Assessment of Nematode Distribution and Yield Losses in Vegetable Crops of Western Uttar Pradesh in India

Rajendra Singh¹, Umesh Kumar²

Plant Nematology Research Lab, Department of Zoology, Bareilly College Campus, MJP Rohilkhand University, Bareilly- 243005, U.P., India

Abstract: Plant-parasitic nematode population densities were determined from 412 root and soil samples collected from vegetable growing areas of Western Uttar Pradesh province of India. The most abundant plant-parasitic nematodes detected, in order of decreasing frequency of infestation (percentage of samples), were Meloidogyne incognita (82.16%), M. javanica (68.42), Rotylenchulus reniformis (43.98%), Xiphinema basiri (23.45%), Hoplolaimus indicus (21.99%), Tylenchorhynchus nudus (16.60%), Pratylenchus zeae (15.77%), Heterodera avenae (7.88%), Aglenchus costatus (7.47%), Tylenchorhynchus mashoodi (6.64%), Aphelenchus avenae (5.39%), Discolaimus (4.15%), Boleodorus similis (3.32%), and Tylenchus (2.07%). Yield losses associated with nematode presence were quantified on 17 vegetable crops during 2012 to 2015. Based on our visual field observations and grower responses the average percentage of production losses due to nematode was 23.70% for 17 vegetable crops in Western UP, which ranged from 4% for bitter gourd to 43% for egg plants. Maximum yield loss of 43% was observed in eggplant followed by 40% in tomato, 38% in okra, 35% in bottle gourd and 32% in potato. Bitter gourd and cabbage were found to have insignificant yield losses of only 4% and 6% respectively.

Keywords: Plant-parasitic nematodes, distribution, vegetables, yield losses, Uttar Pradesh

1. Introduction

Vegetables are important component in nutritional security, economic viability and fit well into the predominant intensive cropping systems prevailing in different parts of India. More than 60 kinds of vegetables are grown in India in tropical, subtropical and temperate agro-climates. During 2013-2014, India produced 162.19 million tonnes of vegetables and vegetable export from our country was worth Rs. 5462.93 crores. This production is still insufficient for second most populous country in the world where India shares 17.5% of global population. The land area of India is about 2.4 per cent of the total surface area of the world but we share only about 1%, in global market of horticultural products. The agricultural production in India is greatly hampered by the parasitism and predation of different pathogen and pests. Among them plant-parasitic nematodes present a formidable pest problem for different crops including vegetables, fruits, field crops, ornamentals and common weeds (Saxena & Singh, 1997; Anes & Gupta, 2014; Ngele & Kalu, 2015). Most species attack and feed on plant roots and underground plant parts. Nematodes not only suppress the plant growth but also interfere in the nodulation, nitrogen fixation and adversely affect the overall yield (Rehman et al., 2012).

Root-knot nematodes, *Meloidogyne* spp. have become a major pest of almost all types of crops, impacting both quantity and quality of marketable yields in the Western Region of Uttar Pradesh. These nematodes reduce the plants ability to extract available soil water and nutrients, the result being lack of vigor and yield loss (Trudgill, 1992).

Several workers have attempted to assess crop losses caused by plant parasitic nematodes and their distribution in India. Van Berkum and Seshadri (1970) were the first to have calculated monetary losses to crops caused by nematode parasites in India. They estimated annual losses of \$10 million on wheat due to 'ear cockle disease' caused by *Anguina tritici*, \$ 3 million on coffee due to *Pratylenchus coffeae* and \$ 8 million due to "Molya disease" caused by *Heterodera avenae* in Rajasthan province alone. Plantparasitic nematodes cause estimated annual crop losses of \$8 billion in the United States and \$78 billion worldwide (Barker *et al.*, 1998). Damage caused by plantparasitic nematodes on 24 vegetable crops in the USA was estimated to be 11% by Feldmesser *et al.*, in 1971.

The objective of this study was to quantify and document of relative occurrence, distribution, density and prevalence of different nematode populations and the yield losses they cause in associated crops cultivated in Western part of Uttar Pradesh, India.

2. Materials and Methods

2.1 Selection of Sites for Survey

A Survey was carried out to take the root and soil samples from crop fields of 118 localities of 5 districts of Western Uttar Pradesh of India at geographic location of 28.364°N 79.415°E and elevation of 268 meters. The climate of Uttar Pradesh is primarily defined as humid subtropical with dry winter and tropical monsoon suitable for different vegetable and cash crops. Seventeen vegetable crops including carrot, chilies, coriander, crucifers (cabbage and mustard), cucurbits (bitter gourd, cucumber, pumpkin, sponge gourd, melon and bottle gourd), eggplant, okra, pea, potato, spinach, and tomato from commercial fields located throughout Western UP were sampled during 2012 to 2015. Presence of nematodes in vegetable roots and soil was determined at the time of harvesting of each crop. A total of 412 root and soil samples were collected over a three year of period.

2.2 Sampling Methodology

Ten root and soil cores were randomly collected for each crop by walking in a zigzag pattern across each field with a tube of 2.5 cm diameter inserted to a depth of 18-20 cm. Soil cores were combined to represent each sampled field. Samples were packed into plastic bags, placed in a cooler for transport and then stored at 4° C until processed in the lab. Roots were separated from soil and carefully washed under tap water to remove adhering soil particles and then towel dried.

2.3 Nematode Extraction

Nematodes were extracted by following the method of McSorely (1987) from a fresh root composite sub sample of 25g by placing them in a mist chamber for 5 days. Isolated nematodes were heat killed at 60-65°C and then fixed in 4% formalin and placed in viols. Prior to counting, solution containing nematodes were agitated thoroughly and 3 ml suspension poured to a counting dish. Nematodes were counted using a stereo binocular microscope. Selected specimens for each of the recorded species were processed for dehydration by Seinhorst's (1959) rapid glycerin method and mounted on glass slides in anhydrous glycerin. Plantparasitic nematodes were identified on the basis of morphology of adults and juveniles to genus and species using a high magnification binocular compound microscope.

2.4 Assessment of yield losses

The criteria used to assess yield loss comprised grower interviews, visual assessment based on foliage growth (necrotic, chlorotic, stunted, and wilted plants), root symptoms and educated guess to expert opinions. The number of growers interviewed was variable and ranged from 5-15 for each crop. The interview of growers included condition of the crop, quantitative and qualitative yield losses based on market value and life span of the crop. Estimates reported in this study are expressed as percentage of yield loss. All data collected were subjected to analyze statistically by using Software GraphPad Prism (version 5.0), USA.

3. Results and Discussion

3.1 Nematode survey

Fourteen species of plant-parasitic nematodes were found associated with 17 vegetable crops grown in the different field crops of Western UP (Table 1). The nematode genera found were Meloidogyne incognita, M. javanica, Rotylenchulus reniformis, Xiphinema basiri, Hoplolaimus indicus, Tylenchorhynchus nudus, Pratylenchus zeae, Heterodera avenae, Aglenchus costatus, Tylenchorhynchus mashoodi, Aphelenchus avenae, Discolaimus, Boleodorus similis, and Tylenchus. Five species M. incognita, Rotylenchulus reniformis, Xiphinema basiri, Pratylenchus zeae, Heterodera avenae, were found in all the samples, however, their frequency and density was highly variable from field to field and within the same locality. Frequency of these nematode species ranged from 82.16% to 2.07%. The two potentially damaging nematode genera identified were Meloidogyne and Rotylenchulus.

Root-knot nematode was found to infest the roots and soil of all the vegetable crops. This nematode induces severe root galling of variable size and numbers on roots of vegetables, such roots had arrested root systems with few feeder roots (personal visual observations). The most common nematode species found in this study were *M. incognita*, *M. javanica* and, *Rotylenchulus reniformis* which were found at the relative percent occurrence (RPO) of 82.16%, 68.42% and 43.98% respectively. Average population density of *M. incognita* was 310 individuals per 100cm³ and 1510 per gram of soil and root, respectively, whereas the number of *Rotylenchulus reniformis* found in the root and soil were 1000 per g and 432 per cm³, respectively (Table 1).

The migratory ectoparasitic nematodes including sting nematode, *B. longicaudatus*, spiral nematode, *H. dihystera.*, lance nematode, *H. columbus*, needle nematode, *L. africanus*, stubby root nematode, *P. minor*, stunt nematode, *T. clarus* and dagger nematode, *X. indicus* have been found to be damaging nematode pests of many vegetable crops as they cause destruction of epidermis during feeding. (Cooke, 1989; McKenry *et al.*, 2001)

 Table 1: Relative Percent Occurrence (RPO) and Relative

 Density (RD) of different nematode genera in Western UP,

 India

		India				
<i>S</i> .	Nematode species	RPO	RD	Nematode population		
No		(%)	(%)	densities		
				100 cm ³	Per g of	
				of soil	root	
1	Meloidogyne incognita	82.16	46.85	310 ± 75	1510 ± 200	
2	M. javanica	68.42	23.12	215 ± 50	800 ± 100	
3	Rotylenchulus reniformis	43.98	15.48	432±50	1000±20	
4	Xiphinema basiri	23.45	4.98	12 ± 4	4±2	
5	Hoplolaimus indicus	21.99	9.44	94±2	12±1	
6	Tylenchorhynchus nudus	16.60	3.17	622±12	424±42	
7	Pratylenchus zeae	15.77	5.07	322±78	212±54	
8	Heterodera avenae	7.88	1.80	112±2	34±4	
9	Aglenchus costatus	7.47	2.10	46±6	8±2	
10	Tylenchorhynchus	6.64	1.10	76±2	14±4	
	mashoodi					
11	Aglenchus avenae	5.39	4.47	22±1	-	
12	Discolaimus	4.15	0.80	12±4	8±2	
13	Boleodorus similis	3.32	1.29	23±12	-	
14	Tylenchus	2.07	1.29	95±10	-	

Plant-parasitic-nematodes found associated with vegetable crops can be classified as epidermal, cortical, and vascular feeders. Epidermal feeders include Hoplolaimus indicus, Tylenchorhynchus spp., H. indicus, Xiphinema basiri and Pratylenchus zeae. These nematode species feed at or close to root tips where they arrest root elongation and disrupt the site of plant growth factors. Root hairs, a single cell extension of the epidermis, enlarge plant surface area and improve efficiency in absorbing water while also providing sites for Rhizobium invasion in legumes. Invasion by ectoparasitic nematodes reduces the ability of the epidermis to absorb water by pruning root hairs, by reducing rooting depth and numbers of branched roots (Anwar and Van Gundy, 1989, Endo, 1975). Cortical feeders including H. indicus, H. galeatus and Pratylenchus sp, are migratory endoparasites. Carneiro et al., (2002) reported that feeding

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activities result in cellular necrosis which interferes with radial transport of water and solutes and leakage of stored photosynthetic products. Vascular feeders in these studies included two species of root-knot nematode, M. incognita and M. javanica, which become sedentary endoparasites. This group of nematodes damages their hosts by redirecting large amounts of energy and nutrients from normal activities into development of the nematodes and their special feeding sites (Anwar, 1995; McClure, 1977). The altered tissues at feeding sites also disrupt the vascular system hampering the upward transport of water and dissolved nutrients by xylem and translocation of photosynthates to other regions of the plant by phloem (Hajera et al., 2009). Roots severely galled by root-knot nematodes can predispose plants to root rots leading to a shorter life span of the crop. These galled tissues become succulent, poorly protected from invasion, and rich in nutrients. The result is a nutrient-rich food source which fungi can rapidly colonize (Abawi and Chen, 1998). Rootknot nematode damage results in poor growth, decline in quality and yield of the crop and reduced resistance to other stresses like drought and disease. Meloidogyne incognita has often been reported as a damaging nematode pest of vegetable crops including recent work by these authors (Anwar and McKenry, 2010). A high level of root-knot nematode damage can lead to total crop loss. Gautam et al., 2014 found the Root-knot nematode infection with an overall incidence of 54.54% in five district of Chhattisgarh. Nematode damaged roots do not utilize water or fertilizers as effectively, leading to additional losses for the grower (Trudgill and Phillips, 1997). In smaller commercial and backyard settings of the Uttar Pradesh plants are often invaded by several different nematode genera and simultaneously cause damage at all three regions of a root: the epidermis, cortex and vessels. Such plants exhibit retarded growth, chlorotic leaves, delayed flower and fruit formation, susceptibility to fungal/bacterial/viral attack plus significant growth and yield reductions (Trudgill, 1992).

3.2 Assessment of Yield Losses

Based on our visual field observations and grower responses the average percentage of production losses due to nematode was 23.70% for 17 vegetable crops in western UP, which ranged from 4% for bitter gourd to 43% for egg plants (Table 2). Each of the seventeen commercially important vegetable crops planted in the western UP suffered greater losses due to nematode damage individually as well as on an average of 23.70% of all seventeen studied crops.

Multiple nematode associations appear to be causing synergistic increases in yield loss. Nematode damage experienced within relatively small-sized farms on very different soils following various cropping histories could be a major source of the greater nematode damage in the Punjab. Reliable crop loss estimates are important for establishing research, extension and budget priorities (Dunn, 1984).

Name of the vegetable crop			ield losses	Nematode invading tissue root		
Common	Scientific Name		Epidermal	Cortical	Vascular	
Bitter gourd	Momordica charantia	4	Hoplolaimus indicus		Meloidogyne incognita	
			Tylenchorhynchus spp			
Carrot	Daucus carota	25	H. indicus		M. incognita	
Eggplant	Solanum melongena	43		Pratylenchus spp	M. incognita	
Cucumber	Cucumis sativus	25	Xiphinema basiri Paratylenchus zeae	Pratylenchus spp.	Meloidogyne spp	
Okra	Hibiscus esculentum	38	H. indicus	Pratylenchus spp.	Meloidogyne spp	
Potato	Solanum tuberosum	32	Tylenchorhynchus spp.	Heterodera avenae	Meloidogyne spp	
Pumpkin	Cucurbita argyrosperma	27	Tylenchorhynchus spp		M. incognita	
Sponge gourd	Luffa cylindrica	15	Hoplolaimus indicus		M. incognita	
Tomato	Lycopersicum esculentum	40			M. incognita	
Watermelon	Citrullus lanatus	12	Tylenchorhynchus spp		M. incognita	
Bottle gourd	Lagenaria siceraria	35	Tylenchorhynchus spp		Meloidogyne spp	
Chilies	Capsicum annuum	20	H. indicus		M. incognita	
			P.zeae			
Pea	Pisum sativum	20	P. zeae		M. javanica	
	Coriandrum sativum	25	H. indicus	Pratylenchus spp	M. incognita	
			P. zeae			
Cabbage	Brassica oleracea	6	H. indicus	Pratylenchus spp	M. incognita	
			P. zeae			
Beet	Beta vulgaris	12	H. indicus		M. javanica	
Bean	Phaseolus vulgaris	27	H. indicus		M. incognita	

Specific estimates of vegetable crop losses due to *M. incognita* and *M. javanica* have ranged from 17 to 20% for eggplant, *Solanum melongena*, 18 to 33% for melon, *Cucumis melo*, 24 to 38% for tomato, *Lycopersicon esculentum*, and 25% for potatoes, *S. tuberosum* (Kathy, 2000). Ten genera of plant- parasitic nematodes other than root-knot have also been reported associated with vegetable crops, including *Rotylenchulus sps.*, *Xiphinema sps*, *Hoplolaimus sps*, *Tylenchorhynchus sps*, *Pratylenchus sps*,

Heterodera sps, Aglenchus sps, Aphelenchus sps, Discolaimus sps, Boleodorus sps., Singh (1999). During a preliminary survey, several vegetable crops including bitter gourd, cabbage, carrot, chilies, cowpea, cucumber, eggplant, lettuce, melon, mustard, okra, potato, pumpkin, sponge gourd, squash and tomato were found to be infected with root-knot nematodes and other plant-parasitic nematode species (Anwar *et al.*, 2007). Nevertheless, the information on the losses inflicted by these nematodes on vegetable

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crops is not available in India.

Crop losses due to plant parasitic nematodes are estimated to be about 12.3% in developed nations and 14.6% in the developing countries (Sasser and Freckman, 1987). However, these early estimates of nematode damage to vegetables appear underestimated when evaluating vegetable crops in third world countries. Data from the US indicate 4.5% damage for 28 vegetable crops (McSorley et al., 1987) but the estimates are 12% in India (Sehgal and Gaur, 1999). We observed 23.70% loss on 17 vegetable crops commercially grown in the Western UP. These are 80% and 46% higher compared to that of USA, and India, respectively. Each of the 13 commercially important vegetable crops planted in the Western UP suffered greater losses due to nematode damage individually as well as on an average (21%) of all 13 compared to that of planted in USA (6.23%). In vegetable production areas of developing nations like India the experience is 43% (Bhatti and Jain, 1977; Reddy, 1980, 1985) or in Sudan the estimate is 70% (Yasin, 1974) yield loss in the tomato crop due to root knot nematode.

These finding support the present work, as here also various parasitic as well as soil nematodes had been noticed affecting the plant growth of vegetable crops. Paruthi & Gupta (1985) also studied the pathogenecity of *M. javanica* on bottle gourd. Later on Netscher & Sikora (1990) investigated different species of root-knot nematode and found moderate occurrence of *M. arenaria* and severe infestation of *M. javanica* in bottle gourd crops whereas in sponge gourd, both, *M. incognita* and *M. javanica* rank highest with moderate infection of *M. javanica*.

The dominance of *M. incognita* (53.37%) in northern Uttar Pradesh including Bareilly region had also been reported by Khan *et al.*, (1994). Khan & Khan (1996) reported the distribution of root-knot nematode species and race infesting vegetable crops in eastern Uttar Pradesh as 69.9% of *Meloidogyne incognita* infection.

Prior to the present work Jain (1992) and Jagpal (1997) had surveyed the nematode infestation of many other vegetables belonging to Family Solanaceae, cruciferae and Cucurbitaceae of different regions of Rohilkhand Division and reported mainly heavy infection of *M. incognita*. Jagpal (1997) has reported upto 92.33% occurrence of *M. incognita* on tomato, eggplant and okra, whereas, in the current study it ranged 82.16% to 88.33% on bottle gourd and sponge gourd respectively.

The present study provides important information to extension specialists, who can be used to create awareness among growers, it should alert the plant scientists to consider nematodes major damaging pests of crops and start searching for resistant cultivars as an option in plantparasitic nematode management.

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Reference

- Abawi, G.S. and Chen (1998). Concomitant pathogen and pest interactions. In: *Plant and nematode interaction* (eds. K.R. Barker, G.A. Pederson, and G.L. Windham), Agronomy Monograph **36**. American Society of Agronomy, Madison, WI, pp. 135-158.
- [2] Anes, K.M. and Gupta, G.K. (2014). Distribution of plant parasitic nematodes in the soyabean (*Glycine max*) growing areas in India. *Indian J Nematology.*, 40 (2):227-231.
- [3] Anwar, S.A. (1995). Influence of *Meloidogyne ingconita, Paratrichodorus minor* and *Pratylencus scribneri* on root-shoot growth and carbohydrate partitioning in tomato. *Pakistan J. Zool.*, **27**:105-113.
- [4] Anwar, S.A. and Mckenry, M.V. (2010). Incidence and reproduction of *Meloidogyne incognita* on vegetable crop genotypes. *Pakistan J. Zool.*, **42**:135-141.
- [5] Anwar, S.A. and Van Gundey, S.D. (1989). Influence of four nematodes on root and shoot growth parameters in grape. *J. Nematol.*, 21:276-283.
- [6] Anwar, S.A.; Zia, A.; Hussain, M. and Kamran, M. (2007). Host suitability of selected plants to *Meloidogyne incognita* in the Punjab, Pakistan. *Int. J. Nematol.*, 17:144-150.
- Barker, K.R.; Pederson, G.A. and Windham, G.L. (1998). *Plant and nematode interactions*. Agronomy Monograph 36. American Society of Agronomy, Madison, WI.
- [8] Bhatti, D.S. and Jain, R.K. (1977). Estimation of loss in okra, tomato and brinjal yield due to *Meloidogyne javanica*. *Indian J. Nematol.*, **7**:37–41.
- [9] Carneiro, R.G.; Mazzafera, P.; Ferraz, L.C.C.B.; Muraoka, T. and Trivelin, P.C.O. (2002). Uptake and translocation of nitrogen, phosphorus and calcium in soybean infected with *Meloidogyne incognita* and *M. javanica. Fitopatol. Bras.*, 27:141-150.
- [10] Cooke, D.A. (1989). Damage to sugar-beet crops by ectoparasitic nematodes, and its control by soil-applied granular pesticides. *Crop Protect.*, **8**:63-70.
- [11] Dunn, R.A. (1984). How much do plant nematodes cost Floridians? *Ent. Nematol. News*, 10:7-8.
- [12] Endo, B.Y. (1975). Pathogenesis of nematode-infected plants. *Ann. Rev. Phytopath.*, **13**: 213-238.
- [13] Feldmesser, J.; Edwards, D.I.; Epps, J.M.; Heald, C.M.; Jenkins, W.R.; Johnson, H.J.B.; Lear, C.W.; McBeth, C.W.; Nigh, E.L. and Perry, V.G. (1971). Estimated crop losses from plant-parasitic nematodes in the United States. *Comm. Crop losses*. Special publication No. 1. Soc. Nematologists, Hyattsville, Maryland.
- [14] Gautam, S.K.; Sahu, G.; Verma, B.K. and Poddar, A.N. (2014). Status of Root-Knot Nematode (*Meloidogyne* sps) disease in vegetable crops of some districts of central plain region of Chhattisgarh state, India. *African journal of Microbiology research*, 8(16):1663-1671.
- [15] Jagpal, S.K. (1997). Biocontrol of nematodes Meloidogyne and Criconemoides with plant extracts.

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Ph.D. Thesis, Rohilkhand University, Bareilly, pp.105.

- [16] Jain, S.K. (1992). Plant parasitic and soil nematodes of Rohilkhand Division: Survey and Control. Ph.D. Thesis, Rohilkhand University, Bareilly, pp. 143.
- [17] Khan, A.A. and Khan, M.W. (1996). Distribution of root-knot nematodes species and races infecting vegetable crops in Eastern Uttar Pradesh. *Indian J Nematol.*, 26(2): 238-244.
- [18] Khan, M.W.; Ashraf, S.S. and Ansari, N.A. (1994). Incidence and intensity of root-knot nematode and identity of species/races associated with vegetable crops in some districts of U.P. India. *Indian J. Nematol.*, 24(1): 388-44.
- [19] McClure, M.A. (1977). *Meloidogyne incognita*: A metabolic sink. J. Nematol., 9:88-90.
- [20] McKenry, M.V.; Kretsch, J.O. and Anwar, S.A. (2001). Interactions of selected rootstocks with ectoparasitic nematodes. *Am. J. Enol. Vitic.*, **52**:304-309.
- [21] McSorley, R.; Arenett, J.D.; Bost, S.S.; Carte, W.W.; Hafez, S.A.; Johnson, A.W.; Kirkpatrick, T. Nyczepir, A.P.; Radewald, J.D.; Robinson, A.F. and Schmitt, D.P. (1987). Bibliography of estimated crop losses in the United States due to plant-parasitic nematodes. *Annls App. Nematol.*, **1**: 6-12.
- [22] Ngele, K. K. and Kalu, U. N. (2015). Studies on different species of plant parasitic nematodes attacking vegetable crops grown in Afikpo North L.G.A, Nigeria. *Direct Research Journal of Agriculture and Food Science*, **3** (4): 88-92
- [23] Netscher, C. and Sikora, A. (1990). Nematode parasites of vegetable. In, Plant Parasitic Nematodes in Subtropical and Tropical Agriculture.(Eds. Sikora, RA Luc, M. and Bridge J.), *CAB international*, 237-283.
- [24] Paruthi, I.J. and Gupta, D.C. (1985). Pathogenicity of root-knot nematode, *Meloidogyne javanica* on bottle gourd. (*Lagenaria siceraria*) using two methods of inoculation. *Indian J. Nematol.*, 15:52-54.
- [25] Reddy, D.D.R. (1980). Root knot nematode. A global menace to crop production. Pl. Dis., **104**:36-41.
- [26] Reddy, D.D.R. (1985). Analysis of crop losses in tomato due to *Meloidogyne incognita*. *Indian J. Nematol.*, 15:55–59.
- [27] Rehman, B.; Ganai, M.A.; Parihar, K.; Siddiqui, M.A. and Usman, I. (2012). Management of root knot nematode, *Meloidogyne incognita* affecting chickpea, *Cicer arietinum* for sustainable production. *Bioscience International*, 1(1): 01-05.
- [28] Sasser, J.N. and Freckman, D.W. (1987). A world perspective on nematology: the role of the society. In: *Vistas on nematology* (eds. J.A. Veechand and D.W. Dickson). Society of Nematologists, Hyattsville, Maryland. USA. pp. 7-14.
- [29] Saxena, R. and Singh, R. (1997). Survey of nematode fauna of groundnut, *Arachis hypogea* in and around Bareilly Region, U.P. India. *Current Nematology* 8(1&2):93-97.
- [30] Sehgal, H.L. and Gaur, H.S. (1999). *Important nematode problems of India*. Technical Bulletin NCIMP, New Delhi, India, pp. 16.
- [31] Seinhorst, J. W., 1959. A rapid method for the transfer of nematodes from fixative to anhydrous glycerin.. *Nematologica*, **4:** 67 - 69.
- [32] Singh, R. (1999). Evaluation of some natural plant

extracts against root-knot nematode, Meloidogyne sps. On cucurbitaceae. Ph.D. thesis. Rohilkhand University, Bareilly, pp.31

- [33] Singh, R.V. and Khera, S. (1984). Plant parasitic nematode associated with vegetable crops around Calcutta. *Indian J. Nematol.*, **14(1)**: 188-190.
- [34] Trudgill, D.L. (1992). Resistance to and tolerance of plant-parasitic nematodes in plants. Ann. Rev. Phytopath., 29:167-192.
- [35] Trudgill, D.L. and Phillips, M.S. (1997). Nematode population dynamics, threshold levels and estimation of crop losses. En: http://fao.org/docrep/v9978e/ v9978e07.htm; consulta: noviembre de 2005.
- [36] Yasin, A.M. (1974). Root-knot nematodes in the Sudan and their chemical control. *Nematol. Mediterr.*, **2**:102-112.