Isolation of Seed Borne Fungi Associated with Pigeon pea (*Cajanus cajan*, *Linn*.) Seeds

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Abstract: Using ISTA techniques, the seed borne fungi of Pigeon pea (Cajanus cajan Linn.) was studied. A total of 15 genera and 29 species of fungi were isolated, and been recorded from seeds of Pigeon pea. The blotter method agar plate method was found to be most suitable technique for detection of fungi in Peageon pea. There was the detection of Fusarium spp, Aureobasidium pullulans, Gleomatix sp., Gleosporium sp., Trichoderma sp. and Penicillium purpurogenum done by these two methods.

Keywords: Cajanus cajan, Linn., Seed Mycoflora, Blotter Paper Method, Agar Plate Method, ISTA, etc

1. Introduction

Pigeonpea [Cajanus cajan (L.) Millsp.] is a perennial member of the family leguminosae. It is a multi-purpose species, is extensively used as food grain and green manure crop for soil fertility amelioration in local cropping systems. Other common names are "Red gram, Congo pea, Gungo pea, Gunga pea, and No-eye pea". It is an important grain legume crop of rain-field agriculture in the tropics and subtropics. Compared with other grain legumes, pigeonpea ranks only sixth in area and production, but it is used in more diverse ways than others [5, 10, 11, 21]. Many fungal diseases (31), involving 45 pathogens, are known; the most serious is wilt disease (Fusarium udum), favored by soil temperatures of 17°-20°C. The fungus enters the plant through the roots and may persist in soilborne stubble for a long time. The only effective control measure is development of resistant cvs (e.g., 'C-ll,' 'C-36,' 'NP-15,' 'NP-38,' and 'T-17'). Rotation with tobacco and intercropping with sorghum is said to decrease the wilt problem. Other fungi include: Cercospora spp., Colletotrichum cajanae, Corticium solani, Diploidia cajani, Leveillula taurica, Macrophomina phaseoli, Phaeolus manihotis, Phoma cajani, Phyllosticta cajani, Phytophthora sp., Rhizoctonia bataticola, Rosellinia sp., Sclerotium rolfsii, and Uredo cajani (rust). So far, economic damages by these have been small or negligible, but rust is locally of some importance. Pigeon pea is also attacked by the bacterium Xanthomonas cajani and the sterility mosaic and yellow mosaic viruses [8]. Fungi reported from seeds of Peageon pea are Alternaria sp., Aspergillus sp., Colletotrichum lagenarium, Coleophoma empetri, Fusarium equiseti, Macrophomina phaseolina, Myrothecium roridum, Rhizoctonia solani, Rhizopus sp., and Sclerotium rolfsii [14, 15, 16, 17]. The present study focused on isolation of seed borne fungi of Peageon pea using Blotter paper and Agar plate method.

2. Materials and Methods

Two hundred seeds from each lot (250 g) were collected from seed market were selected for isolations by using standard blotter paper and agar plate method [12]. From the samples, twenty seeds were placed on three layers of moistened blotter paper in a 9 cm diameter petridish without any pretreatment, while for internal seed-borne fungi, seeds pretreated with 0.1% HgCl2. Same procedure was repeated in case of agar plate method. All the petridishes were incubated at $22 + 1^{\circ}$ C for eight days. After eight days, these plates were examined directly under stereoscopic microscope. The surface sterilized seeds were nicked with a pointed needle to find out the actual location of mycoflora associated with different seed components. The fungal growth appearing on seeds was identified with the help of colony color, sporulation type and shape of spores, up to species level with the help of compound microscope and relevant literature [1, 4,9,20, 22]

3. Result and Discussion

Using blotter paper method and agar plate method, total 15 genera and 32 species were isolated from 10 samples of Pigeon pea collected from Seed Market.

Of the fungi isolated 32 viz., Alternaria alternata, A.citri, Aureobasidium pullulans, Aspergillus flavus, A. fumigatous, A. niger, A. tamarii, A. terreus, A. wentii, Cladosporium cladosporoidae, C. oxysporum, C. sphaerospermum, Drechslera australiensis, Fusarium moniliformae, F. solani, oxysporum, Nigrospora oryzae FPenicillium purpreogenum, P.notatum, Rhizoctonia solani, Rhizopus suinus, Scytalidium sp., Stemphyllium sarciniformii, Trichoderma atroviride, T. flavofuscum, T. harzianum, and reported on seeds of pigeon pea. T. polysporum were External infestation was high on both the blotter paper and agar plate method respectively, which was reduced in case of pre treatment of seeds with HgCl2, (Table I, Plate no.1). Similar results were also obtained by [7], who concluded that by chlorine treatment the infestation of saprophytic fungi could be reduced. Seed surface disinfection with HgCl2 usually suppresses the growth of saprophytic and other superficial fast growing fungi [3]. Only F. oxysporum and F.solani were recorded from embryo of seeds at low frequency (Table II).

Aspergillus sp., Cladosporium sp., Fusarium oxysporum, Fusarium solani and Rhizopus sp., were most frequent in pigeon pea seeds. Rhizopus sp., were consistently isolated from seeds of pigeon pea. Gliomastix, Gleosporium, *Scytalidium and Stemphyllium sarciniformii* were found associated with some discolored and ungerminated seeds and also with seeds having abnormal seedlings.

The standard blotter method yielded maximum number of fungi. Such similar results have been observed from the detection of seed borne fungi in rice [13], Cotton [3], Cajanus [6] and Sunflower [7]. [2] reported that blotter paper method was found most suitable for detection of most infectious fungi of cucurbits.

High percentage of above mentioned potential pathogens having both internal and external mode of infestation. For the production of healthy and certified quality seeds, seed health certification programme has to be followed and seed must be tested and treated with suitable seed dressing fungicides.

Plate no: 1: External and Internal Seed Mycoflora of *Cajanus cajan*, Linn.



Untreated Seeds



Treated Seeds (Hgcl₂ solution)

Fungi	Blotter Paper Method						
	Control Set			Treated Set			
	Mean	St.D	Variance	Mean	St.D	Variance	
Alternaria alternata	13.30 ± 7.37	16.75	279.22	17.74 ± 7.52	14.05	226.417	
Alternaria citri	8.97 ± 0.0	-	-	8.25 ± 0.0	-	-	
Aspergillus flavus	15.0 ± 10.0	14.14	200.00	-	-	-	
Aspergillus fumigates	23.28 ± 7.9	20.46	500.65	22.60 ± 7.83	22.15	490.751	
Aspergillus niger	13.08 ± 6.64	15.25	254.76	8.75 ± 3.75	8.49	72.125	
Aspergillus niger	4.75 ± 0.0	-	-	0.75 ± 0.0	-	-	
Aspergillus tamari	8.25 ± 0.0	-	-	2.0 ± 0.0	-	-	
Aspergillus terreus	12.36 ± 2.84	7.54	56.83	7.57 ± 2.36	6.25	39.09	
Aspergillus wentii	4.75 ± 0.0	-	-	1.25 ± 0.0	-	-	
Aureobasidium pullulas	4.25 ± 0.0	-	-	1.8 ± 0.0	-	-	
Cladosporium cladosporoidae	$16.78{\pm}0.0$	-	-	2.75 ± 0.0	-	-	
C. oxysporum	14.12 ± 8.68	27.26	452.43	9.58 ± 6.28	16.84	318.392	
C.sphaerospermum	8.45 ± 1.67	2.89	8.39	4.50 ± 1.68	3.37	11.417	
D.australiensis	6.28 ± 0.0	-	-	$2.75\pm\ 0.0$	-	-	
Fusarium equisetii	2.0 ± 1.3	2.38	5.68	1.25 ± 0.0	-	-	
F. moniliforme	3.5 ± 0.25	0.43	0.188	5.0 ± 0.72	1.25	1.563	
F. oxysporum	$13.24\ \pm 0.0$	-	-	8.58 ± 4.77	8.27	68.396	
F.solani	5.25 ± 1.6	2.12	4.500	4.85 ± 1.06	3.27	5.644	
Gleosporium sp.	15.8 ± 8.75	17.52	306.932	13.68 ± 8.05	16.09	259.141	
Gliomastix sp.	-	-	-	4.33 ± 1.01	1.75	36.125	
Nigrospora oryzae	7.0 ± 4.25	8.48	72.00	2.25 ± 0.0	-	-	
Peniclillium notatum	3.25 ± 0.0	-	-	-	-	-	
P.purpurogenum	3.87 ± 1.87	-	-	$2.0\pm~0.0$	-	-	
Penicillium sp.	7.0 ± 6.0	-	-	2.5 ± 0.50	0.71	0.50	
Rhizoctonia solani	3.25 ± 0.0	2.65	1.875	2.25 ± 0.0	-	-	
Rhizopus suinus	7.0 ± 6.0	8.48	72.000	13.47 ± 4.62		213.492	
Scytalidium sp.	37.4 ± 16.38	36.6	1341.00	4.73 ± 3.07		28.563	
Stemphyllium sarciniformii	10.76 ± 7.51	10.61	112.50	2.07 ± 0.98	1.70	2.896	
Trichoderma atroviride	11.23 ± 0.0	-	-	-	-	-	
T. harzianum	1.76 ± 0.0	-	-	-	-	-	
T .flavofuscum	7.5 ± 4.25	6.01	36.125	$3.86 \hspace{0.1cm} \pm \hspace{0.1cm} 1.87 \hspace{0.1cm}$	2.65	7.031	
T. polysporum	1.75 ± 0.0	-	-	-	-	-	

Table 1: Occurrence of fungi on Pigeon pea seeds using Blotter Paper Method

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Fungi	Agar Plate Method								
	Control Set			Treated Set					
	Mean	St.D	Variance	Mean	St.D	Variance			
Alternaria alternata	16.10 ± 6.37	10.95	120.021	15.07 ± 5.32	9.22	85.136			
Alternaria citri	4.0± 3.23	4.54	21.124	3.13 ± 1.86	2.64	7.031			
Aspergillus flavus	1.5 ± 0.0	-	-	2.0 ± 0.0	-	-			
Aspergillus fumigates	19.10 ± 6.91	19.56	382.182	19.0 ± 7.68	21.70	472.00			
Aspergillus niger	9.05 ± 3.70	8.29	68.835	9.10 ± 3.59	7.19	51.76			
Aspergillus niger	3.5 ± 0.5	-	-	3.26 ± 0.0	-	-			
Aspergillus tamari	4.73 ± 0.0	-	-	7.43 ± 2.05	-	-			
Aspergillus terreus	9.80 ± 1.84	5.03	25.268	0.5 ± 0.0	5.43	29.451			
Aspergillus wentii	0.5 ± 0.0	-	-	5.0 ± 0.0	-	-			
Aureobasidium pullulas	$3.\ 35\pm2.86$	4.06	16.531	2.76 ± 0.0	-	-			
Cladosporium cladosporoidae	16.78 ± 0.0	-	-	12.70 ± 9.10	-	-			
C. oxysporum	15.42 ± 10.50	23.61	557.394	3.82 ± 0.88	20.34	414.418			
C.sphaerospermum	5.55 ± 1.87	3.76	14.182	-	1.51	2.271			
D.australiensis	-	-	-	$2.75\pm~0.0$	-	-			
Fusarium equisetii	1.24 ± 0.0	-	-	3.0 ± 0.25	-	-			
F. moniliforme	2.51 ± 0.35	0.66	0.437	8.66 ± 4.22	0.35	0.125			
F. oxysporum	5.57 ± 4.22	7.34	53.895	7.48 ± 1.47	7.31	53.521			
F.solani	4.21 ± 1.33	2.92	8.544	18.31 ± 9.68	3.24	10.571			
Gleosporium sp.	14.31 ± 7.15	14.30	204.599	5.15 ± 1.77	19.36	375.474			
Gliomastix sp.	$2.00\ \pm 0.58$	1.01	1.021	3.37 ± 2.87	3.95	15.675			
Nigrospora oryzae	4.50 ± 2.51	3.52	12.500	8.21 ± 2.79	4.00	16.531			
Peniclillium notatum	7.25 ± 2.43	6.43	41.425	7.75 ± 3.22	7.40	54.780			
P.purpurogenum	6.18 ± 2.47	4.95	24.556	3.81 ± 0.74	6.44	41.583			
Penicillium sp.	3.62 ± 1.37	1.95	3.780	3.81 ± 1.72	1.07	1.125			
Rhizoctonia solani	4.05 ± 2.26	3.93	15.437	-	3.44	11.891			
Rhizopus suinus	-	-	-	2.83 ± 1.72	-	-			
Scytalidium sp.	4.58 ± 3.71	6.43	41.395	8.81 ± 3.98	2.98	8.896			
Stemphyllium sarciniformii	15.46 ± 6.24	12.47	155.901	1.91 ± 0.44	7.96	63.432			
Trichoderma atroviride	7.68 ± 4.81	9.64	92.891	1.91 ± 0.44	0.763	0.582			
T. harzianum	3.41 ± 1.96	3.40	11.582	5.75 ± 2.29	3.968	15.750			
T .flavofuscum	7.5 ± 4.25	6.01	36.125	3.86 ± 1.87	2.65	7.031			
T. polysporum	$1.75\ \pm 0.0$	-	-	-	-	-			

Table 2: Occurrence of fungi on Pigeon pea seeds using Agar Plate Method

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