

# Light and Electron Microscopic Details of blood-Spleen Barrier in Nandanam Chicken (*Gallus domesticus*)

T.A.Kannan<sup>1</sup>, Geetha Ramesh<sup>2</sup>

<sup>1</sup>Professor, Centre for Stem Cell Research and Regenerative Medicine, Madras Veterinary College, Chennai-600 007, India

<sup>2</sup>Professor and Head, Department of Veterinary Anatomy, Madras Veterinary College, Chennai-600 007, India

**Abstract:** *Light and electron microscopic studies of blood-spleen barrier was done in various age groups of Nandanam Chicken ranging from day-old to forty weeks. The spleen is the largest secondary lymphoid organ which can perform filtration of blood and also the primary site for immunocyte proliferation and differentiation. The results showed that the parenchyma was made up of white and red pulp. The white pulp appeared as islands enclosed by red pulp and there was no distinct marginal zone between the red and white pulp. The trabeculae were not evident in all the age groups studied. The arterioles from the periphery of the white pulp observed to enter the red pulp as sheathed capillaries. These capillaries were found to be surrounded by reticular cells and macrophages which formed the ellipsoids or sheathed capillaries. Under transmission electron microscope, the ellipsoids were found to have a meshwork of stellate reticular cells, reticular fibres and a few macrophages.*

**Keywords:** Light microscopy - electron microscopy - blood-spleen barrier – Chicken

## 1. Introduction

The spleen is the largest secondary lymphoid organ in Chicken, containing around 25% of the total number of lymphocytes and also the primary site for immune cell proliferation and differentiation [29] and is a specialised organ to perform filtration of blood [29]. The splenic parenchyma plays a crucial role in immune responses such as exposure to blood-borne antigens [5,29] due to the scarcity of lymph nodes and poorly developed lymphatic vessels in avian species [16,22,27,29,20].

The blood-spleen barrier [BSB] was first described in Mice by Weiss [Weiss *et al.*, 1986] and it has been confirmed by light and electron microscope [26] and also functional studies with various extracellular tracers [15]. In mouse, the BSB is composed of endothelial cells, macrophages, reticular tissue and fibres [10]. Whereas, Zhang *et al.* [2015] described that BSB in Chicken spleen is made up of ellipsoids and peri-ellipsoidal lymphatic sheath [PELS].

The ellipsoid or Schweigger-Seidel sheath, is a specialised capillary segment in the spleen of chicken [19]. Ellipsoids are common in avian species, turtles, cat, dogs, human but not in rodents and lagomorphs [11,21,19,24]. However, the structure of ellipsoid varies between species.

## 2. Literature Survey

Although, there are several detailed reports available on histoarchitecture of Chicken spleen, the study on structural details of BSB in Nandanam Chicken is very limited. Nandanam strain of chicken is a dual purpose, coloured variety with good disease resistance, developed in Institute of Poultry Production and Management, TANUVAS. Hence, the present work was aimed to study the location and

structural details of BSB using light and electron microscopy.

## 3. Materials and Methods

Over 36 specimens of thymic tissue were collected from six different age groups such as day-old, four, eight, twelve, twenty and forty weeks. Six birds were used in each age group. The spleen was removed immediately after high cervical dislocation and fixed for light and electron microscopy [8].

For light microscopy, tissue pieces were fixed in 10% neutral buffered formalin, dehydrated in ascending grades of alcohol, cleared in xylene and embedded in paraffin wax. Five micron thick sections were made and stained with Haematoxylin and Eosin [1].

For electron microscopy, small pieces of tissue were fixed for 2 hours in 3 % glutaraldehyde buffered to pH 7.4 with 0.1 M sodium cacodylate buffer at 4°C. The tissue was washed overnight in three changes of buffer at 4°C, post-fixed in 1 % osmium tetroxide in 0.1 M sodium cacodylate buffer at 4°C for 2 hours and washed overnight in buffer. Dehydration in ethanol and propylene oxide was followed by embedding in propylene oxide: epoxy resin mixture and embedded in Epon-araldite mixture. Semi thin [1 micron] sections were stained by toluidine blue. Ultra thin sections [600 Å to 900 Å] were prepared on Leica ultracut microtome, mounted on uncoated copper grids and stained with saturated solution of uranyl acetate and lead citrate [18]. The stained ultra thin sections were examined under Phillips [Teknai-10] computer augmented transmission electron microscope operated at 60-kilowatt ampere [KVA].

## 4. Results and Discussion

### Light Microscopy

In the present study, the parenchyma of the spleen was observed to have stroma composed of red pulp, white pulp and blood vessels. The white pulp appeared as islands enclosed by red pulp and the demarcation between the two was not distinct [2,29]. This could probably be the reason for less clear zonal demarcation of white and red pulp and absence of marginal zone as in the spleen of rodents and absence of marginal zone as in the spleen of rodents [12,14,17]. The trabeculae were not evident in all the age groups studied. The arterioles from the periphery of the white pulp observed to enter the red pulp as sheathed capillaries. These capillaries were found to be surrounded by reticular cells and macrophages which formed the ellipsoids or sheathed capillaries.

### Electron Microscopy

Under transmission electron microscope, the ellipsoids were found to have a meshwork of polymorphic reticular cells, reticular fibres and a few macrophages [23]. These ellipsoids may act as an antigen trapping zone of the spleen [4]. The reticular cells were stellate shaped, they had smaller nuclear-cytoplasmic ratio and more organelles in the cytoplasm [Fig-1]. Rough endoplasmic reticulum was found to be more [Fig-2]. Well developed mitochondria, ribosomes and a prominent Golgi zone were also observed. The macrophages were ovoid or stellate shaped and showed vacuoles and phagocytosed material in the cytoplasm. The nucleus was heterochromatic [Fig-3].

The presence of reticular cells and macrophages helps the ellipsoids to regulate the movement of cells and antigens between the blood vessels and white pulp [6,9]. It is hypothesised that the ellipsoid together with PELs may be comparable to the marginal zone or comparable with that of high endothelial venules of lymph node / spleen [12,25].

In the present study, BSB was located in the ellipsoid and PELs as reported by [29,3] and in contrast to the findings of [7] in rodents. There were no structural changes noticed in ellipsoid of spleen in different age groups studied.

## 5. Conclusion

The present study on Chicken spleen described the location, light and electron microscopic details of blood-spleen barrier in different age groups. The ellipsoids were found to have a meshwork of polymorphic reticular cells, reticular fibres and a few macrophages. The structure of BSB was similar in all the age groups studied and it may provide a structural basis for homing of lymphocyte in chicken. There were no structural changes noticed in ellipsoids and periarterial lymphatic sheath of spleen in different age groups. This basic study helped in better understanding of the immune function of the spleen.

## 6. Future Scope

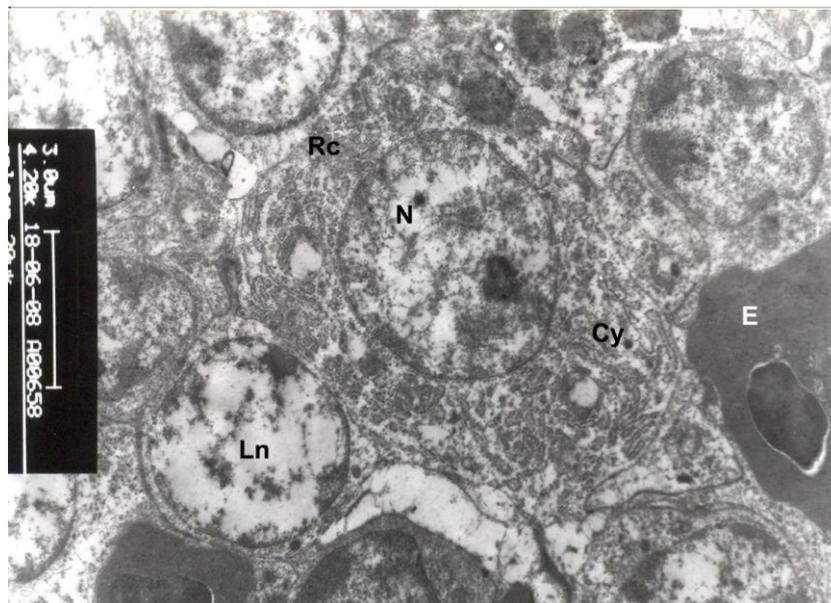
Though, the present study described the location and structure of BSB, quantitative and qualitative estimation of the T-lymphocyte subsets viz., CD-4 and CD-8 cells in ellipsoid and periellipsoidal lymphatic sheath will help in better understanding the immune function of spleen in Chicken. These observations will form a basis for further immunohistochemical and flow cytometric studies in chicken spleen.

## References

- [1] Bancroft, J.D. and A. Stevens, 2007. Theory and practice of histological techniques. Churchill Livingstone, London.
- [2] Baishya, G. and R. Bhattacharyya, 2012. Gross and Micro-Anatomy of the Spleen of Adult Indigenous Fowl of Assam. Indian Journal of Veterinary Anatomy 24 [2]: 84-86.
- [3] Bao, H.J., Li, M.Y., Wang, J., Qin, J.H., Xu, C.S., Hei, N.N., Yang, P., Gandahi, J.A., Chen, Q.S., 2009. Architecture of the blood-spleen barrier in the soft-shelled turtle, *Pelodiscus sinensis*. Anatomical Record 292, 1079–1087.
- [4] Biro, E., Kocsis, K., Nagy, N., Molnar, D., Kabell, S., Palya, V., Olah, I., 2011. Origin of the chicken splenic reticular cells influences the effect of the infectious bursal disease virus on the extracellular matrix. Avian Pathology 40, 199–206.
- [5] Brendolan, A., Rosado, M.M., Carsetti, R., Selleri, L., Dear, T.N., 2007. Development and function of the mammalian spleen. Bioessays: News and Reviews in Molecular, Cellular and Developmental Biology 29, 166–177.
- [6] Colombatti, A., Poletti, A., Carbone, A., Volpin, D., Bressan, G.M., 1989. Extracellular matrix of lymphoid tissues in the chick. Journal of Histochemistry and Cytochemistry 37, 757–763.
- [7] Cubas, A., Rolland, L., Bricaire, F., Gentilini, M., Monjour, L., 2000. The anti-malaria role of the spleen. Presse Medicale 29, 1186–1190.
- [8] Gilmore, R.S.C. and J.B. Bridges, 1974. Histological and ultrastructural studies on the myoid cells of the thymus of the domestic fowl, *Gallus domesticus*. J. Anat., 118: 409 - 16.
- [9] Gumati, M.K., Magyar, A., Nagy, N., Kurucz, E., Felfoldi, B., Olah, I., 2003. Extracellular matrix of different composition supports the various splenic compartments of guinea fowl [*Numida meleagris*]. Cell and Tissue Research 312, 333–343.
- [10] Guo GJ, Ye MF, Jiang DJ, Zhang K, Zhang TF. 2000. Morphologic study on blood-spleen barrier. J Fourth Mil Med Univ 21:177–179.
- [11] Hatae, T., 1978. Electron microscope studies on the ellipsoid of the cat spleen with special reference to the filaments in the endothelial cell. Archivum Histologicum Japonicum 41, 177–186.
- [12] Jeurissen, S.H., H.E. Claassen and E.M. Janse, 1992. Histological and functional differentiation of non-lymphoid cells in the chicken spleen. Immunology, 77: 75 – 80.

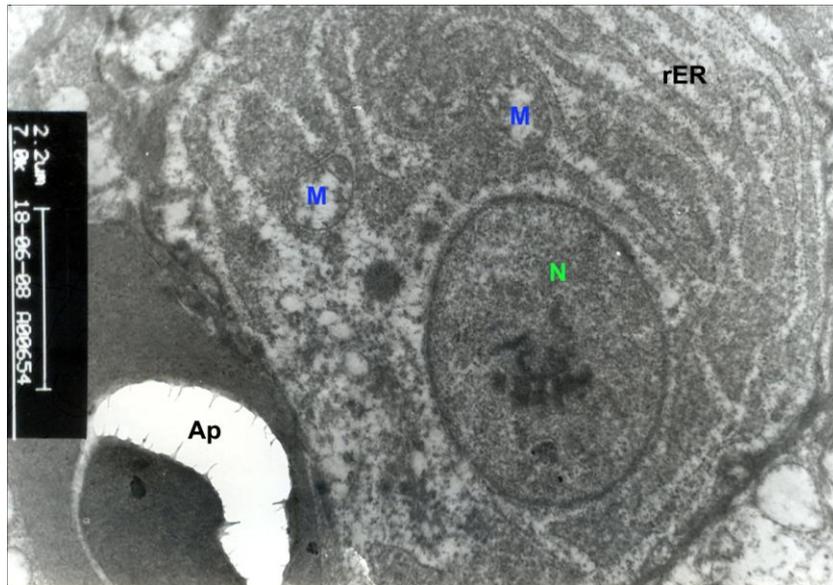
- [13] Jeurissen, S.H.M., 1991. Structure and function of the chicken spleen. *Research in Immunology* 142, 352–355.
- [14] Jeurissen, S.H., L. Vervelde and M. E. Janse, 1994. Structure and function of lymphoid tissue of the chicken. *Poult. Sci. Rev.*, **5**: 183 – 207.
- [15] Jiang DJ, Guo GJ, Chen WP, Zhang TF, Zhang K. 2002. An experimental study on structure and function of blood-spleen barrier. *Chin J Hepatobiliary Surg* 8:49–52.
- [16] John, J.L. 1994. The avian spleen: a neglected organ. *Quartly Review of Biology* **69**: 327-351.
- [17] Kannan, T.A., Geetha Ramesh, S.Ushakumari, G.Dhinakarraj and S.Vairamuthu, 2012. Light microscopic studies on Spleen of Chicken [*Gallus domesticus*]. *Haryana Vet.*, 51[12] ; 114 - 115.
- [18] Kannan, T.A., Geetha Ramesh, S.Ushakumari, G.Dhinakarraj and S.Vairamuthu, 2015. Histological and Ultrastructural studies of Caecal tonsil in Chicken [*Gallus domesticus*]. *Int.J.Curr.Microbiol.App.Sci.*, 4[6] : 63-68.
- [19] Kasai, K., Nakayama, A., Ohbayashi, M., Nakagawa, A., Ito, M., Saga, S., Asai, J., 1995. Immunohistochemical characteristics of chicken spleen ellipsoids using newly established monoclonal antibodies. *Cell and Tissue Research* 281, 135–141.
- [20] Kita, K., 2014. The spleen accumulates advanced glycation end products in the chicken: Tissue comparison made with rat. *Poultry Science* 93, 429–433.
- [21] Kroese, F.G., Van Rooijen, N., 1983. Antigen trapping in the spleen of the turtle, *Chrysemys scripta elegans*. *Immunology* 49, 61–68.
- [22] Nagy, N., Biro, E., Takacs, A., Polos, M., Magyar, A., Olah, I., 2005. Peripheral blood fibrocytes contribute to the formation of the avian spleen. *Developmental Dynamics* 232, 55–66.
- [23] Olah, I. and N. Glick, 1982. Splenic white pulp and associated vascular channels in chicken spleen. *Am. J. Anat.*, **165**: 445 - 80.
- [24] Onkar, D.P., Govardhan, S.A., 2013. Comparative histology of human and dog spleen. *Brazilian Journal of Morphological Sciences* 30, 16–20.
- [25] Venkatesan, S., Geetha Ramesh and Vijayaragavan, C. 2005. Age related changes in histomorphology of the spleen of the Japanese quail. *Indian Journal of Veterinary Anatomy*, **17**: 19-23.
- [26] Weiss L, Geduldig U, Weidanz W. 1986. Mechanisms of splenic control of murine malaria: reticular cell activation and the development of a blood-spleen barrier. *Am J Anat* 176:251–285.
- [27] Yang, S.B., Zhang, Y.N., Zhang, Y.L., Gao, J.J., Yang, L.H., Nuan, W.M., 2010. Development and localization of B lymphocytes in chicken spleen. *Chinese Journal of Veterinary Science* 30, 379–383.
- [28] Zhang, Y.N., Yang, S.B., Yang, L., Nuan, W.M., 2010. Dynamic observation of histological development of spleen in chicken. *China Poultry* 32, 13–15.
- [29] Zhang, Q., B. Chen, P. Yang, L. Zhang, Y. Liu, S. Ullah, L. Wu, Y. Waqas, Y. Le, W. Chen, Q. and Chen, 2015. Identification and structural composition of the blood-spleen barrier in chickens. *The Veterinary Journal* 204 : 110–116.
- [30] Zhu AL, Jiang HC, Liu LX, Piao DX, Pan SH, Qiao HQ. 2005. The study on the morphology character of blood-spleen barrier. *Chin J. Surg* 43:591–594.

### List of Plates and Legends



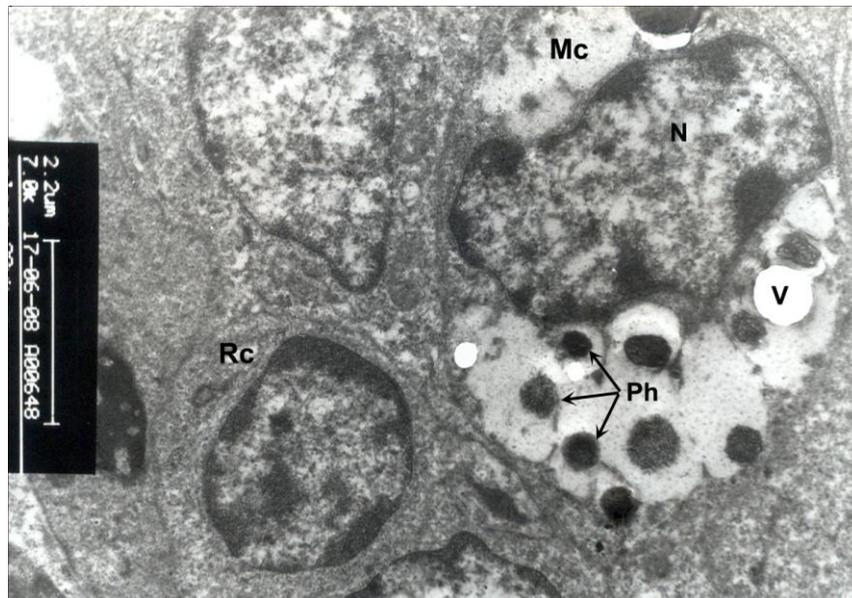
**Figure 1:** Transmission electron micrograph of spleen of a fortytwo week-old chicken showing the details of reticulum cell in white pulp x 4200

Cy - Cytoplasm  
 N - Nucleus of reticulum cell  
 E - Erythrocyte  
 Ln - Nucleus of Lymphocyte  
 Rc - Reticulum cell



**Figure 2:** Transmission electron micrograph of spleen of a forty week-old chicken showing the cytoplasmic organelles of reticulum cell in white pulp x 7000

Ap - Apoptotic cell M - Mitochondria  
N - Nucleus rER - Rough endoplasmic reticulum



**Figure 3:** Transmission electron micrograph of spleen of twenty week-old chick showing a macrophage in the white pulp x 7000

Mc - Macrophage N - Nucleus of macrophage  
Ph - Phagocytosed material Rc - Reticulum cell  
V - Vacuole