

filtrates were poured in petridishes and then were incubated at 37°C for drying to produce crude ethanolic and methanolic extracts. Water extract was obtained by lyophilizing the filtrate for the removal of water. The extracts were then used for the antimitotic and *in vitro* cytotoxicity assay.

2.3. Tumour Cells

Sarcoma-180 (S-180) cells were collected from the Department of Biotechnology, Indian Institute of Technology, Kharagpur. Cells were maintained by intraperitoneal inoculation of 1×10⁶ cells/mice [16] and developed a milky white fluid containing rounded tumor cells.

2.4. Study of Mitotic index^[17]

The red variety of *Allium cepa* were collected from local market, the bulbs of *Allium cepa* were sprouted in water saturated sand tub for 3 to 4 day at room temperature. The roots thus developed were treated by dipping these in the aqueous, methanolic and ethanolic solution of *Anacardium occidentale* leaf extract at the concentration 100mg/ml for 24 hours. Treatment of roots with distilled water served as control. The roots thus treated with above solutions were then cut to separate root tips and the root tips are transferred to fixing solution, 1:3v/v aceto-alcohol for 1 hour in room temperature. Then the root tips were taken out and preserved in 70% alcohol in refrigerator. At the time of staining, the root tips were taken out from 70% alcohol and immersed in 2% acetic acid and 1N HCl and boiled at the smearing point. Then the root tips were placed on grease free slide added with 2% acetic acid, covered with cover slip then squashed to prepare smear and observed under microscope. The numbers of cells in each stage of cell division were counted in four fields for each group.

$$\text{Mitotic index} = \frac{\text{Number of dividing cells}}{\text{Number of Total cells}} \times 100$$

2.5. Study of Short-Term in Vitro Cytotoxicity: [18]

Short term cytotoxic activity of MEAOL, EEAOL, and WEAOL were assayed by determining the percentage viability of Sarcoma-180 cells using the trypan blue dye exclusion technique (by Modified Dongre S H. et al). Sarcoma-180 cells were cultured in healthy albino mice weighing between 20-28 g by serial intraperitoneal inoculation of a suspension of Sarcoma-180 cell (1×10⁶ cells/ml). The cells were aspirated aseptically from the peritoneal cavity of mice on 18th day of inoculation and washed with PBS and centrifuged for 5 min. at 1000 rpm in a cooling centrifuge. The pellet was re-suspended with RPMI 1640 medium (10% FBS and antibiotic solution) and seeded in flat bottom culture plate with MEAOL EEAOL and WEAOL at the concentration of 100 µg/1×10⁶cell/ml. The culture plates were incubated for 3hrs at 37°C and 5% CO₂. Then trypan blue dye exclusion technique was performed to determine the percentage of viability at the end of 1st, 2nd and 3rd hrs. The dead cells failed to exclude the stain and appeared in blue colour. The percentage

viability was calculated by counting number of viable cells/100 cells in each microscopic field.

2.6. Statistical Analysis

The values are given as mean ±SEM and the data was analyzed by Student's t-test. P values less than 0.05 were considered statistically significant.

3. Results

The effects of EEAOL, MEAOL and WEAOL on percentage of cells in different stages of mitosis and mitotic index in *Allium cepa* root tips are shown in Fig-1(A, B). All the treated group shows significantly reduced number of dividing cell compared to water treated group. The mitotic index of EEAOL, MEAOL and WEAOL treated group and water control group were 40, 30, 30, and 51.58 respectively.

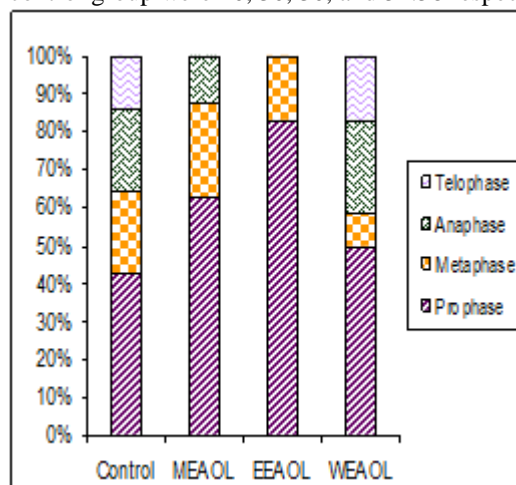


Figure 1A

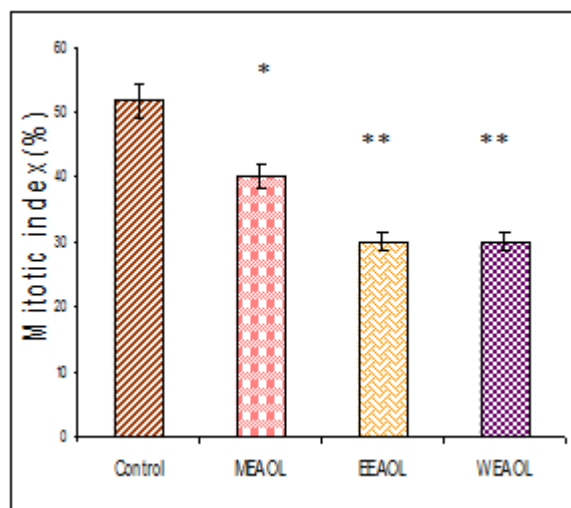


Figure 1B

Fig-1. The bar diagram shows the effect of EEAOL, MEAOL and WEAOL on % of cells in different stages of mitosis (Fig.1, A) and mitotic index (Fig.1, B) in *Allium cepa* root tips. Data are expressed as mean ± SEM. Probability values are given in asterisks. * indicates P< 0.05, ** indicates p<0.01; values are taken in respect of control.

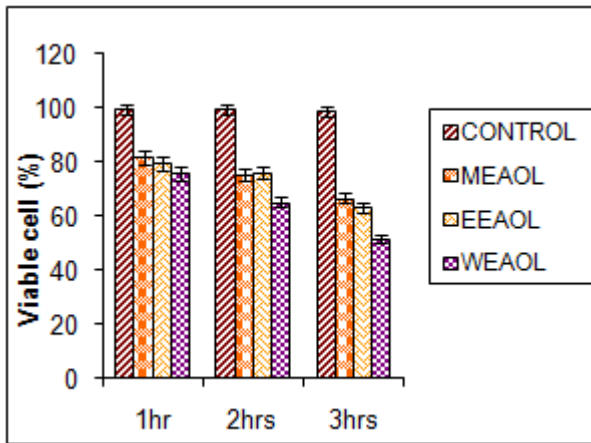


Fig-2. The bar diagram shows the effect of MEAOL, EEAOL, and WEAOL on in vitro short term cytotoxicity against S-180 cells. Data are expressed as mean \pm SEM. Probability values are given in asterisks. * indicates $P < 0.05$, ** indicates $p < 0.01$; values are taken in respect of tumour control.

The cytotoxic activity of MEAOL, EEAOL and WEAOL against S-180 cells is shown in Fig-2. The MEAOL, EEAOL and WEAOL exhibited a time-dependent cytotoxic effect on the S-180 cells at the concentration of 100 μ g/ml. Less numbers of viable S-180 cells were observed in MEAOL, EEAOL and WEAOL treated groups compared to S-180 control (tumour control) group. Among the three extracts, WEAOL showed maximum cytotoxic effect after three hours of treatment.

4. Discussion

The present study showed that MEAOL, EEAOL and WEAOL had prominent antimetabolic activity. The maximum percentage of root tips cells of the treated groups were observed to be in prophase indicating inhibition of transition from prophase to metaphase and subsequent phases. Maximum numbers of non-dividing cells were observed in the extract treated group compared to the control group. The extracts of *Anacardium occidentale* leaf seem to stay at prophase stage of cell division. Mitotic index of root tips of *Allium cepa* treated with EEAOL, MEAOL and WEAOL showed significantly lower mitotic index compared to control group, among the three extracts the EEAOL showed the highest degree of suppression of cell division. A group of anticancer drugs competitively inhibit enzyme that participates in DNA and RNA synthesis, and thus cytotoxic drugs suppress the cell cycle [19]. Logically, they may possess a greater toxic effect on rapidly dividing cells such as malignant and myeloid cells [20]. *Anacardium* is a medicinal plant that contains various flavonoids, alkaloids, etc. These chemicals may exhibit antimetabolic property [21], which may suppress the cell division of another species like *Allium cepa*.

In the present study the cytotoxic property of MEAOL, EEAOL, and WEAOL was assayed on S-180 cells by trypan blue dye exclusion technique. The extracts were also effective in decreasing the cell viability of Sarcoma-180 cells. The result showed that WEAOL exhibited more cell death compared to MEAOL and EEAOL in time-dependent

manner. The extracts may possess some active components, which showed prominent anti-tumour effect. Anti-tumour drugs that interact with microtubules and tubulin are known to block mitosis and induce cell death by apoptosis [22]. The extracts may mediate the effects through other mechanisms also. Those include restriction in RNA and or DNA synthesis, DNA damage, loss of membrane integrity, breakdown of cytoskeleton etc. [23]. Moreover the active compounds present in extracts may bind with different cell proteins, which are responsible for cell division and may make them inactive [24].

Apoptosis is a critical molecular target for the prevention of cancer [25]. The potential use of *Anacardium occidentale* leaves as therapeutic agent holds great hope for the isolation of one or more cytotoxic chemicals from crude extracts and the judicious use of such chemicals can control the progression of cancer and also can prevent the formation of tumour in susceptible individuals.

Declaration of Interest

Authors declare that there are no conflicts of interests.

Acknowledgement

The authors are thankful to Vidyasagar University, Midnapore for providing the facilities to execute these studies.

References

- [1] Alison MR. Cancer. In: eLS. John Wiley & Sons Ltd: Chichester. 2001. 10.1038/npg.els.0001471; doi: 10.1038/npg.els.
- [2] Varma PS, Agarwal VK. Cell biology, Genetics, Molecular biology, S. Chand & Company Ltd, New Delhi, 2002; 277-287.
- [3] Iwasaki S., Antimetabolic agents; Chemistry and recognition of tubulin molecule, *Med Res Rev*, 1993;13: 183 – 198.
- [4] Fonrose X, Ausseil F, Soleilhac E, Masson V and David P, Parthenolide inhibits tubulin carboxypeptidase activity, *Cancer Res*, 2007; 67: 3371 – 3378.
- [5] Mollinedo F, Gajate C. Microtubules, microtubule-interfering agents and apoptosis. *Apoptosis*, 2003; 8: 413-450.
- [6] Naem Q, Bashir AC, Altaf D. Phytochemical study of aerial parts of *Lantana camara* for the pharmacological active compounds. *J App Pharm*, 2009; 1(1):19-26.
- [7] Gonçalves JL, Lopes RC, Oliveira DB, Costa SS, Miranda MM, Romanos MT, Santos NS and Wigg MD. In vitro anti-rotavirus activity of some medicinal plants used in Brazil against diarrhea. *J Ethnopharmacol*, 2005;99:403-7.
- [8] Konan NA, Bacchi EM. Antitumorogenic effect and acute toxicity of a hydroethanolic extract from the cashew (*Anacardium occidentale* L) leaves. *J Ethnopharmacol*, 2007; 112:237-242.
- [9] Kamtchouing P, Sokeng SD, Moundipa, PF, Watcho P, Jatsa HB and Lontsi, D. Protective role of *Anacardium occidentale* extract against

- streptozotocin-induced diabetes in rats. *J. Ethnopharmacol.*,1998; 62:95-9.
- [10] Andrade LF, Campos JMS and Davide LC. Cytogenetic alterations induced by SPL (spent potliners) in meristematic cells of plant bioassays, *Ecotoxicol Environ Saf*, 2008; 71: 706–710.
- [11] Leme DM and Marin-Morales MA. Chromosome aberration and micronucleus frequencies in *Allium cepa* cells exposed petroleum polluted water – a case study. *Mutat Res*, 2008; 650:80–86.
- [12] Havey MJ, Rabinowitch HD, Currah L. Genome organization in *Allium*. In *Allium Crop Science*, (Eds.). Recent Advances, CABI Publishing, United Kingdom, 2002. 59-79.
- [13] Kura's M, Nowakowska J, S' liwin' ska E, Pilarski R, Ilasz R, Tykarska T, Zobel A and Gulewicz K. Changes in chromosome structure, Mitotic activity and nuclear DNA content from cells of *Allium test* induced by bark water extract of *Uncariatomentosa* (Willd.) DC., *J Ethnopharmacol*, 2006; 107:211–221.
- [14] Fiskesjo G, *Allium* test for screening chemicals; evaluation of cytological parameters. In *Plants for Environmental Studies*, CRC, Lewis Publishers, New York, 1997. (Eds.) 307-333.
- [15] Ramakrishna Y, Manohar AL, Mamata P, Shreekanth KG, Plants and novel antitumor agents: A review. *Indian Drugs*, 1984; 21:173-185.
- [16] Justo GZ, Duran N, Queiroz MLS, Myelopoietic response in tumor-bearing mice by an aggregated polymer isolated from *Aspergillusoryzae*. *Eur J Pharmacol*, 2000; 388: 219–226.
- [17] Pardesi Goldee S, Gadgoli C, Vaidya Madhav D, Hasni Hamid Y, More Babita H and Bhuskat Pallavi P. Preliminary studies on antimetabolic and anti-cancer activity of *Calotropis gigantean*. *Pharmacologyonline*, 2008; 1: 38-47.
- [18] Dongre SH, Badami S, Natesan S, Raghu Chandrasekhar H Antitumor activity of methanol extract of *Hypericum hookerianum* against Ehrlich Ascites Carcinoma in Swiss Albino mice. *J Pharmacol Sci*, 2007; 103:354-359.
- [19] Ravi Rajagopalan, PT, Zhiquan Z, McCourt L. Interaction of dihydrofolate reductase with methotrexate: Ensemble and single-molecule kinetics. *Proceedings of the National Academy of Sciences*, 2002; 99:21.
- [20] Johnston A, Gudjonsson J, Eli S, Hekla S . The antiinflammatory action of methotrexate is not mediated by lymphocyte apoptosis, but by the suppression of activation and adhesion molecules. *Clinical Immunology*, 2005; 114 (2): 154–63.
- [21] Kubo I, Ochi M, Vieira PC, Komatsu S., Antitumor agents from the cashew *Anacardium occidentale* apple juice. *Journal of Agricultural and Food Chemistry*, 1993; 41:1012–1015.
- [22] Jordan MA. Mechanism of action of antitumor drugs that interact with microtubules and tubulin. *Curr Med Chem Anticancer Agents*, 2002; 2: 1-17.
- [23] Awad AB, Downie D, Fink CS. Inhibition of growth and stimulation of apoptosis by Bsitosterol treatment of breast cancer MDA-MB-231 cells in culture. *Int J Mol Med* 2000; 5:541- 5.
- [24] Awad AB, Gan Y, Fink CS. Effect of Bsitosterol, a plant sterol on growth, protein phosphatase 2A and phospholipid D in LNCaP cells. *Nut Cancer*, 2000; 36:74-8.
- [25] Thenmozhi A, Nagalakshmi A, Mahadeva R. Study of Cytotoxic and Antimetabolic Activities of *Solanum nigrum* by Using *Allium cepa* Root Tip Assay and Cancer Chemo preventive Activity Using MCF-7- Human Mammary Gland Breast Adenocarcinoma Cell Lines. *International Journal of Science & Technology*, 2011; 2250-141X Vol. 1. 2.

Author Profile

Pralay Maity (Research student): Completed M.Sc in Human Physiology from Vidyasagar University, Midnapore in 2010. He is currently working as Research Scholar and pursuing his doctoral degree programme in the Department of Human Physiology with Community Health, Vidyasagar University, Midnapore, West Bengal, India.

Madhubanti Bepari (Research student): Completed M.Sc in Human Physiology from Vidyasagar University, Midnapore in 2010. She is currently working as a UGC Research Fellow and pursuing her doctoral degree programme in the Department of Human Physiology with Community Health, Vidyasagar University, Midnapore, West Bengal, India.

Dr. Sujata Maiti Choudhury (Research Guide): Working as an Associate Professor in the Department of Human Physiology with Community Health, Vidyasagar University, Midnapore, West Bengal, India.