Insilico Prediction of B-Cell and T-Cell Epitopes for Chaperones from *Mycobacterium tuberculosis*

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Abstract: With *Mycobacterium tuberculosis* control becoming a global necessity and history of causing more death than HIV in 2015 crucial consideration is to be given to create an effective vaccine to cope up with the multi-drug resistant forms of *M. tuberculosis*. The only licensed TB vaccine Bacillus Calmette-Guérin (BCG) has drawbacks that limit its efficacy and applicability for which alternate approaches like peptide or epitope based vaccine that can offer safe and specific immune response without any cross reactivity also offering easy production except for an obstacle that for designing peptide vaccines a potential candidate selection from the hundreds or thousands of potential candidates in which many number of epitopes might be present in an antigenic protein but all may not antigenic and does not involve in immune response too. In the present study, two protein fimC and papD were selected. Antigenicity, B-cell and T-cell epitopes of these proteins were identified using Vaxijen, BCPrEd, ProPred I, ProPred. Binding affinity of the predicted epitope with MHC/HLA alleles were calculated using MHC Pred. Surface exposed epitope may be effectively involved in immune response which is predicted by PSORTb tool. Good binding epitopes were selected based on FC50 value against DRB1*0101 and DRB1*0401 allele. The finally selected potential epitopes are IRVNNTPSY, YRPAGLSDR from fimC and one epitope LGSAPVLGY from papD. This study suggested that the predicted epitopes can elicit both B-Cell and T-cell mediated immune response, however experimental validation are required.

Keywords: Antibiotic, Drugs, MDR, Vaccine, Epitope, MHC/HLA allele, prediction

1. Introduction

Tuberculosis one of the important health hazards worldwide possess a major role in India as identified by WHO. The drug susceptibility and molecular characterization of drug resistant forms need to be explored. *M. tuberculosis* follows mutations in genes that encodes drug targets or drug activating enzymes which are responsible for drug resistance. The multi-drug resistance forms of Tuberculosis acts as the major threat to all TB control programs. To obstruct the multi drug resistance (MDR) and extensive drug resistance (XDR) of tuberculosis, Peptide-based epitope vaccines offer great advantage over whole cell vaccines; that avoids non-protective responses and unwanted side effects. Peptide based vaccines provides hope for treatment and prevention of chronic infectious disease that are caused by virus and cancer cells (Hailemichael, Y and Wang, C.Y., 2013)

Humoral and adaptive immune responses are mediated by B-cell and T-cell epitopes, of which the former are linear (Continuous) and conformational (Discontinuous). Conformational epitope identification is solely based on X-Ray structure of antigen-antibody complex which is a time taking step. Computational algorithms are available for linear B-cell epitope prediction (Barlow et al., 1986). MHC binding peptide is a prerequisite for immune response, the MHC Class I (recognize CD8 + T-cells), MHC class II (recognize CD4+ T-cells). For the recognition of peptide bounded with MHC on the Antigen Presenting cells (APCs) both Helper T-Lymphocytes (HTL) and Cytotoxic T-Lymphocytes (CTL) are required (Pancre et al., 1996). The epitope functional study aid in the immunopathogenesis study understanding. It also helps in the better designing of protective epitopes and better improved epitope or peptide designing. (PMID 24715888 – Functions and potentials of *M. tuberculosis* epitope). Epitope based designs takes understanding a broader area of the multidrug resistance mechanism accelerating the vaccine productions.

In the present approach we have selected two chaperone protein fimC and papD from the *Mycobacterium tuberculosis* which can play role in Multi Drug resistance. Computational techniques were followed to predict novel peptide –epitope which can elicit both humoral and adaptive immune response.

2. Methodology

Antigenic properties of Target protein

The amino acid sequences of *Mycobacterium tuberculosis* chaperone protein fimC and papD were retrieved from UniprotKB database. Immunogenic properties of the retrieved sequences were identified by using Vaxijen 2.0 server with default value ≥ 0.4 to increase the prediction accuracy and to minimize the false positives. The sequences with vaxijen score ≥ 0.4 and were selected.

B-Cell and T-cell epitope prediction

The first screening steps were carried out in BCPrEd server with a cut-off score of 0.8 for 20-mer with vaxijen score ≥0.4. The predicted antigenic B-cell epitopes were subjected to T-cell epitope prediction based on binding affinity with MHC class I and II molecules using Propred I (Singh, H. and Raghava, G. P. S. 2003) and Propred (Singh, H. and Raghava, G. P. S. 2001). MHC class II binding peptides were predicted from 51 alleles and MHC class I binding peptides were predicted from 47 alleles. Epitopes binded with MHC I and MHC II were selected as promiscuous 9-mer epitope. A second screening was based on epitope localization on target protein which is carried out by Pepitope server (Mayrose et al., 2007) a platform based on PepSurf (Mayrose et al., 2007) and Mapitope (Bubli et al., 2007) algorithms used for best cluster prediction based on

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the structure based QSAAR simulation. For the best epitope cluster, IC50 values were calculated using MHCPred (Guan et al., 2003) to know its binding efficiency to the specific human alleles DRB1*0101 and DRB1*0401.

MDR protein modeling and Epitope characterization:
The protein structures were modeled using Phyre2 server and validated using PROCHECK. The modeled structure was then used in Pepitope server to predict the epitope localization on target protein.

3. Results

Protein localization & Antigenic character identification:
The Mycobacterium tuberculosis chaperone proteins fimC and papD were identified from Vaxijen score which is ≥ 0.4 that indicates the proteins are antigenic. The accession number, sequence length and Vaxijen score are shown in Table 1.

Table 1: Sequence length and antigenic properties of the fimC and papD protein

<table>
<thead>
<tr>
<th>S.No</th>
<th>Protein Name</th>
<th>Uniprot ID</th>
<th>Sequence length</th>
<th>Vaxijen Score</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>fimC</td>
<td>A0A1K3GKW8</td>
<td>234</td>
<td>0.67</td>
</tr>
<tr>
<td>2</td>
<td>papD</td>
<td>A0A1K3C1H2</td>
<td>252</td>
<td>0.46</td>
</tr>
</tbody>
</table>

Linear B-cell epitope prediction:
The number of identified B-cell epitopes from fimC and papD are 12 and 9. Among the identified B-cell epitopes, 10 were from fimC and 8 from papD found to be antigenic based on Vaxijen score ≥ 0.4. The predicted antigenic B-cell epitopes of fimC and papD are represented in Table 2.

Table 2: Predicted B-cell epitopes of fimC and papD

<table>
<thead>
<tr>
<th>Protein Name</th>
<th>Sequence position</th>
<th>B-cell epitopes</th>
<th>BCPred score</th>
<th>Vaxijen score</th>
</tr>
</thead>
<tbody>
<tr>
<td>fimC</td>
<td>157</td>
<td>SFQSGSHERVINNTSPYHIT</td>
<td>0.992</td>
<td>1.2076</td>
</tr>
<tr>
<td></td>
<td>68</td>
<td>PVPFELETTPISRINPQGANQ</td>
<td>0.973</td>
<td>0.7975</td>
</tr>
<tr>
<td></td>
<td>112</td>
<td>LDIPPSVETSSLKEKNSGQL</td>
<td>0.971</td>
<td>0.7899</td>
</tr>
<tr>
<td></td>
<td>30</td>
<td>YPKGAHDITARIINESATAA</td>
<td>0.848</td>
<td>1.1629</td>
</tr>
<tr>
<td></td>
<td>211</td>
<td>RGDHLTVSNINDQGNGVFT</td>
<td>0.793</td>
<td>1.2667</td>
</tr>
<tr>
<td></td>
<td>57</td>
<td>PGDSEALPAKVLVTFPLETEP</td>
<td>1</td>
<td>1.3646</td>
</tr>
<tr>
<td></td>
<td>115</td>
<td>PPVSESSLKKEKNSGOLAVR</td>
<td>1</td>
<td>1.1028</td>
</tr>
<tr>
<td></td>
<td>139</td>
<td>VFPYRPGALDRNATQRVSF</td>
<td>1</td>
<td>0.8093</td>
</tr>
<tr>
<td></td>
<td>30</td>
<td>YPKGAHDITARIINESATAA</td>
<td>1</td>
<td>1.1629</td>
</tr>
<tr>
<td></td>
<td>163</td>
<td>HSIRVNNTPSYHITETIT</td>
<td>1</td>
<td>1.1373</td>
</tr>
<tr>
<td>papD</td>
<td>164</td>
<td>TRQGDKITYVINNPYTTIS</td>
<td>0.992</td>
<td>1.0702</td>
</tr>
<tr>
<td></td>
<td>40</td>
<td>LNSERASIESINQNKELP</td>
<td>0.963</td>
<td>0.9094</td>
</tr>
<tr>
<td></td>
<td>216</td>
<td>SLGSAPVLGVYINDYGRPPK</td>
<td>0.88</td>
<td>0.4570</td>
</tr>
<tr>
<td></td>
<td>157</td>
<td>WQESTILTRQGDKJTYVNNPT</td>
<td>1</td>
<td>0.8977</td>
</tr>
<tr>
<td></td>
<td>203</td>
<td>APNSEEVLGNNAAASLGSAPV</td>
<td>1</td>
<td>0.9558</td>
</tr>
<tr>
<td></td>
<td>34</td>
<td>DTRVILNGSAESVEISN</td>
<td>1</td>
<td>0.1780</td>
</tr>
<tr>
<td></td>
<td>119</td>
<td>REIPPKTDKANTLQIALQTR</td>
<td>1</td>
<td>0.6700</td>
</tr>
<tr>
<td></td>
<td>180</td>
<td>VTISSASAAYGSKDISGFEA</td>
<td>0.997</td>
<td>0.8034</td>
</tr>
</tbody>
</table>

T-cell epitope prediction:
8 T-cell epitopes of fimC and 7 T-cell epitopes of papD were commonly binding with Class I and Class II MHC molecules. Among the identified commonly binding T-cell epitopes, 7 epitopes of fimC and 5 epitopes of papD were found to be antigenic based on Vaxijen score ≥ 0.4. The predicted antigenic T-cell epitope from fimC and papD are shown in Table 3. The predicted T-cell epitope binding with the number of class I and Class II MHC molecules are represented in Fig 1, 2.

Table 3: T-cell epitope of fimC and papD binding with class I and class II MHC

<table>
<thead>
<tr>
<th>Protein Name</th>
<th>Position</th>
<th>Common in ProPred I and ProPred II</th>
<th>Vaxijen score</th>
</tr>
</thead>
<tbody>
<tr>
<td>fimC</td>
<td>167</td>
<td>VNNTPSYH</td>
<td>0.7834</td>
</tr>
<tr>
<td></td>
<td>165</td>
<td>IRVNTSPY</td>
<td>0.6557</td>
</tr>
<tr>
<td></td>
<td>158</td>
<td>FOQSGSHER</td>
<td>1.8821</td>
</tr>
<tr>
<td></td>
<td>37</td>
<td>ITARINES</td>
<td>1.4025</td>
</tr>
<tr>
<td></td>
<td>41</td>
<td>INNESATAA</td>
<td>0.8367</td>
</tr>
<tr>
<td></td>
<td>141</td>
<td>YRPAGLSDR</td>
<td>0.9376</td>
</tr>
<tr>
<td></td>
<td>172</td>
<td>YHITET</td>
<td>0.8859</td>
</tr>
<tr>
<td>papD</td>
<td>224</td>
<td>YINDYGRP</td>
<td>1.0321</td>
</tr>
<tr>
<td></td>
<td>217</td>
<td>LGSAPVLGY</td>
<td>0.9953</td>
</tr>
<tr>
<td></td>
<td>208</td>
<td>VLGGSNAAL</td>
<td>1.3725</td>
</tr>
<tr>
<td></td>
<td>188</td>
<td>YGKGDIGSF</td>
<td>0.9230</td>
</tr>
<tr>
<td></td>
<td>180</td>
<td>VTISSASAAYGSKDISGFEA</td>
<td>0.8144</td>
</tr>
</tbody>
</table>

Epitope characterization:
To analyze the surface localization of the peptide to the respective target protein, 3-D structure of papD (PDB ID: 5K93) was retrieved from PDB. Three dimensional structure of fimC protein was modeled using Phyre2 server and validated using Ramachandran plot. More than 85% of the...
amino acids of all the modeled protein were present in allowed region of the Ramachandran Plot.

The best epitope clusters were identified using P-epitope server. Among the identified clusters, epitope located on outer surface of the protein was selected. The number of short listed epitope from each protein includes 2 epitope from fimC whose cluster score is 272.45 and 5 epitope from papD which has a cluster score of 134.06. Epitope located with the surface of fimC and papD are shown in Fig 3.

Figure 3: Surface localized epitopes of fimC and papD

Epitope validation
The potential T-cell epitopes of fimC are IRVNNTSPY, YRPAGLSDR and T-cell epitopes of papD are YINDYGGRP, LGSAPVLGY, VLGGNAASL, YKGKDISGF, VTISSASAA. The binding affinities of these potential T-cell epitopes were predicted against DRB1*0101 and DRB1*0401 allele. The final shortlisted epitopes based on IC_{50} value are IRVNNTSPY, YRPAGLSDR from fimC and one epitope LGSAPVLGY from papD. The results are represented in Table 4.

<table>
<thead>
<tr>
<th>Protein Name</th>
<th>Sequence position</th>
<th>T-cell epitope</th>
<th>IC_{50} Value (nM)</th>
<th>DRB1*0101</th>
<th>DRB1*0401</th>
</tr>
</thead>
<tbody>
<tr>
<td>fimC</td>
<td>165</td>
<td>IRVNNTSPY</td>
<td>8.11</td>
<td>341.98</td>
<td></td>
</tr>
<tr>
<td>papD</td>
<td>141</td>
<td>YRPAGLSDR</td>
<td>5.24</td>
<td>95.28</td>
<td></td>
</tr>
<tr>
<td></td>
<td>224</td>
<td>YINDYGGRP</td>
<td>10.64</td>
<td>767.36</td>
<td></td>
</tr>
<tr>
<td></td>
<td>217</td>
<td>LGSAPVLGY</td>
<td>42.95</td>
<td>325.09</td>
<td></td>
</tr>
<tr>
<td></td>
<td>208</td>
<td>VLGGNAASL</td>
<td>229.09</td>
<td>1291.22</td>
<td></td>
</tr>
<tr>
<td></td>
<td>188</td>
<td>YKGKDISGF</td>
<td>239.33</td>
<td>717.79</td>
<td></td>
</tr>
<tr>
<td></td>
<td>180</td>
<td>VTISSASAA</td>
<td>76.03</td>
<td>762.08</td>
<td></td>
</tr>
</tbody>
</table>

4. Discussion

Experimental approaches of vaccine designing are costly and time consuming to determine the epitope which elicit humoral and T-cell immunity (Sakib et al., 2014). In silico prediction of epitopes from immunogenic protein a familiar tool that can save time from large scale screening of experimental approach (Li et al., 2005). Proteins located on the surface of pathogenic organism mainly interact with host immune system since surface proteins are highly immunogenic. (Da Dana et al., 2003; Tran et al., 2006). Identification of target antigen being the initial step in rational vaccine design, the surface exposed proteins become more favorable for a potential vaccine (Almeida et al., 2003).

The protein sequence availed from UniprotKB database; provides information about protein localization. The proteins fimC and papD are found to be antigenic based on VaxiJen server whose prediction accuracy is nearly 70%-89 %. B-cell epitope prediction is essential, as the cells are the sites of antigen antibody interaction since antibodies are produced by B-lymphocytes. Antigen- antibody interactions are key events in clearance of pathogen by the immune system (Irving et al., 2001; Larsen JEP et al., 2006; Gershoni et al., 2007). This study revolves around the prediction of 20-mer B-cell epitope using BCPred server. Several studies reported that, multiple prediction tools are promising strategy for consensus approaches (Manzahavalk et al., 2008; Barh et al 2010). Antigenic score was predicted for the B-cell epitopes predicted by both the methods, in order to improve the accuracy of 20-mer B-cell epitope, two prediction methods the BCPred and AARD prediction were used which gives different B-cell epitopes.

Adaptive immunity is mediated by recognition of peptide antigens (T-cell epitopes) bound with major histocompatibility complex (MHC) molecules. The prediction of MHC peptide binding is lead priority for anticipating potential T-cell epitopes (Lafuente et al., 2009). The identification of both CD8 and CD4 T-cells are basic for the development of epitope based vaccines and in understanding the disease pathogenesis (Tchernev et al., 2006; Reche et al., 2006).

In the present study T-cell epitopes (9-mer) from antigenic B-cell epitope were predicted by using Propred I and Propred tool. Epitope which binds to MHC I and MHC II good antigenic score is selected as required epitopes.

Surface localization of peptide is one of the significant properties of peptide vaccine. We have predicted surface localization of T-cell epitope through P-epitope server. The modeled antigenic proteins were validated using PROCHECK. From this server, best cluster epitopes were selected and again its localization was cross checked with PSORTb 3.0.2 tool. In human, MHC molecules known as Human Leucocyte Antigen(HLA) whose polymorphic diversity in peptide binding specificity leads to effective immune response (Reche et al 2003; Kuhns et al., 1999). The Peptide binding to more than one HLA alleles is essential in vaccine development with impartial and extensive human population coverage. Selected peptide binding affinity predicted against human DRB1*0101 and DRB1*0401 HLA allele based on IC50 value, in which (i) IC50< 50 nM represent good binding. (ii) IC50=50-100 nM represent intermediary binder. (iii) IC50=500-5000 nM represent weak binder. (iv) IC50 > 5000 nM represent non binder. The predicted epitopes can induce both humoral and adaptive immune response but experimental approaches are required.

5. Conclusion

This study reported the epitopes from the chaperones of Mycobacterium tuberculosis. 3 potential epitopes from two
proteins were selected can be developed as peptide based vaccines in future against *Mycobacterium tuberculosis*. The epitopes predicted have the potential to elicit immune responses. Further evaluation of these in silico identified epitopes are highly warranted to confirm their immunogenicity and vaccine potential.

References


