain. The field and laboratory strains of R. dominica were highly susceptible to spinosad, and one of the field strains was relatively less susceptible to spinosad than the laboratory strain. Our results confirm a range of biological variability in field populations, which is consistent with findings for other compounds, and underscores the need to adopt resistance management programs with stored grain insect pests. The baseline data generated on the susceptibility of the four insect species to spinosad will be useful for monitoring resistance development and for setting field rates.

Chakraborty and Mondal (2014) observed the hatching of eggs laid by *C. chinensis* was 92% and 95.5% in two successive generations in cowpea. The increment of the egg laying percentage shows that cowpea is preferable host for deposition of more eggs. Tabu *et al.* (2012) observed the mean average percentage of eggs hatched were 91.6 % with a range of 91 to 93% which was in close agreement with the report of Teshome (1991), who reported that the average egg hatchability of *C. chinensis* on chickpea was 89.6% with a range of 83 to 96.4%.

The edible oil of G. abyssinica, B. juncea and L. usitatissimum (at the highest concentration) showed significantly the highest mortality when compared to the untreated check. It was even effective at the lowest concentration although the efficacy is gradual in killing the adult bruchids. In general, these results suggested that it is possible to control C. chinensis effectively by using edible oils though there appeared variation of efficacy due to concentration and exposure period. According to Inge (2004), oil may also kill the insect eggs. When the egg is already present at the surface of the seed or inside the seed, the oil coating prevents gaseous exchanges. So the larvae inside the egg or the kernel will die due to lack of oxygen. Abraham (2003) compared oils of maize, sunflower, G. abyssinica and A. indica against the maize weevil at Bako, Ethiopia. He further obtained effective control of the maize weevil with all oils at the rates ranging from 5 to 10 ml kg-1. Similarly, Talekar (1976) also examined that, coating seeds with 5-10 ml of vegetable oils per kg of seeds, protect stored seeds from bruchid infestation. Mixing of soya bean or ground nut oil at the rate of 2-3 ml/100 g seeds gave mung bean considerable in bruchid infestation for up to two months.

References

- [1] Abbot W S 1925. A method of computing the effectiveness of an insecticide. *J. Econ. Ent.* **18**: 265-267.
- [2] Abd-Elhaldy H K 2012. Insecticidal activity and chemical composition of essential oil from *Artemisia judiaca* L. against *Callosobruchus maculatus* (F.) (Coleoptera: Bruchidae). *J. Plant Prot. Res.* **52**: 347–352.
- [3] Abraham T 2003. Studies on some non-chemical insect pest management options on farm-stored maize in Ethiopia. Ph D Thesis, Justus Leibig University of Giessem, Germany, pp 246.
- [4] Allahvaisi S, Maroufpoor M, Abdolmaleki A, Hoseini S A and Ghasemzadeh S 2011. The effect of plant oils for reducing contamination of stored packaged foodstuffs. *J. Plant Prot. Res.* **51**: 82–86.
- [5] Bekele A J, Ofori D O and Hassanali A 1995. Products derived from the leaves of *Ocimum kilimandscharicum* (Labiatae) as post-harvest grain protectants against the infestation of three major stored product insect pests. *Bull Ent. Res.* **85**: 361-367
- [6] Bert B L, Lerson L L, Schoonover J R, Sparks T C and Thompson G D 1997.Biological propertis of spinosad. *Down to earth* **52**: 6-13.
- [7] Chakraborty S and Mondal P 2014. studies on the biology of pulse beetle (*C. chinensis* Linn) infesting cowpea. *International J. Curr. Res.***7**: .23512-23515.
- [8] Chiasson H, Delislen U, Bostanian N J and Vincent C 2008. Recherche, développement et commercialisation de FACIN, un biopesticide d'origine végétale. Étude d'un cas de réussiteen Amérique du Nord. [Research, development and commercialization of facin, a vegetable biopesticide. A case study of success in North America], 2nd ed. Lavoisier, Tech. & Doc., Paris, France, 546 pp.
- [9] Fang Lang, Bhadriraju Subramanyam and Frank H. Arthur 2002. Effectiveness of Spinosad on four classes of wheat against five stored-product Insects. *J. Econ. Ent.* 95: https://doi.org/10.1603/0022-0493-95.3.640
- [10] Fatiha A, Righi K, Khelil A and Pujade-Villar J 2014. Biological control against the cowpea weevil (*Callosobruchus chinensis* L., Coleoptera-Bruchidae) using essential oils of some medicinal plants. *J. Plant Protect. Res.* 54: 211-217.
- [11] Huang F, Subramanyam B and Toews M D 2004. Susceptibility of laboratory and field starins of four stored-product insect species to spinosad. *J. Econ. Ent.* **97**: 2154-2159.
- [12] Inge de Groot 2004. Protection of stored grains and pulses: The Storage of Tropical Agricultural Products. *Grodok* **18**: 56-67
- [13] Isman M B 2006. Botanical insecticides, deterrents, and repellents in modern agriculture and an increasingly regulated work. *Ann. Rev. Ent.* **51**: 45–66.
- [14] Isman M B 2008. Botanical insecticides: for richer, and for poorer. *Pest Management Sci.* **64**:8-11.
- [15] Mian L S and Mulla M S 1982. Biological activity of IGRs against four stored product coleopterans. *J. Econ. Ent.* **75**: 80-85.
- [16] Niango A S, Clementine M Ba L and Pittendriggh Binso-Dabire Barry R 2010. Effectiveness of spinosad (naturalytes) in controlling the cowpea storage pest,

Volume 8 Issue 6, June 2019

www.ijsr.net

Licensed Under Creative Commons Attribution CC BY

ResearchGate Impact Factor (2018): 0.28 | SJIF (2018): 7.426

- Callosobruchus maculatus (Coleoptera: Bruchidae). J. Econ. Ent. 103: 203-210.
- [17] Teshome L 1991. The biology and control of the Adzuki bean beetle (Callosobruchus chinensis L.) (Coleoptera: Bruchidae) on chickpea Cicer arietinum. M Sc Thesis presented to the school of Graduate studies of Alemaya University, Ethiopia, pp 109.
- [18] Thiam A and Ducommun G 1993. Protection Naturelle des Végétaux en Afrique. [Natural Plant Protection in Africa]. Environment and Development Action in the Third World (ENDA TM), Dakar, Senegal, 212 pp.
- [19] Ratnasekara D and Rajapakse R 2012. The potential use of indigenous plant materials against *Callosobruchus* chinensis L. and *Callosobruchus maculatus* L. (Coleoptera, Bruichidae) in stored legumes in Sri Lanka. *JBiopest* 5: 88-94
- [20] Regnault-Roger C, Philogene B J R and Vincent C 2008. *Biopesticides of vegetable origin*. 2nd Edition, Lavoisier, Paris, 550p.
- [21] Talekar S 1976. Biology: damage and control of bruchid pest of mungbean. The Asian Vegetable Research and development Center, Ethiopia
- [22] Yousefnezhad-Irani R and Asghar P A 2007. Effect of spinosad on adults of TRIBOLIUM CASTANEUM Herbst (Coleoptera: Tenebrionidae) and SITOPHILUS ORYZAE L. (Coleoptera: Curculionidae). *Pakistan J. Bio. Sci.* **10**:2505-2509.

Volume 8 Issue 6, June 2019 www.ijsr.net

Licensed Under Creative Commons Attribution CC BY