





the number of corn pests were significantly decreased. *Chilo agamemn* infestation decreased to 20±3.3 and 25±3.1 individuals after 90 days during season, 2014 and 2013 as compared to 98±9.3 and 99±9.3 individuals in the control plots in both two seasons. The obtained results are similar to other studies carried out by Castillo *et al.* (2000) and Espin *et al.* (1989) on their work on *C. capitata*.

The yield loss found in table (4) which showed that the weight of corn in the field during season 2013, 4089±82.80 and 4999±66.73 after treatments with *Nano-M. flavoviride* and *Nano-I. fumosorosea* respectively as compared to 2611±33.80 in the control. the percentage of yield loss 47% in the control and 0.1% in the plots treated with *Nano-I. fumosorosea*.

During season 2014 the results show that the weight of corn was significantly decreased to in the control as compared to and in plots treated with *Nano-I. fumosorosea* and *Nano-M. flavoviride*., respectively. The percentage of yield loss are 0.02% and 52% in plots treated with *Nano-I. fumosorosea* and in the control (Table 4).

Figure (1 & 2) show that the percentage of infestations were significantly decreased during both two seasons

These results are also agree with Sabbour & Shadia Abd El-Aziz (2002 and 2010) and Shadia Abdel Aziz & Nofel (1998), who proved that the application with bioinsecticides increased the yield and decreased the infestation with insect pests. Also, results were in accordance with Castillo *et al.* (2000) who reported that the virulence of *B. bassiana* against *C. capitata* ranged between 8 to 30% and decrease the infestation among the olive fruits. Espin *et al.* (1989) recorded that *C. capitata* mortality ranged between 69 and 78% after bioinsecticides treatments.

The results were matched with Abdel-Rahman & Abdel-Mallek (2001), Abdel-Rahman (2001) and Abdel-Rahman *et al.* (2004), when they controlled cereal aphids with entomopathogenic fungi. They found that the infestation was reduced after fungi applications under laboratory and field conditions. Sabbour & Sahab (2005, 2007), Tanda and Kaya (1993) and Sahab and Sabbour (2011) found that the fungi reduced insect infestations of cabbage and tomato pests under laboratory and field conditions.

## References

- [1] Abbott, W.W. (1925). A method of computing the effectiveness of an insecticide. J. Econ. Entomol 18: 265-267.
- [2] Abdel-Rahman, M.A.A.; Abdel-Mallek, A.Y. and Hamam, G.A. (2006). Comparative abundance of entomopathogenic fungi of cereal aphids in Assiut. Egypt. J. Boil. Pest Cont., 16: 39-43.
- [3] Finney, D.J. (1971). Probit Analysis, Cambridge: Cambridge University Press.
- [4] Abdel-Rahman, M.A.A. (2001). Seasonal prevalence of entomo-pathogenic fungi attacking cereal aphids infesting wheat in southern Egypt. Inter. Symposium. Agric. Agadir-Morocco, 7-10: 381-389.
- [5] Abdel-Rahman, M.A.A. and Abdel-Mallek, A.Y. (2001). Paramilitary records on entomopathogenic fungi attacking cereal aphids infesting wheat plants in southern Egypt. First Conference for safe Alternatives to pesticides for pest managements, Assiut: 183-190.
- [6] Abdel-Rahman, M.A.A.; Abdel-Mallek, A.Y.; Omar, S.A. and Hamam, A.H. (2004). Natural occurrence of entomopathogenic fungi on cereal aphids at Assiut. A comparison study between field and laboratory observations. Egypt. J. Boil. Sci., 14: 107-112.
- [7] Castillo, M.A.; Moya, P.; Hernandez, E. and Primo-Yufera, E. (2000). Susceptibility of *Ceratitiss capitata* Wiedenmann (Diptera: Tephritidae) to entomopathogenic fungi and their extract. Biol. Cont. 19: 274-282.
- [8] El-Husseini, M.M.; Shahira, S.M.; Amal, A.M.; El-Zoghby, A.; Sahar S. Ali, Naglaa, A.M. Omar; Agamy, E.A.; Abou Bakr, H.E.; Nada, M.S.; Sherin Tamer; Kamal, H.M. and Ibrahim, A.M. (2004). Isolation, Production and use of entomopathogenic fungi for controlling the sugar beet insect pests in Egypt. Egypt. J. Biol. Pest Control. 14(1): 265-275.
- [9] Espin, G.A. T. laghi De .S.M.; Messias, C.L. and Pie-Drabuena, A.E. (1989). Pathogenicidad de *Metarhizium anisopliaenas* diferentes fases de desenvolvimento de *Ceratitiss capitata* (Wied.) (Diptera: Tephritidae). Revista Brasileira de Entomologia, 33: 17-23.
- [10] Finney, D.J. (1971). Probit Analysis, Cambridge: Cambridge University Press.
- [11] Leiderer, P. and Dekorsy, T.. (2008). Interactions of nanoparticles and surfaces Tag der mÄundlichenPrÄaufung: 25. April.<http://www.ub.unikonstanz.de/kops/volltexte/2008/5387/>;
- [12] [http:// nbn-resolving.de/ urn:nbn:de:bsz:352-opus-53877](http://nbn-resolving.de/urn:nbn:de:bsz:352-opus-53877).
- [13] Qiao, Meihua; Daniel E. Snyder, Jeffery Meyer, Alan G. Zimmerman, Meihau Qiao, Sonya J. Gissendanner, Larry R. Cruthers, Robyn L. Slone, Davide R. Young (12 September 2007). "Preliminary Studies on the effectiveness of the novel pulicide, spinosad, for the treatment and control of fleas on dogs". Veterinary Parasitology: 345–351. Retrieved 3 May 2012.
- [14] Rice, R.E. (2000). Bionomics of the olive fruit fly *Bactrocera (Dacus) oleae*. Univ. of California Plant Prot. Quart, 10:-1-5.
- [15] Roberts, D.W. and Humber, R.A. (1981). Entomogenous fungi In Biology of Conidial Fungi (G.T. Cole and W.B. Kendrick, eds), vol. 2, pp. 201-236. Academic Press, New York.
- [16] Rombach, M.C.; Aguda, R.M. and Robert, D.W. (1988). Production of *Beauveria bassiana* in different liquid media and subsequent conditions mycelium. Entomo., 33: 315-234.
- [17] Sabbour, M.M. and Shadia E. Abed El-Aziz (2002). Efficacy of some botanical oils formulated with microbial agents against the cotton leafworm and greasy cutworm attacking cotton plants. Bull. Ent. Soc. Egypt. ser. 28, 2001-2002: 135-151.
- [18] Sabbour, M.M. and Sahab, A.F. (2005). Efficacy of some microbial control agents against cabbage pests in Egypt. Pak. J. Biol. Sci. 8: 1351-1356.

- [19] Sabbour, M.M. and Sahab, A.F. (2007). Efficacy of some microbial control agents against *Agrotis ipsilon* and *Heliothis armigera* in Egypt. Bull. N.R.C. Egypt. 13
- [20] Sabbour, M.M. and Shadia, E. Abd-El-Aziz (2010). Efficacy of some bioinsecticides against *Bruchidius incarnatus* (BOH.) (Coleoptera: Bruchidae) Infestation during storage. J. Plant Prot. Res. 50 (1): 28-34.
- [21] Sahab, A.F. and Sabbour, M.M. (2011). Virulence of four entomo-pathogenic fungi on some cotton pests with especial reference to impact of some pesticides, nutritional and environmental factors on fungal growth. Egypt. J. Boil. Pest Cont., 21 (1): 61-67.
- [22] Sameh, A. Moustafa; Ahmed, E. Abd El-Mageed; Mostafa, M. El-Metwally and Nabil, M. Ghanim (2009). Efficacy of Spinosad, Lufenuron and Malathion against olive fruit fly, *Bactrocera oleae* (Gmelin) (Diptera: Tephritida e) Egypt. Acad. J. biolog. Sci., 2 (2): 171- 178.
- [24] Shadia, E. Abdel Aziz and Nofel, M.A. (1998). The efficacy of bacteria, fungi and natural products in baits against the greasy cutworm *Agrotis ipsilon* (Hufn.) (*Lepidoptera: Noctuidae*) in Egypt. J. Egypt. Ger. Soc. Zool., 27. Ent. 129-139.
- [25] Tanda, Y. and Kaya, H.K. (1993). Insect Pathology. Academic Press, San Diego, CA, USA.

**Table 1:** Effect of nano-*M. flavoviride* against the target insect pests larvae under laboratory conditions

Insects	LC <sub>50</sub>	slope	Variance	95% confidence limits
<i>Ostrinia nubilalis</i>	88x10 <sup>4</sup>	0.1	1.01	69-129
<i>Sesamia cretica</i>	76x10 <sup>4</sup>	0.2	1.00	65-140
<i>Chilo agamemnon</i>	106x10 <sup>4</sup>	0.1	1.03	119-187

**Table 2:** Effect of nano-*Isaria fumosorosea* against target insect pests under laboratory conditions

Insects	LC <sub>50</sub>	slope	Variance	95% confidence limits
<i>Ostrinia nubilalis</i>	56 x10 <sup>4</sup>	1.01	0.02	41-76
<i>Sesamia cretica</i>	68 x10 <sup>4</sup>	0.10	1.01	54-87
<i>Chilo agamemnon</i>	95 x10 <sup>4</sup>	0.10	1.01	85-109

**Table 3:** Effect of different treatments on the three target insect pests under field conditions

Post 1 <sup>st</sup> application date	Treatments	Number of infestation (Mean±SE) during the two seasons					
		<i>Ostrinia nubilalis</i>		<i>Sesamia cretica</i>		<i>Chilo agamemnon</i>	
		2013	2014	2013	2014	2013	2014
20	Control	44±6.2	68±9.8	61±2.5	79±9.7	69±9.9	78±9.9
50		69±8.9	76±9.1	79±3.4	88±8.9	85±7.9	91±9.3
90		79±8.3	89±9.1	88±5.1	97±2.9	98±9.9	99±9.9
20	<i>Nano-M. flavoviride</i>	12±0.1	14±0.1	16±4.1	15±1.0	16±0.3	17±0.2
50		16±0.2	15±0.1	20±0.7 24±0.4	23±0.2	19±0.4	19±0.4
90		20±1.0	18±0.2		19±0.3	20±0.3	20±0.1
20	<i>Nano-I. fumosorosea</i>	19±3.1	20±0.1 21±1.0	21±2.1	21±2.0	21±3.1	21±2.1
50		20±1.2 22±0.2	22±1.0	22±3.1 25±1.1	23±0.2	24±2.1 25±1.1	22±1.0
90					26±2.1		24±1.3
F-value		22.2	37.3	34.1	28.8	30.1	24.8
LSD at 5%		11.2	14.4	13.8	15.0	16.8	16.7

**Table 4:** Assessments of damage caused in corn crop field after the nano- entomopathogenic fungi treatment

Treatments	Season 2013		Season 2014	
	Weight of corn crop (kg/ feddan)	% yield loss	Weight of corn crop (kg/feddan)	yield loss %
<i>Nano-M. flavoviride</i>	4089±82.80	0.1	4990±69.83	0.02
<i>Nano-I. fumosorosea</i> Control	4999±66.73	-	5099±49.74	-
	2611±33.80	47	2400±33.10	52
F-value	33.6		32.9	
LSD at 5%	126.7		126.5	

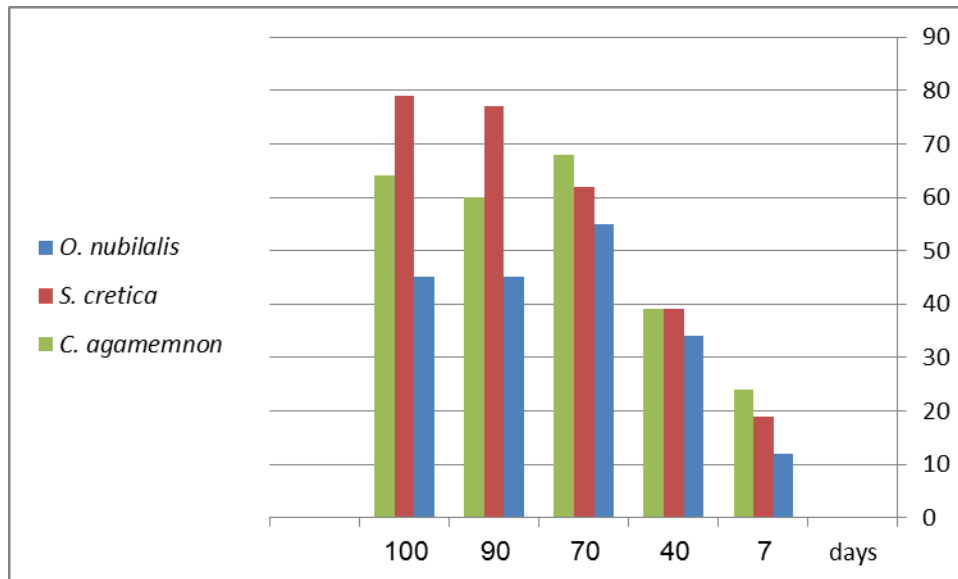


Figure 1: Effect of the nano- fungus *I. fumosorosea* under field conditions

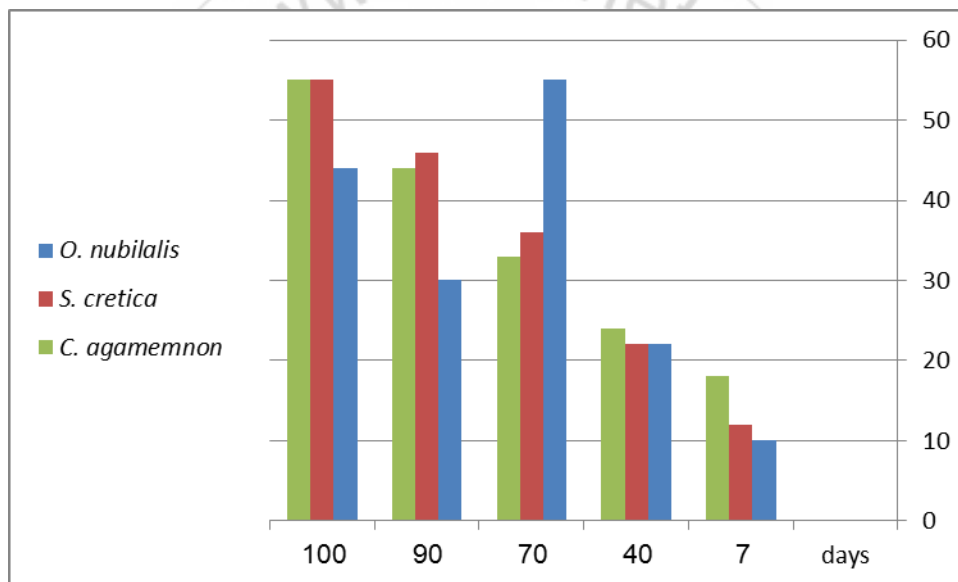


Figure 2: Effect of the nano- fungus *M. flavoviride* under field conditions