

Evaluation of Antioxidant and Cytotoxic Activity of *Decalepis hamiltonii* on Hep G2 Cancer Cell Lines

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Abstract: *Decalepis hamiltonii* (Swallow root) belongs to the Asclepiaceae family is a monogenetic climbing shrub endemic to the Deccan peninsula and is used as a culinary spice due to its highly aromatic roots. It is useful as an appetizer, blood purifier, preservative and as a source of bioinsecticide for stored food grains. The present research work aims to identify the enzymatic and non enzymatic antioxidants in animal models, LDH and cytotoxicity activities using HEP G2 cancer cell lines were studied in vivo models to find the anticancer activity of *D. hamiltonii* methanolic and water extracts. In treated group of animals significant increased levels of enzymatic antioxidants like Superoxide Dismutase, Catalase and Glutathione Peroxidase were observed when compared to toxic group of animals. High concentration of LDH in 320µg/ml and 640µg/ml were identified as compared to other concentrations like 40µg/ml, 80µg/ml and 160µg/ml. Similarly significant levels of HEP G2 Cell Line viability were predicted at 640µg/ml of *D. hamiltonii* extract. These results might conclude the antioxidant and anticancer activity of *D. hamiltonii* extract.

Keywords: antioxidants, cytotoxicity, hepatic cancer, cell lines

1. Introduction

Plants have been the traditional source for raw materials and finished medicines since the dawn of civilization (Srinivasa Rao *et al.*, 2004). In India, herbal medicines have been the basis of treatment and cure for various diseases, physiological conditions in traditional methods practiced such as Ayurveda, Unani and Siddha. It has been estimated that about 20,000 plant species are known to have worldwide and phytochemical tests have been performed in about 5000 and nearly 1110 species are extensively utilized in Ayurveda (80%) and Allopathic medicine (33%) used as drugs (Swapna and Srinivasa Rao, 2013). Indian folk medicine comprises numerous herbal prescriptions for therapeutic purposes which may be as varied as healing wounds, treating inflammation due to infection, skin lesions, leprosy, diarrhoea, scabies, venereal diseases, snake bite and ulcers etc. The realization that many infectious pathogenic microorganisms are fast developing resistance against prevailing drugs has necessitated a search for new sources of antimicrobial compounds. Plants can synthesize a variety of secondary metabolites capable of providing protection against the infectious diseases (Srinivasa Rao *et al.*, 2005).

Decalepis hamiltonii, an endemic and endangered medicinal plant of southern peninsula. It is a liane and is locally called as maredu kommulu or maredu gaddalu (Devi and Latha 2012). Milky latex is present in the entire plant. Roots are harvested during summer months (Vedavathy, 2004). Roots of *D.hamiltonii* is a natural source of antioxidant and hence reduces the oxidative stress level and beneficial for health (Srivastava *et al.*,2005).

Majority of world population can't afford to conventional pharmaceutical products due to its high cost and thus have to rely upon the use of traditional and plant derived medicines (Srinivasa Rao *et al.*, 2006). As a result, plant based antihepatocellular carcinoma drugs are being searched all over the world as an alternative to existing drugs. Global estimates indicate that over 3/4th of the world population cannot afford the products of the Western pharmaceutical

industry due to its high cost and thus have to rely upon the use of traditional, plant-derived medicine (Avni Desai *et al.*,2008).

Developing countries can ill afford the products of the Western pharmaceutical industry due to their high and rising cost. The World Health Organization, therefore, recommends the inclusion of herbal medicines which are time tested, safe and available at affordable prices in the national health programmes. Moreover, traditional medicines show better culture acceptability, better compatibility and lesser side effects than synthetic drugs. This recognition has made the herbal system once again the center stage of health care program me, not only in India, but the world over (Umadevi *et al.*, 2013).

The most common histologic type of primary liver cancer, hepatocellular carcinoma (HCC), is a malignant tumor arising from hepatocytes, the liver's parenchymal cells. In agreement, in the early twentieth century, Wiliam Osler in the United States also reported that primary liver cancer was rare and are common in Asia and Africa, but are uncommon in Europe and North America. The highest liver cancer incidence rates in the world are reported by registries in Asia and Africa. Approximately 85% of all liver cancers occur in these areas, with Chinese registries alone, reporting over 50%.

Free radicals have been implicated in many diseases such as cancer, atherosclerosis, diabetes, neurodegenerative disorders and aging. It is believed that higher intake of antioxidant decreases risk of degenerative diseases particularly cardiovascular diseases and cancer. Several studies have shown that plant derived antioxidant related degenerative effects. Antioxidants are being used widely because of their therapeutic effects (Valko *et al.*,2007) .

HepG2 is a perpetual cell line consisting of human liver carcinoma cells, derived from the liver tissue of a 15-year-old Caucasian male who had a well-differentiated hepatocellular carcinoma. Hepatocellular carcinoma is, worldwide, the fifth most-common cancer. The morphology

of HepG2 cells is epithelial and they have 55 chromosome pairs. HepG2 cells can be grown successfully at a large scale, and secrete many plasma proteins, such as transferrin, fibrinogen, plasminogen and albumin. They can be stimulated with human growth hormone. HepG2 are adherent, epithelial-like cells growing as monolayers and in small aggregates. Not tumorigenic in immunocompromised mice.

2. Materials and Methods

Media: Eagle's Minimum Essential Medium, 10% FBS, 0.05% Trypsin-EDTA.

Plant material

The *Decalepis hamiltonii* root used for the investigation was collected from Tirumala Hills, Tirupati, India. The plant roots were authenticated by taxonomist Dr.K.Madhava Chetty, Department of Botany, Sri Venkateswra University, Tirupati, India.

Preparation of extract

The collected root of *D.hamiltonii* was dried in shade, crushed to coarse powder and was extracted with different solvents. Sequential extraction of the root powder was done with different solvents with increasing polarity i.e. hexane, chloroform, ethyl acetate, acetone, methanol and water. A total of 50 g of root powder was extracted in 0.5 l of the solvent in glass conical flask on a shaker for 24 h at room temperature. The extract was filtered with Whatmann paper no.1 and dried by lyophilization. The methanolic extract was prepared by soxhlet extraction in methanol. A 100 g of root powder was extracted (1 l) at 50°C for 24 h. Methanol was evaporated and the extract weighed (28 g). The solvents in plant extracts were evaporated under reduced pressure and extracts were stored in desiccators for further investigation of Antioxidant and Cytotoxic activities.

DPPH radical scavenging assay

The DPPH activity assay is the widely used assay (Goze et al., 2009; Saha et al., 2008; Prasad et al., 2009; Li et al., 2008). DPPH is a radical, which in the presence of antioxidants, is reduced from a stable free radical, 1,1-diphenyl-2-picryl hydrazyl (purple) to a diphenyl picryl hydrazine (yellow). Reaction of DPPH with hydroxyl group (R-OH) and R-NO₂ produces 2-(4-hydroxyphenyl)-2-phenyl-1-picryl hydrazine and 2-(4-nitrophenyl)-2-phenyl-1-picryl hydrazine, respectively (Naidoo, 2005). The amount of DPPH reduced is equivalent to the amount of antioxidants present in the study sample, thus the results obtained gives a true reflection of the extract's ability as an antioxidant.

The ability of the extracts to annihilate the DPPH radical (1,1-diphenyl-2-picrylhydrazyl) was investigated by the method described by (Blois, 1958). Stock solution of the plant root extracts was prepared to the concentration of 1

mg/ml. 100 µg of each extracts were added, at an equal volume, to methanolic solution of DPPH (0.1 mM). The reaction mixture is incubated for 30 min at room temperature; the absorbance was recorded at 517 nm. The experiment was repeated for three times. BHT was used as standard controls. The results are tabulated in Table 1. DPPH scavenging activity of this plant root extract was measured according to the method of Blois (1958). IC₅₀ values of the extracts and concentration of the extracts necessary to decrease the initial concentration of DPPH by 50% were calculated.

3. Results and Discussion

Decalepis is rich source for the various phytochemical constituents such as aldehydes, phenols, inositols, flavanoids, steroids, terpenoids etc. The roots are used for indigestion, appetite and as blood purifier etc.

MTT Cell Proliferation Assay

Cell proliferation and Viability are major factors to identify the cancer cells. This is a colorimetric assay that measures the reduction of yellow 3-(4,5-dimethyl thiazol-2yl)-2,5 diphenyl tetrazolium bromide [MTT] by mitochondrial succinate dehydrogenase. The MTT enters the cells and passes into the mitochondria where it is reduced to an insoluble, colored (dark purple) formazan product. The cells are then solubilized with isopropanol and the released solubilized formazan is measured spectrophotometrically. Since the reduction of MTT can occur in metabolically active cells the level of activity is a measure of the viability of the cells.

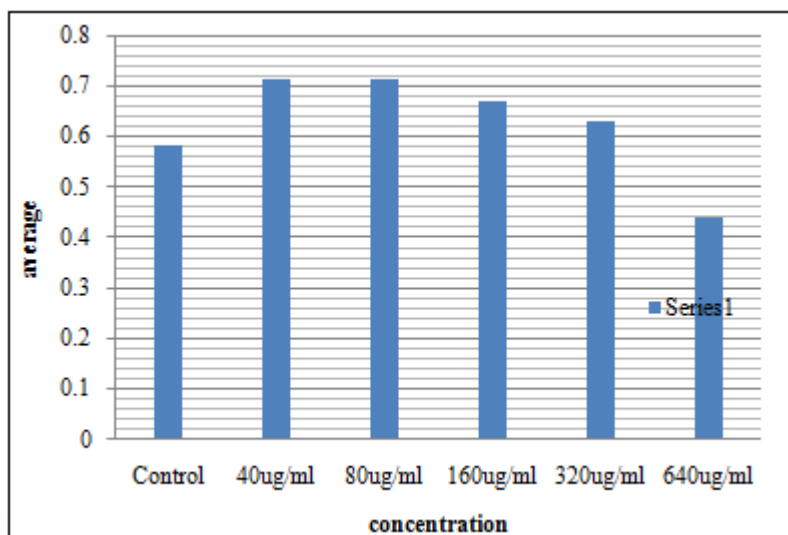
Measurement of cell viability and proliferation forms the basis for numerous in vitro assays of a cell population's response to external factors. The reduction of tetrazolium salts is now widely accepted as a reliable way to examine cell proliferation. The yellow tetrazolium MTT (3-(4, 5-dimethylthiazolyl-2)-2, 5-diphenyltetrazolium bromide) is reduced by metabolically active cells, in part by the action of dehydrogenase enzymes, to generate reducing equivalents such as NADH and NADPH. The resulting intracellular purple formazan can be solubilized and quantified by spectrophotometric means. The MTT Cell Proliferation Assay measures the cell proliferation rate and conversely, when metabolic events lead to apoptosis or necrosis, the reduction in cell viability. The number of assay steps has been minimized as much as possible to expedite sample processing. The MTT Reagent yields low background absorbance values in the absence of cells. For each cell type the linear relationship between cell number and signal produced is established, thus allowing an accurate quantification of changes in the rate of cell proliferation. The herbal extract showed a significant effect on growth of the cancer cell lines.

Table 1: Cell proliferation and viability assay of HEP G2 Cell Line

	1st Concentration	2nd Concentration	3rd Concentration	Average	% Viability
Control	0.61	0.554	0.584	0.583	100.0
40ug/ml	0.71	0.744	0.684	0.713	122.3
80ug/ml	0.689	0.772	0.682	0.714	122.6
160ug/ml	0.579	0.75	0.685	0.671	115.2
320ug/ml	0.611	0.702	0.579	0.631	108.2
640ug/ml	0.415	0.469	0.436	0.440	75.5

From the **Table 1** it is illustrated that different concentrations 40 µg/ml, 80µg/ml, 160µg/ml, 320µg/ml and

640µg/ml of plant extracts and their responses were identified.



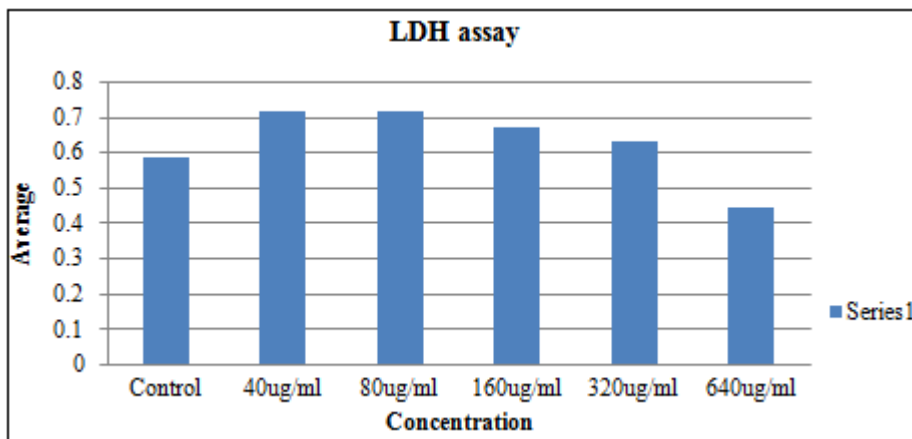
LDH Assay

Apoptosis and necrosis are two major forms of cell death observed in normal and disease pathologies. Although there are many assays for detection of apoptosis, relatively few assays are available for measuring necrosis. A key signature for necrotic cells is the permeabilization of plasma membrane (**Francis Ka-Ming Chan et al., 2014**). This event can be quantified in tissue culture settings by measuring the release of the enzyme lactate dehydrogenase (LDH). When combined with other methods, measuring LDH release is a useful method for detection of necrosis. In this we describe the step-by-step procedure for detection of LDH release from necrotic cells using a microtiter plate based colorimetric absorbance assay. the release of lactate was monitored after 48 hours of treatment in this study. This is because the content of lactate in the medium will affect the cell growth profile. Schneider (1996) reported that the toxic action of lactate was probably due to the acidic pH and osmolarity activity on the cells, particularly at high concentration (>20 mM). In the presence of c-myc, the genes of glycolytic enzymes, namely lactate dehydrogenase–A (LDHA) and GLUT1 would be transactivated to enhance both glucose uptake and lactate production (Shim, 1997).

LDHA was also reported could be upregulated in several tumors and it is essential for c-myc-mediated transformation (Shim, 1997). Actually, lactate was released as the by-product during cell growth. However, if the concentration of lactate in the medium was too high, it would affect the cell growth. Therefore, the content of lactate is crucial to monitor in order to avoid the side effect of lactate to the growth process of cells. The lactate release profile showed that in fact, the amount of lactate present in the medium did not significantly affect the cell growth (Fig. 3(b)). Hence, the growth profile of the cells was mainly Influenced by the plant extracts, but not because of the content of lactate released into the medium.

Table 2: Lactate Dehydrogenase (U/L)

	1 st Concentration	2 nd Concentration	3 rd Concentration	Average
Control	28	21	28	26.000
40ug/ml	28	29	27	28.000
80ug/ml	25	24	25	24.667
160ug/ml	24	21	24	23.000
320ug/ml	23	31	30	28.000
640ug/ml	44	35	33	37.333



4. Conclusion

Interest in the pharmacological effects of bioactive compounds on cancer treatments and prevention has increased dramatically over the past twenty years. It has been shown to possess numerous anti-cancer activities in various cancer cells through different forms of cytotoxic effects without exhibiting considerable damage to normal cells (Katiyar *et al.*, 2009; Mantena *et al.*, 2006a). Hepatocellular carcinoma (HCC) is the third most common cause of cancer-related deaths worldwide, and about half a million individuals die from this disease annually (El-Serag, 2004; Parkin *et al.*, 2005). Adriamycin (ADM) has been commonly used as a chemotherapeutic drug in the treatment of HCC (Schwartz, 2004). However, high concentrations of ADM tend to cause severe toxicity to normal tissues, including cardiotoxicity (Bristow *et al.*, 1978). Accumulating evidences suggest that marine yeast may be a potential chemotherapeutic or a chemopreventive agent based on its ability to induce apoptosis in cancer cells with relatively low toxicity to normal cells. Further studies with in vivo and clinical trials needs to be conducted to establish this herbal drug as a safe agent for cancer therapy (Lau *et al.*, 2001).

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