Ameliorative Effects of Methanolic Extract of Sennaalata leaves on Streptozotocin (Stz)-Induced Oxidative Stress and Hepatotoxicity in Adult Wistar Albinorats

Akinwale A.O1, Adedosu O.T2, Kehinde B.D2, Ibiwunne G.J1

1, 2Department of Science Laboratory Technology, Faculty of Pure and Applied Sciences, Ladoke Akintola University of Technology Ogbomoso, Nigeria

Abstract: Sennaalata leaf is widely available in the tropics and has very important applications in folkloric medicine. It contains several biological properties and antioxidant properties. In the northern part of Nigeria, particularly in Adamawa and Taraba States, the root, stem and leaves are used by practitioners of herbal medicines to treat burns, skin and wound infections, diarrhea, gastrointestinal and upper respiratory tract infections. Objectives: In this work, liver morphology was studied, markers of hepatic oxidative stress and some liver enzymes depicted normal hepatocytes with normal morphology and without necrosis. Conclusion: significantly decreased (P <0.05) the activities of ALT and AST when compared with STZ-induced untreated group (group B). The experimental type 1 diabetes in animals (Punithavathi et al., 2009). Several hypothesis such as oxidation of glucose, the nonenzymatic and progressive glycation of proteins with consequently increased formation of glucose-derived advanced glycation end products (AGEs) have been proposed to explain the genesis of free radicals in diabetes (Booth et al., 1997; Vlassara Palace, 2001). The increased production of reactive oxygen species results in oxidative stress, a state where the concentration of reactive intermediates(such as reactive oxygen species, ROS) overwhelms the concentration of antioxidants (Maxwell et al., 1997; Nwaogu and Onyeze, 2010). Oxidative stress is the result of a redox imbalance between the generation of ROS and the compensatory response from the endogenous antioxidant network. Living organisms have evolved a highly complicated and robust defense system comprizing enzymes that can degrad functional antioxidants in ROS.

Keywords: Oxidative stress, Streptozotocin, Sennaalata leaves; hepatoprotective effects, wistar albino rats.

1. Introduction

The liver is the principal organ of metabolism and plays vital roles in many body processes most especially detoxification of chemical compounds (Sunmonu and Oloyede, 2007). The liver is the first processing and purifying station for chemical compounds (Sunmonu and Oloyede, 2007). The increased production of reactive oxygen species results in oxidative stress, a state where the concentration of reactive intermediates(such as reactive oxygen species, ROS) overwhelms the concentration of antioxidants (Maxwell et al., 1997; Nwaogu and Onyeze, 2010). Oxidative stress is the result of a redox imbalance between the generation of ROS and the compensatory response from the endogenous antioxidant network. Living organisms have evolved a highly complicated and robust defense system comprizing enzymes that can degrad functional antioxidants in ROS.

STZ is the direct result of irreversible damage to the pancreatic beta cells resulting in degranulation and loss of capacity to secrete insulin (Guet al., 1997). Hyperglycemia in diabetes has been associated with increased formation of reactive oxygen species (ROS) and oxidative damage to tissue compounds (Nakahae et al., 2009). Several hypothesis such as oxidation of glucose, the nonenzymatic and progressive glycation of proteins with consequently increased formation of glucose-derived advanced glycation end products (AGEs) have been proposed to explain the genesis of free radicals in diabetes (Booth et al., 1997; Vlassara Palace, 2001). The increased production of reactive oxygen species results in oxidative stress, a state where the concentration of reactive intermediates(such as reactive oxygen species, ROS) overwhelms the concentration of antioxidants (Maxwell et al., 1997; Nwaogu and Onyeze, 2010). Oxidative stress is the result of a redox imbalance between the generation of ROS and the compensatory response from the endogenous antioxidant network. Living organisms have evolved a highly complicated and robust defense system comprizing enzymes that can degrad functional antioxidants in ROS.
defense system and the body acts against free radical-induced oxidative stress involved by different defense mechanism like preventative mechanisms, repair mechanisms, physical defenses and antioxidant defenses (Valko et al., 2007). Under normal circumstances, sufficient amounts of reactive oxygen species are removed by the antioxidant defense systems including enzymes such as superoxide dismutase (SOD), catalase and glutathione peroxidase (Mckee and Mckee, 1999). However, in diabetic condition, the prevailing hyperglycaemia and the consequent excess reactive oxygen species generation can either decrease the activity of these antioxidant enzymes or deplete their metabolites. The decrease in antioxidant enzyme activity under diabetic conditions could be due to glycation of these enzymes, which occurred at persistently elevated blood glucose levels (Taniguchi, 1992). Glycation of SOD reduces its activity, leading to the insufficient dismutation of superoxide anions (O$_2^-$) (Taniguchi, 1992; Majithiya and Balaram 2005; Ravi et al., 2004). The excess formation of ROS, due to a depleted endogenous antioxidant system (Young et al., 1995; Baydas et al., 2002), for instance, leads to a decrease in reduced glutathione (GSH) concentration and SOD activity, and an increase in lipid peroxidation (Tavares de Almeida et al., 2012). Moreover, chronic hyperglycaemia induces carbonyl stress, which in turn can lead to increased lipid peroxidation (Bayanes and Thrope, 1999). The increased concentration of lipid peroxidation worsens oxidative damage by increasing peroxide radicals and hydroxyl radicals (Levy et al., 1999). The increased lipid peroxidation in the plasma and tissues of diabetic animals may be due to the observed remarkable increase in the concentration of MDA as a main product of lipid peroxidation in the plasma and liver (Vijayakumar et al., 2006). Excessive free radical generation has been linked with abnormal alterations in the dynamic properties of cellular membranes (Lin et al., 2000). Free radicals can interact and disrupt the hepatic cell membranes thereby causing the hepatic marker enzymes such as alanine aminotransferase (ALT) and aspartate aminotransferase (AST) to leak out and increase their serum concentrations (Patrick-Iwuanyanwu et al., 2011).

The use of bioactive plant-derived compounds is on the rise, because the main preoccupation with the use of synthetic drugs is the side effects which can be even more dangerous than the diseases they claim to cure (Kamboj, 2000). In contrast, plant derived medicines are based upon the premise that they contain natural substances that can promote health and alleviate illness and proved to be safe, better patient tolerance, relatively less expensive and globally competitive. (Kamboj, 2000). *Senna alata* (L.) Roxb belongs to the Fabaceae family (subfamily Caesalpinioideae) and commonly known as candle bush, with reference to the shape of its inflorescences, or ringworm tree because of a traditional Use (Abo et al., 2008). It is commonly referred to as “Asuwonoyinbo” by the Yoruba ethnic group in Southwestern Nigeria (Awale et al., 2004).

It is widely available in the tropics and has very important applications in folkloric medicine (Rai and Curr, 1987). In the northern part of Nigeria, particularly in Adamawa and Taraba States, the root, stem and leaves are used by practitioners of herbal medicines to treat burns, skin and wound infections, diarrhea, gastrointestinal and upper respiratory tract infections (El-mahoodet al., 2008). Recent researches have shown that the antioxidants of plant origin with free-radical scavenging properties could have great importance as therapeutic agents in several diseases caused due to oxidative stress (Ramchoun et al., 2009). Preliminary phytochemical study showed the presence of tannins, phlobatannins, alkaloids, anthraquinones, flavonoids, cardiac glycosides and saponins in the chloroform extract of leaves of *Senna alata*. However, flavonoids (Wegner and Fintelmann, 1999) alkaloids (Manjunath, 2006), saponins (Trans et al., 2001) and glycoside (Vijayane et al., 2003) have been reported to have hepatoprotective activity. Moreover, some researchers have found antioxidant compounds such vitamin E (Tavares de Almeida et al., 2012) and quercetin (Coskun et al., 2005), and antioxidant- containing plant *Eucalyptus globules* (Nakahae et al., 1999) to prevent and ameliorate streptozotocin-induced oxidative stress and hepatotoxicity. The aim of this present study was therefore to investigate the possible protective effect of *Senna alata* on STZ-induced oxidative stress and hepatotoxicity in wistar rats.

2. Materials and Methods

**Plant materials**

Fresh leaves of *Senna alata* were collected from under G area, Ogbomoso, Oyo state. The plant was authenticated at the department of Pure and Applied Biology, Ladoke Akintola University of Technology, Ogbomoso, Oyo state by Dr.Ogunkunle. The leaves were rinsed severally with clean tap water to remove dust particles and debris and thereafter allowed to completely drain. The collected leaves were then chopped into bits on a chopping board and air dried at room temperature 25°C -30°C for three weeks before taking to the experimental site.

**Preparation of methanolic extract of Senna alata leaves**

Eight hundred and twenty grams of air-dried and pulverized leaves of *Senna alata* was immersed in 5L of methanol for five days to ensure sufficient extraction of the active components. The suspension was filtered and the solvent evaporated to dryness in water bath. The stock was then prepared from the residue and the corresponding doses of 100mg/kg and 300mg/kg body weight were calculated and administered to the experimental animals as required.

**Animal management**

A total number of 40 malewistar rats weighing between 190 -245 g were used for this study. The rats were acclimatized to the experimental room having temperature of 25°C, controlled humidity conditions (65°C) and 12:12h light; dark cycle for two weeks. The experimental animals were housed in standard plastic cages, fed with standard diet (pelletized growers mash) obtained from Bovajay Feed Mill at Orita Naira in Ogbomoso and water ad libitum.

**Induction of Streptozotocin(STZ)**

Experimental Streptozotocin (STZ) was induced by intraperitoneal injection of 60mg/kg STZ freshly dissolved in 0.1M Sodium Citrate at pH 4.5 buffered at 4.5. Hyperglycemia was confirmed two days after injection by measuring the tail vein blood glucose level with an Accu-
Check Sensor Comfort Glucometer (Roche, Mexico City). Only the animals with fasting blood glucose level ≥ 200mg/dl were considered diabetic.

**Experimental design**
A total of 40 wistar rats were used for this study. The animals were randomly divided into four groups of 10 animals per group. The experimental period was three weeks beginning after the induction of STZ-induced oxidative stress. Group A: normal control untreated rats. Group B: STZ-induced untreated group (60mg/kg body weight). Group C and D were STZ-induced rats given methanolic extract of *Senna alata* leaves (150mg/kg and 300mg/kg body weight) daily using an intragastric tube for three weeks (21 days).

**Sample preparation**
Each clothed blood sample was centrifuged at 2000 revolution per minute for 20 minutes to obtain the serum. The supernatant (serum) was siphoned using micropipette.

**Tissue Homogenates**
The liver was cut into pieces and homogenized in equal volume of chilled 10mM Tris/HC1 sucrose buffer pH 7.4 in a mortar and pestle.

**Serum Biochemical Assay**
Serum alanine aminotransferase (ALT) and aspartate aminotransferase (AST) activities were determined by the colorimetric method of Reitman and Frankel as described by Ochei and Kolhatkar (2005).

**Estimation of SOD, MDA and GSH in the Liver**
The activity of superoxide dismutase (SOD) was determined by using the method of Misra and Fridovich (1972). Lipid peroxidation was determined based on the principle of Varshney and Kale (1990). Estimation of lipid peroxidation was based on the reaction of malondialdehyde (MDA) with thiobarbituric acid (TBA) forming a MDA-TBAR adduct that absorbed strongly at 532nm. Reduced glutathione (GSH) level in the liver was assayed following the method of Ellman (1959), modified by Hissein and Hill (1973).

**Tissue processing**
The tissues were allowed to fix in 10% formol saline for 48 hours, tissues were grossed and cut into smaller pieces of 3mm thick in prelabelled tissue cassette. They were processed using Automatic tissue processor (LEICA TP1020) where they passed through various reagents including Alcohol (of various concentrations starting from 70%, 80%, 90%, and two 100% or absolute alcohol) for dehydrations, two changes of xylene and three changes of molten paraffin wax set at 65 degree centigrade. The processing time was 12 hours (Carleton, 1967).

**Statistical analysis**
Data were expressed using Graph pad prism 6. Data were expressed as Mean ± Standard error of mean (Mean ± S.E.M). Values were compared using one-way analysis of variance (ANOVA) followed by student t-test. *P*-Values less than 0.05 were taken to be statistically significant and insignificant at *P* > 0.05.

**3. Results**
Table 1 and figure 1 showed that malondialdehyde concentration in the liver was significantly higher in the STZ – induced untreated group (group B) when compared with the normal control (group A) at *P* < 0.05. However, treatment with methanolic extract of *Senna alata* leaves significantly reduced the MDA levels in both group C (STZ- induced +ML) & group D (STZ-induced +MH) with higher reduction in group D (STZ-induced +MH).

The result showed in table 1 and figure 1 also indicated a significant decrease in GSH level in the STZ-induced untreated group as compared with the normal control group (group A) at *P* < 0.05. However, treatment with methanolic extract of *Senna alata* leaves significantly increased the reduced glutathione levels in dose dependent manner respectively.

Table 1 also represented the results of the oxidative status of the rats. The activity of superoxide dismutase was significantly (*P* < 0.05) lowered in Streptozotocin induced untreated group (Group B) when compared with the normal control group A. However, treatment with 150mg/kg and 300mg/kg of methanolic extract of *Senna alata* significantly increased superoxide dismutase activity in dose dependent manners in group C (STZ-induced +ML) and group D (STZ-induced +MH) respectively.

The results from table 2 and figure 2 below showed significantly (*P* < 0.05) higher activity of serum biomarker enzymes, Alanine aminotransferase (ALT) and Aspartate aminotransferase (AST) in the STZ-induced untreated group (Group B) when compared with the normal control group (Group A). However, treatment of rats with methanolic extract of *Senna alata* leaves significantly (*P* < 0.05) reduced the activity of ALT and AST when compared with the STZ-induced untreated group. The reduction in the activity of ALT was not dose dependent.

**Table 1:** Effect of methanolic extract of *Senna alata* leaves on malondialdehyde (MDA) level, reduced glutathione(GSH) level and Superoxide dismutase activity of the streptozotocin induced oxidative stress in rats.

<table>
<thead>
<tr>
<th>PARAMETERS</th>
<th>GROUP A (Control)</th>
<th>GROUP B (STZ-induced untreated)</th>
<th>GROUP C (STZ-induced+ML)</th>
<th>GROUP D (STZ-induced+MH)</th>
</tr>
</thead>
<tbody>
<tr>
<td>MDA(mmol/gtissue)</td>
<td>7.08±0.19</td>
<td>13.82±1.08</td>
<td>11.05±0.22</td>
<td>9.61±0.10</td>
</tr>
<tr>
<td>GSH(µmol/gtissue)</td>
<td>23.70±0.58</td>
<td>11.29±0.31</td>
<td>13.71±0.33</td>
<td>19.92±0.40</td>
</tr>
<tr>
<td>SOD(µmol/min)</td>
<td>88.80±3.07</td>
<td>23.14±1.38</td>
<td>30.90±0.90</td>
<td>56.90±1.79</td>
</tr>
</tbody>
</table>

ML = methanolic low dose & MH = methanolic high dose. Values are expressed as Mean ± S.E.M (Standard Error of Mean); MDA, GSH and SOD in liver of STZ-induced oxidative stress in wistar rats using student t- test.*Represents
significant increase at $P<0.05$ when compared with Group A (control), b Represents significant decrease at $P<0.05$ when compared with Group B (STZ- induced).

![Figure 1](image1.png)

**Figure 1:** Effect of methanolic extract of *Senna alata* leaves on MDA and GSH Concentration and SOD activity of STZ-induced oxidative stress in Wistar rats

**Table 2:** Effect of methanolic extract of *Senna alata* leaves on the activity of Alanine aminotransferase (ALT) and Aspartate aminotransferase (AST) in Serum of wistar rats after treatment.

<table>
<thead>
<tr>
<th>PARAMETERS</th>
<th>GROUP A (Control)</th>
<th>GROUP B (STZ-induced untreated)</th>
<th>GROUP C (STZ-induced+ ML)</th>
<th>GROUP D (STZ-induced+ MH)</th>
</tr>
</thead>
<tbody>
<tr>
<td>ALT (U/L)</td>
<td>55.71±8.29</td>
<td>198.60±10.30</td>
<td>128.30±1.18</td>
<td>60.90±3.29</td>
</tr>
<tr>
<td>AST (U/L)</td>
<td>22.53±2.35</td>
<td>55.61±2.42</td>
<td>47.88±0.74</td>
<td>41.88±0.79</td>
</tr>
</tbody>
</table>

ML = methanolic low dose, MH = methanolic high dose. Values are expressed as Mean ± S.E.M (Standard error of Mean) on serum analysis of Alanine Aminotransferase (ALT) and Aspartate Aminotransferase (AST) in liver of STZ – induced oxidative stress in wistar rats using student t- test. a Represents significant decrease at $P<0.05$ when compared with Group A (control), b Represents significant increase at $P<0.05$ when compared with Group B (STZ- induced).

![Figure 2](image2.png)

**Figure 2:** Effect of methanolic extract of *Senna alata* leaves on serum ALT and AST activities of STZ-induced oxidative stress in Wistar rats.
previous researchers on the effect of STZ on the oxidative increase in malondialdehyde (MDA) concentration compared sections of group C induced with 60mg/kg STZ and treated with 150mg/kg of methanolic extract of Sennaalata leaves showed normal liver microarchitecture, central vein(green arrow), sinusoids(red arrow), hepatocytes(blue arrow), no haemorrhage and there is no infiltration of inflammatory cells. A photomicrograph of liver sections of group B induced with 60mg/kg STZ showed perportal fatty infiltration(PFI) with focal necrosis of hepatocytes(blue arrow), enlarged sinusoids(red arrow) and central vein(green arrow). A photomicrograph of liver sections of group C induced with 60mg/kg STZ and treated with 150mg/kg of methanolic extract of Sennaalata leaves showed normal central vein(green arrow), reduced distortion in the histology of the hepatocytes(blue arrow), enlarged sinusoids with perivenous fatty infiltration (red arrow). A photomicrograph of liver sections of group D induced with 60mg/kg STZ and treated with 300mg/kg of methanolic extract of Sennaalata leaves showed normal liver microarchitecture featured hepatocytes(blue arrow), sinusoids(red arrow) and central vein(green arrow) no haemorrhage and there is no infiltration of inflammatory cells compared with normal control group.

STZ: streptozotocin; ML: methanolic low dose; MH: methanolic high dose

4. Discussion

The result obtained in this study indicated that exposure of rats to STZ resulted in significant decrease (P <0.05) in the activity of superoxide dismutase (SOD) and concentration of reduced glutathione (GSH) as well as significant (P <0.05) increase in malondialdehyde (MDA) concentration compared with the control group. This is consistent with the results of previous researchers on the effect of STZ on the oxidative status of STZ- exposed rats (Nakhaee et al., 1999; Coskun et al., 2005; Tavares de Almeida et al., 2012). The observed significant decrease (P <0.05) in the activity of superoxide dismutase (SOD) and concentration of reduced glutathione (GSH) along sides significant (P <0.05) increase in malondialdehyde (MDA) concentration indicates oxidative stress in STZ-induced rats (Ujowundet al., 2012). SOD catalyses the consumption of superoxide anion (O₂⁻) which peroxidises cell membrane. The activity of SOD in the cell is therefore a predictor of oxidative status of that cell. The significantly low SOD activity recorded in the liver of the STZ- induced untreated rats could mean the tissue was undergoing oxidative attack occasioned by superoxide anions (Table 1 and Figure 1). However, treatment with 150mg/kg and 300mg/kg of methanolic extract of Sennaalata leaves significantly increased superoxide dismutase activity in dose dependent manner in group C (STZ-induced +ML) and group D (STZ-induced +MH).

In organ and tissue damage, GSH makes up the first line of defense against free radicals resulting from the ingestion of xenobiotics (Ujowund et al., 2012). The significantly low GSH level in the Streptozotocin induced untreated group compared with the normal control group might have confirmed damage to the liver presumably by the oxidant (Table 1 and Figure 1). Nonetheless, treatment with chloroform extract of Sennaalata also significantly increased the reduced glutathione concentration in dose dependent manner. The observed increase in malondialdehyde (MDA) concentration in STZ-induced untreated group as compared to the controlindicates increased lipid peroxidation which could have resulted from depletion of GSH concentration. Again treatment with 150mg/kg and 300mg/kg of methanolic extract of Sennaalata leaves significantly reduced the MDA levels in both group C (STZ- induced +ML) and group D(STZ-induced +MH), with higher reduction in group D(STZ-induced +MH).

The observed significant increase in SOD activity and reduced glutathione concentration with corresponding significant decrease in MDA level in rats induced with STZ and treated with methanolic extract of Sennaalata leaves indicates the ameliorative effect of methanolic extract of Sennaalata leaves on STZ-induced oxidative stress and hepatotoxicity. Methanolic extract of Sennaalata leaves may have conferred protection against oxidative damage of the hepatocytes by the antioxidant activity of its phytoconstituents as the preliminary phytochemical screening of methanolic extract of Sennaalata leaves revealed the presence of antioxidants such as flavonoids, alkaloid, glycosides and saponins (Wegner and Fintelmann, 1999, Manjunath, 2006, Trans et al., 2001, Vijayan et al., 2003). Previous studies have established antioxidant compounds such as vitamin E (Tavares de Almeida et al., 2012) and quercetin (Coskun et al., 2005), and antioxidant-containing plant Eucalyptus globules (Nakhaee et al., 1999) to prevent and ameliorate streptozotocin-induced oxidative stress and hepatotoxicity. Moreover, flavonoids, alkaloids, saponins, glycosides have been reported to possess hepatoprotective activity (Wegner and Fintelmann, 1999, Manjunath, 2006, Trans et al., 2001, Vijayan et al., 2003).

Plate 1: Hepatic histology after three weeks of treatment (H&E stain)

A: Normal control; B: STZ-induced untreated; C: STZ-induced + ML D: STZ-induced + MH

In organ and tissue damage, GSH makes up the first line of defense against free radicals resulting from the ingestion of xenobiotics (Ujowund et al., 2012). The significantly low GSH level in the Streptozotocin induced untreated group compared with the normal control group might have confirmed damage to the liver presumably by the oxidant (Table 1 and Figure 1). Nonetheless, treatment with chloroform extract of Sennaalata also significantly increased the reduced glutathione concentration in dose dependent manner. The observed increase in malondialdehyde (MDA) concentration in STZ-induced untreated group as compared to the controlindicates increased lipid peroxidation which could have resulted from depletion of GSH concentration. Again treatment with 150mg/kg and 300mg/kg of methanolic extract of Sennaalata leaves significantly reduced the MDA levels in both group C (STZ- induced +ML) and group D(STZ-induced +MH), with higher reduction in group D(STZ-induced +MH).

The observed significant increase in SOD activity and reduced glutathione concentration with corresponding significant decrease in MDA level in rats induced with STZ and treated with methanolic extract of Sennaalata leaves indicates the ameliorative effect of methanolic extract of Sennaalata leaves on STZ-induced oxidative stress and hepatotoxicity. Methanolic extract of Sennaalata leaves may have conferred protection against oxidative damage of the hepatocytes by the antioxidant activity of its phytoconstituents as the preliminary phytochemical screening of methanolic extract of Sennaalata leaves revealed the presence of antioxidants such as flavonoids, alkaloid, glycosides and saponins (Wegner and Fintelmann, 1999, Manjunath, 2006, Trans et al., 2001, Vijayan et al., 2003). Previous studies have established antioxidant compounds such as vitamin E (Tavares de Almeida et al., 2012) and quercetin (Coskun et al., 2005), and antioxidant-containing plant Eucalyptus globules (Nakhaee et al., 1999) to prevent and ameliorate streptozotocin-induced oxidative stress and hepatotoxicity. Moreover, flavonoids, alkaloids, saponins, glycosides have been reported to possess hepatoprotective activity (Wegner and Fintelmann, 1999, Manjunath, 2006, Trans et al., 2001, Vijayan et al., 2003).
The result of this present study also indicated hepatic injury as evident by a significant increase (P<0.05) in the activities of serum marker enzymes namely serum alanine transaminase (ALT) and serum aspartate transaminase (AST) in the group treated with streptozotocin only when compared with the normal control (Table 2 and Figure 2). Increased levels of serum ALT and AST have been reported to be sensitive marker of hepatic damage(Ochei, Kolhatkar, 2005; Achuba and Ogwumu, 2014). This may be due to leakage from the cells through peroxidative damage of the membrane. However, the reduction in the levels of these marker enzymes in groups administered with methanolic extract (Groups C and D) is suggestive of regeneration process and repair of hepatic damage induced by streptozotocin.

Histological findings showed normal histoarchitecture in the normal control group. Photomicrograph plates of Group A (Plate 1) reflects normal hepatocytes separated by sinusoids. The central veins show normal histoarchitecture, no haemorrhage and there is no infiltration of inflammatory cells seen within the liver parenchyma. The histological section of STZ–induced untreated group showed enlarged central vein, periportal fatty infiltration (PFI) with focal necrosis of hepatocytes. The sinusoids were enlarged with fatty infiltration (Plate 1).

However, treating rats with methanolic extract of Sennaalata leaves improved the histological features of the liver cells. Although the liver of rats treated with 150mg/kg (group C) still featured some lesions seen in group B, the liver of the group treated with 300mg/kg (group D) showed normal histological features as observed in the normal control group. This suggests the antioxidative actions and hepatoprotective potential of Sennaalata leaves extract is dose-dependent and more efficient at higher doses.

5. Conclusion

This study demonstrated that methanolic extract of Sennaalata leaves through its marked antioxidant properties salvaged streptozotocin-induced oxidative stress and hepatic damage. Therefore, Sennaalata leaves can be used for the treatment of oxidative stress-induced hepatic disorders.

References


