effects of abiotic stress. Seed priming techniques have been used to increase germination, improve germination uniformity, improve seedling establishment and stimulate vegetative growth in more field crops (Ansari *et al.*, 2012; Patade *et al.*, 2011; Foti *et al.*, 2008) under stressed conditions. Also, Ansari and Sharif-Zadeh,(2012) reported that priming by salicylic acid and gibberellins have been used to increase germination characteristics in rye seeds. Also, the priming strategies enhanced activities of free radical scavenging enzymes such as CAT and APX (Ansari and Sharif-Zadeh, 2012; Rouhi *et al.*, 2012).

2. Material and Methods

The present investigation was conducted during Rabi seasons of 2012-13 and 2013-14. The investigation was carried out in field with two varieties of wheat (Triticum aestivum L.) HD 2733 (susceptible) and KRL 210 (tolerant). A field at the experiment at the research farm of the Department of Genetics and Plant Breeding, Narendra Deva University of Agriculture & Technology (Kumarganj), Faizabad (U.P.). The experiment was conducted in sodic soil with two varieties HD 2733 (susceptible) and KRL 210 (tolerant). The whole experiment was planned under Randomized block design (factorial) with three replications with six treatments. Solutions of desired concentrations of plant growth regulators (PGRs) and chemicals were prepared. After that bold and healthy seeds of HD 2733 and KRL 210 were primed in the solutions of KNO₃-3%, KCl-1%, GA₃ -150 ppm and Cycocel-500 ppm for 12 hours before sowing. For hydropriming treatment, seeds were soaked in distilled water. Non primed seeds were taken as untreated control. The seeds were dried in shade for 2 hours and sowing was done with the help of chisel in the first week of December 2012 and December 2013 at a row, spacing of 22cm at depth of 4-5cm. After 15 days of sowing, thinning was done to maintain and provide proper spacing. About 30 uniform plants of same vigour were selected as well as tagged in each plot. The data regarding germination were recorded 15 days after sowing. Various data were recorded at 30, 60 and 90 Days after sowing of seeds. Plant height was measured with the help of meter scale from the base of the shoot to tip. Plants were oven dried at 80°C for 24 hours and dry weight was taken at each stages of observation. Chlorophyll content was estimated according to the method of Arnon (1949) and expressed as mg g⁻¹ fresh weight of leaves. Starch content was measured by the method of Mc Cready (1950). Both biochemical parameters (Chlorophyll content and Starch content) were recorded at reproductive stage. The statistical analysis of experimental data was done by method described by Fisher and Yates (1949) using Randomized block. Seed germination was recorded upto 14 days after start of the experiment. Germination percentage was calculated using the following formula:

Germination (%) = $\frac{\text{Number of seeds germinated after } 14 \text{ days } x100}{\text{Number of seeds sown}}$

3. Result and Discussion

Seed priming had significant positive effect on different aspects such as seed germination, growth and biochemical

parameters. Sodicity significantly reduced the seed germination percentage irrespective of wheat variety but the effect of sodicity was more pronounced in susceptible variety HD 2733 in comparison to tolerant variety KRL 210.

Seed priming significantly enhanced the seed germination percentage in wheat irrespective of variety but the effect of priming was more prominent in tolerant variety KRL 210 in comparison to susceptible variety HD 2733 (Table 1). Maximum germination percentage was recorded in KNO₃ (3%), KCl (1%) and GA₃ (150ppm) while hydropriming influenced least on germination percentage. Faroog et al. (2006) reported that hydropriming break down seed dormancy by the activation of hydrolytic enzymes like αamylase. This increase in germination may also be due to the activity of α-amylase due to osmopriming. Amylases are key enzymes that play a vital role in hydrolyzing the seed starch reserve, thereby supplying sugars to the developing embryo. The improved germination of primed seeds was attributed to enhanced counteraction of free radicals and re-synthesis of membrane bound enzymes compared to unprimed seeds (Srinivasan and Saxena 2001). Similar findings were also reported by Golizadehet al., (2015) in Cannabis seed, Lara et al., (2014) in tomato, Ghobadi et al., (2012), Lemrasky et al. (2012) and Abbasdokht (2011) in wheat.

Table 1: Effect of seed priming with chemicals and PGRs on germination and biochemical changes of wheat varieties under sodic soil

Variety Treatments	Germination percentage	Total chlorophyll content	Starch content
Control	68.75	2.71	93.63
Hydropriming	73.08 2.76		102.06
Priming with KNO ₃	80.92 3.11		113.83
Priming with KCl	77.75	3.05	108.71
Priming with GA ₃	76.25	3.12	105.62
Priming with cycocel	74.50	3.04	103.09
SEm±	0.87	0.025	1.30
CD at 5%	2.55	0.073	3.82
HD 2733	72.08	2.84	100.74
KRL 210	78.33	3.09	108.24
SEm±	0.50	0.014	0.75
CD at 5%	1.47	0.042	2.21

In the present investigation, seed priming with chemicals, plant growth regulators and water had maintained significantly higher total chlorophyll in both tolerant and susceptible varieties (Table-1). Maximum chlorophyll content was recorded with KNO₃(3%) followed by KCl (1%), GA₃(150ppm), Cycocel (500ppm) and hydropriming as compared to untreated control. Chlorophyllase enzyme which is responsible for chlorophyll degradation, might have been inhibited by priming treatment. The Similar increase in chlorophyll content has been reported by Menon *et al* (2013), Wasif and Mohammd (2012) in *moringa oleifera*, Azooz (2009) in sorghum, Hamid *et al.*, (2008) in wheat and EI-Tayed (2005) in barley.

Seed priming significantly enhanced the starch content in both the wheat varieties over control(Table 1). The maximum starch content was found with KNO₃ followed by KCl, GA₃, cycocel and hydropriming as compared to untreated control. Increase in starch content might be due to

International Journal of Science and Research (IJSR)

ISSN (Online): 2319-7064

Index Copernicus Value (2013): 6.14 | Impact Factor (2013): 4.438

induced hydrolysis of reserve polysaccharide or rapid utilization of total soluble starch. These results are corroborated with the findings of Afzal *et al.*,(2009b) observed that seeds primed with 50 mM CaCl₂ a strong association with increased total sugars is reported in

marigold and tomato (Handa *et al.*, 1983). Iqbal *et al.* (2006) in wheat, Afzal *et al.* (2008) in wheat and Bakht *et al.* (2011) in maize.

Table 2: Effect of seed priming with chemicals and PGRs on yield and yield contributing traits of wheat varieties under sodic soil

Variety Treatments	Plant height	Number of tillers plant ⁻¹	Dry biomass plant ⁻¹	EBT plant ⁻¹	No. of grain ear ⁻¹	Grain yield plant ⁻¹
Control	82.89	6.45	27.05	5.92	33.97	7.84
Hydropriming	85.38	6.74	28.95	6.17	37.28	8.44
Priming with KNO ₃	93.70	7.92	29.59	7.42	44.78	9.51
Priming with KCl	90.65	7.33	30.43	7.39	41.50	9.34
Priming with GA ₃	89.48	7.33	29.38	7.00	39.36	8.93
Priming with cycocel	81.17	7.12	29.29	6.42	38.17	8.72
SEm±	1.29	0.30	0.502	0.31	0.71	0.21
CD at 5%	3.80	0.88	1.474	0.90	2.08	0.61
HD 2733	83.80	6.45	27.59	5.97	34.52	8.03
KRL 210	90.63	7.84	30.64	7.46	43.83	9.56
SEm±	0.74	0.17	0.290	0.18	0.41	0.12
CD at 5%	2.19	0.51	0.851	0.52	1.20	0.36

In general, plant height increases with the increase of plant age. Seed priming significantly enhanced the plant height in tolerant variety KRL 210 as compared to susceptible variety HD 2733(Table-2). Maximum plant height was recorded with KNO₃ (3%) followed by GA₃ (150ppm) and hydropriming. However the minimum was recorded in case of cycocel (500ppm). Priming with cycocel had no effect on plant height. The increase in plant height might be due to stimulation of cell elongation, cell division and enlargement as reported by Tolbert (1974). These findings are in accordance with the results reported by Golizadeh*et al.*, (2015) in Cannabis seed, Jalilian *et al.*, (2014) in barley, Shabbir *et al.* (2013) in fennel, Abnavi and Ghbadi (2012) and Ghobadi *et al.* (2012) in wheat.

Seed priming significantly affected the number of tillers plant⁻¹. The tolerant variety KRL 210produced more tiller as compared to susceptible variety HD 2733 (Table-2) under priming. Priming with KNO₃(3%), KCl (1%), GA₃(150ppm),cycocel (500ppm) and hydropriming had maintained more tillers plant⁻¹ while lowest number was observed in untreatedcontrol. These results are corroborated with the findings of Farooq *et al.* (2008) in wheat, Farooq *et al.* (2006) in rice, Zhang *et al.* (2005) in potato, Cox and Otis, (1989) in wheat, Child *et al.* (1988) in oil seed and Woodward and Marshall (1987) in barley.

Seed priming treatments significantly affected dry biomass plant⁻¹ in both the varieties (Table-2) but the effect of seed priming was more pronounced on tolerant variety KRL 210 as compared to susceptible variety HD 2733. Maximum dry biomass plant⁻¹ was recorded with KNO₃(3%) followed by KCl (1%), GA₃(150ppm), cycocel(500ppm) and hydropriming as compared to untreated in both the varieties. The increased vegetative growth in turn resulted into production of higher dry matter in plants. The increase in plant dry weight due to priming treatments indicated that the photosynthetic activity and efficiency of the leaves have been increased which contributed to dry matter production. The results are in agreement with the earlier findingsShabbir

et al. (2013) in fennel, Abnavi and Ghbadi (2012) in wheat, Aymen and Cherif (2013) in coriander, Ghobadi et al. (2012) in wheat, Hamid et al. (2008) in wheat, Gurmani et al., (2006) in rice and Khodary (2004) in maize.

Yield is the synthesis and outcome of physiological and biochemical process. Priming with chemicals and plant growth regulators had proved significant effect on yield and yield components. Seed priming enhanced the yield and yield components (ear bearing tiller⁻¹, no. of grain ear⁻¹ and grain yield plant⁻¹) in both the varieties (Table-2).High accumulation of sodium in plant under saline soil leads to high pollen in fertility which results in increased sterility percentage. Yield is a summation of all metabolic processes and growth events during life cycle of a crop plants and any abiotic or biotic stress during their growth and development influence the potential productivity of crop yield. Plants grown under saline soil have chlorotic leaves which reduce their capacity of fix CO₂ as a result total biomass is affected. As we know total biomass is important character of maintain the grain yield under saline condition, poor translocation of metabolites to the reproductive sink may be also are of the reason for lower yield. The maximum increase in all the vield components were observed with KNO₃ (3%) followed by KCl (1%), GA₃ (150 ppm),cycocel (500 ppm)and hydropriming in both the varieties as compared to control.Similar findings are also reported by Farooq et al. (2006) in rice, Farooq et al. (2008) in wheat, Arif et al.,(2007) in wheat, Akhter et al., (2009) in wheat, Yari et al. (2011) in wheat, Afzal et al., (2012) in rice, Amin et al., (2012) and Ali et al., (2013) in wheat, Bakht et al. (2011) in maize.

References

[1] Abbasdokht, H. (2011). The effect of hydropriming and halopriming on germination and early growth stage of wheat (*Triticum aestivum L.*). *Desert*; **16(1)**:61-68.

Volume 4 Issue 7, July 2015

International Journal of Science and Research (IJSR)

ISSN (Online): 2319-7064

Index Copernicus Value (2013): 6.14 | Impact Factor (2013): 4.438

- Abnavi, M. S. and Ghobadi, M. (2012). The effects of [2] source of priming and post-priming storage duration on seed germination and seedling growth characteristics in wheat (Triticum aestivem L.). J. of Agril. Sci. (Toronto); 4(9):256-268.
- Afzal, I.; Butt, A.; Rehman, H.; Basra, S.M.A. and [3] Afzal, A. (2012). Alleviation of salt stress in fine aromatic rice by seed priming. Aust. J. Crop Sci., 6: 1401-1407.
- Afzal, I.; Ashraf, S.; Qasim, M.; Basra, S.M.A. and Shahid, M.(2009). Does halopriming improve germination and seedling vigour in marigold (Tagetus sp.). Seed Sci. Technol., 37: 436-445.
- Afzal, I.; Rauf, S.; Basra, S. M. A.and Murtaza G. (2008). Halopriming improves vigor, metabolism of reserves and ionic contents in wheat seedlings under salt stress. Plant Soil Environ., 54: 382-388.
- Akhter Abro, S., A.R. Mahar and A.A. Mirbahar. (2009). Improving yield performance of landrace wheat under salinity stress using on-farm seed priming. Pak. J. Bot., 41(5): 2209-2216.
- Ali, Hakoomat, Iqbal Nadeem, Shahzadi Ahmad Naeem, Sarwari Naeem, Ahmadi Shakeel and Mehmood Athar (2013). Seed priming improves irrigation water use efficiency, yield, and yield components of late-sown wheat under limited water conditions Turk J Agric For 37: 534-544.
- Almansouri M, Kinet JM, Lutts S. (2001). Effect of salt and osmotic stresses on germination in durum wheat (Triticum durum Desf.).Plant and Soil. 231: 243-254
- Amin R, Khan AZ, Khalil SK, Khalil, I. H. (2012). Effect of osmopriming sources and moisture stress on wheat. Pakistan Journal of Botany 44(3), 867-871.
- [10] Ansari O, Choghazardi HR, Sharif Zadeh F, Nazarli H. (2012). Seed reserve utilization and seedling growth of treated seeds of mountain ray (Seecalemontanum) as affected by drought stress. CercetăriAgronomiceîn Moldova. 2 (150): 43-48
- [11] Ansari, O. and Sharif-Zadeh F. (2012). Does Gibberelic acid (GA), Salicylic acid (SA) and Ascorbic acid (ASc) improve Mountain Rye (Secalemontanum) seeds Germination and Seedlings Growth under Cold Stress?. International Research Journal of Applied and Basic Sciences. 3 (8):1651-1657
- [12] Arif, M., M. Waqas, K. Nawab and M. Shahid. (2007). Effect of seed priming in Zn solution on chickpea and wheat. AfricanCrop Sci. Con. Pro., 8: 237-240.
- [13] Arnon, D.I. (1949). Copper enzyme is isolated chloroplast, polyphenyl oxidase in Beta vulgaria. Pl. Physio. 24: 1-15.
- [14] Ashraf, M, Foolad MR. (2005). Presowing seed treatment-a shotgun approach to improve germination growth and crop yield under saline and none-saline conditions. Advan. Agron. 88: 223-271
- [15] Aymen, E.M., Cherif, H. (2013). Influence of seed priming on emergence and growth of coriander (Coriandrum sativum L.) seedlings grown under salt stress. Acta Agriculturae Slovenica, 1, pp. 41-47.
- [16] Azizpour K, Shakiba MR, Khosh Kholgh Sima N, Alyari H, Moghaddam M, Esfandiari E, Pessarakli M (2010). Physiological response of spring durum wheat genotypes to salinity. Journal of Plant Nutrition 33:859-873.

Paper ID: 14061502

- [17] Azooz.,Mm.(2009).salt stress mitigation by priming with salicylic acid in two faba bean genotypes differing in salt tolerance int.j.agril,and boil.,11: 343-
- [18] Bakht J, Shafi M, Shah SR (2011). Response of maize cultivars to various priming sources. Pak J. Bot. 43:
- [19] Child R.D., Evans D.E., Hutcheon J.A., Jordan V.W., Stinchcombe G.R., 1988 - Influence of time of application of growth retardants on canopy structure, disease and yield in oilseed rape. Brighton Crop Protection Conference, Pest and Diseases. CAB International, pp, 881-886.
- [20] Cox W.J., Otis D.J., 1989 Growth and yield of winter wheat as influenced by chlormequat chloride and ethephon. Agronomy Journal, 1, 264-270.
- [21] El-Tayeb, M.A., 2005. Response of barley grains to the interactive effect of salinity and salicylic acid. Plant Growth Regul., 45: 215–224.
- [22] Esfandiari E, Shekari F, Shekari F, Esfandiari M. (2007). The effect of salt stress on antioxidant enzymes activity and lipid peroxidation on the wheat seedling.Not. Bot. Hort. Agrobot. Cluj. 35 (1): 48-56
- [23] Farooq M, Basra SMA, Rehman H, Saleem BA (2008) Seed priming enhances the performance of late sown wheat (Triticm aestivum L.) by improving chilling. J Agron Crop Sci 194: 55-60.
- [24] Farooq, M., S. M. A. Basra, I. Afzal and A. Khaliq (2006). Optimization of hydropriming techniques for rice seed invigoration. Seed Sci. Technol., 34: 507-
- [25] Foti R, Abureni K, Tigere A, Gotosa J, Gerem J. (2008). The efficacy of different seed priming osmotica on the establishment of maize (Zea mays 1.) caryopses. J arid Environ. 72: 1127-1130
- [26] Gadallah, M.A.A. (1999). Effect of kinetin on growth, grain yield and some mineral elements in wheat plants growing under excess salinity and oxygen deficiency. Plant Grwoth Regul, 27, pp 63-
- [27] Ghobadi, M.; Shafiei-Abnavi, M.; Jalali-Honarmand, S.; Ghobadi, M. E. and Mohammadi, G. R. (2012). Does KNO3 and hydropriming improve wheat (Triticum aestivum L.) seeds germination and seedlings growth? Annals of Biological Research; 3(7):3156-
- [28] Golizadeh Karbalaye Shirin, Mahmoodi Mir Tooraj and Khaliliaqdam Nabi (2015). Effect of Priming of (KNO3, ZnSo4, Distilled water) on rate Germination and Seedling Establishment on Cannabis seed (Cannabis sativa L.) Biological Forum - An International Journal 7(1): 190-194.
- [29] Gurmani A. R., Bano A. and Salim Muhammud (2006) Effect of growth regulators on growth, yield and ions accumulation of rice (Oryza sativa L.) under salt stress Pak. J. Bot., 38(5): 1415-1424.
- [30] Hamid, M.; Ashraf, M.Y.; Khalil, Ur. R.; Arshad, M. (2008). Influence of salicylic acid seed priming on growth and some biochemical attributes in wheat grown under saline condition. Pakistan J. Botany, 40: 361-367.
- [31] Handa, S., R.A. Bressan, A.K. Handa, N.C. Carpita and P.M. Hasegawa, 1983. Solutes contributing to

International Journal of Science and Research (IJSR)

ISSN (Online): 2319-7064

Index Copernicus Value (2013): 6.14 | Impact Factor (2013): 4.438

- osmotic adjustment in cultured plant cells adapted to water stress. *Plant Physiol.*, 73: 834–843
- [32] Iqbal, M. and M. Ashraf, (2006). Wheat seed priming in relation to salt tolerance: growth, yield and levels of free salicylic acid and polyamines. *Ann. Bot. Fenn.*, 43: 250–259.
- [33] Jalilian Jalal, Khalilzadeh Razieh and Khanpaye Edris (2014) Improving of Barley Seedling Growth by Seed Priming Under Water Deficit Stress *Journal of Stress Physiology & Biochemistry, Vol. 10 No. 2 2014, pp. 125-134 ISSN 1997-0838.*
- [34] Khodary, S.E.A. (2004). Effect of salicylic acid on the growth, photosynthesis and carbohydrate metabolism in salt stressed maize plants. *Int. J. Agri. Biol.*, 6:5-8.
- [35] Lara Tulio S., Jean Marcel S. Lira, Amanda C. Rodrigues, Miroslava Rakocevic and Amauri A. Alvarenga (2014). Potassium Nitrate Priming Affects the Activity of Nitrate Reductase and Antioxidant Enzymes in Tomato Germination Journal of Agricultural Science; Vol. 6, No. 2.
- [36] Lemrasky, M. G. and Hosseini, S. Z. (2012). Effect of seed priming on the germination behavior of wheat. *International J. of Agril. and Crop Sci. (IJACS)*; 4(9):564-567.
- [37] Mass EV, Nieman RH (1978). Physiology of plant tolerance to salinity. In Jung G.A. (eds.) Crop tolerance to suboptimal land conditions. ASA CSSA SSSA Madison Wisconsin, p. 277.
- [38] Menon Noor-Un-Nisha, Gandahi, Moula Bux,; and Pooja, Vajanti Mala,; and Sharif Nasm (2013) *IJASR* (3) 163-174.
- [39] Murungu FS, Nyamugafata P, Chiduza C, Clark LJ, Whalley WR. (2003). Effects of seed priming, aggregate size and soil matric potential on emergence of cotton (Gossypiumhirsutum L.) and maize (Zea mays L). Soil and Tillage Research. 74: 161-168
- [40] Parida AK, Das AB (2004). Salt tolerance and salinity effect on plants: A review. Ecotox. Environ. Safe, 60: 324-349.
- [41] Patade VY, Maya K, Zakwan A. (2011). Seed priming mediated germination improvement and tolerance to subsequent exposure to cold and salt stress in capsicum. Res J Seed Sci. 4 (3): 125-136
- [42] Rouhi HR, Aboutalebian MA, Moosavi SA, Karimi FA, Karimi F, Saman M, Samadi M. (2012). Change in several antioxidant enzymes activity of Berseem clover (TrifoliumalexandrinumL.) by priming. International Journal of AgriScience. 2(3): 237-243
- [43] Saha R, Manndal AK, Basu RN (1990). Physiology of invigoration treatments in soybean (*Glycine max L.*). Seed Sci. Technol. 18: 269-276.
- [44] Shabbir, I., Shakir, M., Ayub, M., Tahir, M., Tanveer, A., Shahbaz, M., Hussain, M. 2013. Effect of seed priming agents on growth, yield and oil contents of fennel (Foeniculum Vulgare Mill.). Advances in Agriculture & Biology, 1, pp. 58-62.
- [45] Shafi M, Bakht J, Guoping Z, Razi-U-Din E, Islam, Aman M (2010). Effect of cadmium and salinity stresses on root morphology of wheat. Pak. J. Bot. 42: 2747-2754.
- [46] Shafi M, Bakht J, Hassan MJ, Razi UM, Guoping Z (2009). Effect of Cadmium and salinity stress on

- growth and antioxidant activities of wheat. Bull. Environ. Contam. Toxicol. 82: 772-776.
- [47] Shafi M, Bakht J, Raziuddin, Zhang G (2011). Genotypic difference in Photosynthesis and Chlorophyll fluorescence of Wheat (*Triticumaestivum* L.) as affected by Cadmium and Salinity. J. Plant Nutr. 34: 315-323.
- [48] Soltani, A.; Gholipoor M, and Zeinali E. (2006). Seed reserve utilization and seedling growth of wheat as affected by drought and salinity. Environmental and Experimental Botany. **55**: 195–200.
- [49] Srinivasan, K. and S. Saxena (2001). Priming seeds for improved viability and storability in Raphanus sativus cv. Chinese Pink. Indian J. Plant Physiol., 6: 271–274.
- [50] Tolbert, N. E. (1974). Photorespiration. In Algal physiology and biochemistry, pp. 474-504. Ed. by W. D. P. Stewart. University of California Press, Los Angeles.
- [51] Wasif, N. and Mohammad, T. (2012). Enhancement of everyone potential and establishment of Moringa oleifera lam by seed priming. Turk. J. agric. For, 36: 227-235.
- [52] Woodward E.J. and Marshall C., (1987). Effects of seed treatment with plant growth regulator on growth and tillering in spring barley (*Hordeumdistichum* cv. Triumph). Annals of Applied Biology 110, 629-639.
- [53] Yari, L.; Khazaei, F.; Sadeghi, H. and Sheidaei, S. (2011). Effect of seed priming on grain yield and yield component of bread wheat (*Triticum aestivum L.*). *J. of Agril. and Biological Sci.*, 6(6): 1-5.
- [54] Zhang, Z.J.; B.Z. Mao; H.Z.Li, W.J.; Zhou, Y.; Takeuchi and Y. Yoneyama (2005). Effect of salinity on physiological characteristics, yield and quality of microtubers *In vitro* in potato. *Acta Physiol. Planta*, 27:481-490.

: 2319