



(E)

Figure 2: The activity of various antioxidant enzymes, including catalase represented by a graph (A), the Mn/FeSOD and Cu/Zn SOD represented by (B) and (C) respectively; similarly the concentration TBARS shown by (D) and the concentration of Nitric Oxide shown by (E) in normal control subjects and in patients with diabetes of family 1 and family 2.

We evaluated the levels of antioxidant enzymes in maternally inherited type 2 diabetes mellitus and in a healthy control subjects. The results of this study, clearly corroborates the oxidative stress in both family 1 and family 2. Catalase is the important antioxidant enzyme found nearly in all cells [40] which converts hydrogen peroxide into water molecule and oxygen [41]. Hydrogen peroxide if not removed from the biological samples it may convert into hydroxyl radicals, which may impart more oxidative stress. Hydroxyl radicals are the potent free radical, which is reported as the deadliest free radical which has ever been found. Hence catalase plays an important role in free radical scavenging [42]. Statistical analysis affirms that not only in family 1 but also in family 2 there is a significant decrease in the activity of catalase enzyme in both the diabetic families. This is in accordance with various other studies [43,44]

Like the catalase enzyme, Manganese/ Iron Superoxide Dismutase and Copper/ Zinc containing Superoxide Dismutase also plays a central role in the defence mechanism against the free radicals damage [45]. The main function of the Superoxide Dismutase enzyme is to eradicate superoxide anion which can cause damage to cells, cellular membranes, DNA and other biological organelles. Superoxide Dismutase converts superoxide anion radical into the hydrogen peroxide molecule [46,47]. Many studies showed that there is decreased activity of Mn/Fe SOD and Cu/Zn SOD in type 2 diabetes mellitus than normal control samples [48]. This study also shows a significant decrease in the activity of Mn/Fe SOD and Cu/Zn SOD in patients with type 2 diabetes.

Malonaldehyde formation is an indication of increased lipid peroxidation in diabetes mellitus [49]. It is used to determine the oxidative stress through determining the balance between free radicals and antioxidant enzymes [50]. We analysed the significant increase in the concentration of malonaldehyde in both diabetic families. The increased concentration of malonaldehyde indicates an oxidative stress in patients with diabetes. Various other studies showed an increase in oxidative stress in diabetes mellitus [43,51,52].

Statistics showed somewhat similar results for the concentration of nitric oxide to those of malonaldehyde. Nitric oxide may react with superoxide anion and can form peroxynitrite molecules, which can harm cellular mechanism. Hence, it is very essential to maintain the concentrations of

free radicals like nitric oxide and superoxide radicals [53]. Although, the life span of nitric oxide is very less, increased concentration can induce an oxidative stress and may play a pivotal role in diabetes mellitus [54]. The present study, exhibited significant increase in the concentration of nitric oxide in both the families of diabetic patients. Certain other studies have also demonstrated an increase concentration of nitric oxide in patients with diabetes than control [55,56].

Table 2 : A Statistical analysis of various antioxidant enzymes, lipid peroxidation and nitric oxide.

ENZYMES	GROUPS	MEAN±SD	P-VALUE
CATALASE (Units/Mg protein/ML)	CONTROL	0.3659±0.0931	
	FAMILY1	0.1805±0.0752	<0.01
	FAMILY2	0.2160±0.0400	<0.01
Mn/Fe SOD (Units/Mg protein/ML)	CONTROL	0.0376±0.0074	
	FAMILY1	0.0244±0.0064	<0.01
	FAMILY2	0.0215±0.0115	<0.05
Cu/Zn SOD (Units/Mg protein/ML)	CONTROL	0.1325±0.0308	
	FAMILY1	0.0379±0.0203	<0.001
	FAMILY2	0.0532±0.0305	<0.01
MDA (nM × 10 ⁻³ /ML)	CONTROL	0.2048±0.1247	
	FAMILY1	0.4487±0.1656	<0.05
	FAMILY2	0.3767±0.1022	<0.05
NO (nM × 10 ⁻² /ML)	CONTROL	0.1800±0.0303	
	FAMILY1	0.2483±0.0611	<0.05
	FAMILY2	0.2733±0.0592	<0.05

Mitochondrial DNA lacks histone proteins and DNA repair mechanisms, which increases the chances of its damage by free radicals [57]. Colossal work on mitochondrial DNA exhibited its role in maternally inherited type 2 diabetes mellitus [21,22,23]. For last few decades, researchers have been working on mitochondrial DNA to identify the various mitochondrial DNA mutations, which can play a pivotal role in the pathogenesis of type 2 diabetes mellitus. Over more than 40 different mitochondrial DNA mutations associated with type 2 Diabetes Mellitus have been identified yet [58]. In this study, the absence of 3243 A/G mutation has been observed in the selected families (Fig: 2), this was according to the study of Naveed AK et.al. [59].

5. Conclusion

This study cogitated on oxidative stress and 3243 A/G mitochondrial DNA mutation in two families with a history of maternally inherited type 2 diabetes mellitus. Decrease activities of various antioxidant enzymes like catalase, Mn/Fe SOD, Cu/Zn SOD with increase concentration of nitric oxide and lipid peroxidation providing a straight forward evidence of oxidative stress in both diabetic families. Increased oxidative stress can cause other complications like DNA and cellular damage in the patients. Therefore, monitoring oxidative stress in patients with maternally inherited type 2 diabetes mellitus could be of utmost importance to prevent these complications. The undertaken study did not observe a 3243A/G mutation in patients with a history of maternally inherited type 2 diabetes mellitus, other mutations might be present in selected families.

6. Acknowledgement

This work has been sanctioned by University Grant Commission, Regional Office Pune, Maharashtra, India. We would like to thank all the patients for their valuable contribution and co-operation for the study.

References

- [1] Wild S, Gojka R, Green A, Sciref R, King H. Global prevalence of diabetes estimates for the year 2000 and projection for 2030. *Diabetes Care* 2004;27:1047-1053.
- [2] World Health Organization (WHO). Facts and figures. http://www.who.int/diabetes/facts/world_figures/en/
- [3] Jean-Claude HENQUIN. Triggering and amplifying pathways of regulation of insulin secretion by glucose. *Diabetes* 2000;49: 1751-1760.
- [4] Cadenas E, Davies K J. Mitochondria free radical generation, oxidative stress and aging. *Free Radical Biology and Medicine* 2000;29:222-230.
- [5] Ide T, Tsutsui H, Kinugawa S, Utsumi H, Kang D, Hattori N, et al. Mitochondrial electron transport complex I is a potential source of oxygen free radicals in the failing myocardium. *Circulation Research* 1999;85:357-363.
- [6] Machlin L J and Bendich A. Free radical tissue damage: protective role and antioxidant nutrients. *The FASEB journal* 1987;6:441-445.
- [7] Halliwell B. Reactive species and antioxidants. Redox biology is a fundamental theme of aerobic life. *Plant physiology* 2006;141:312-322.
- [8] Keller J.N., Kindy M S, Holtsberg F W, Clair DKS, Yeb HC, Grmeyer A, et.al. Mitochondrial manganese superoxide dismutase prevents neural apoptosis and reduces ischemic brain injury: Suppression of peroxynitrite production, lipid peroxidation and mitochondrial dysfunction. *The Journal of Neuroscience* 1998;18:687-697.
- [9] Miki T, Nagashima K and Seino S. The structure and function of ATP sensitive potassium channel in insulin secreting pancreatic beta cells. *Journal of Molecular Endocrinology* 1999;22:113-123.
- [10] Craig T J, Ashcroft F M and Proks P. How ATP inhibits the open KATP channel. *J. Gen Physiol* 2008;132:131-144.
- [11] Ohkubo K, Yamano A, Nagashima M, Mori Y, Anzai K, Akehi Y, et al. Mitochondrial gene mutation in the tRNA Leu(UUR) Region and diabetes: Prevalence and Clinical Phenotypes in Japan. *Clinical Chemistry* 2001;47:1641-1648.
- [12] Lynn S, Wardell T, Johanson MA, Chinnery PF, Dally ME, Walkar M, et al. Mitochondrial Diabetes: investigation and identification of a novel mutation. *Diabetes* 1998;47:1800-1802.
- [13] Dongre U J, Meshram VG. Is mitochondrial DNA responsible for maternally inherited type 2 diabetes mellitus: A hypothetical review. *Int. J. Pharm. Sci. Rev. Res* 2014;28:179-187.
- [14] Min B and Ahn D.V. Mechanism of lipid peroxidation in meat and meat products-A review. *Food Sci Biotechnol* 2005;14:152-163.
- [15] HABIB S and ALI A. Biochemistry of nitric oxide. *Ind J Clin Biochem* 2011;26:3-17.
- [16] Flekac M, Skrha J, Higtartova J, Lacinova Z and Jarolimkova M. Gene polymorphism of superoxide dismutase and catalase in diabetes mellitus. *BMC Medical genetics* 2008;30: 1-9.
- [17] Dongre UJ & Meshram VG. A statistical analysis of antioxidants and biochemical parameters in maternally inherited type 2 diabetes mellitus. *Asiatic Journal of Biotechnology Resources* 2014;4:94-96.
- [18] Dongre UJ & Meshram VG. Evaluation of glutathione dependant antioxidant enzymes in maternally inherited type 2 diabetes mellitus. *J. Pharm. Sci & Res.* 2015;7:137-140.
- [19] Moraes, C.T., F. Ciacci, E. Bonilla, C. Jansn, M. Hirano, N. Rao, R.E. Lovelace, L.P. Rowland, E.A. Schon, and DiMauro. Two novel pathogenic mitochondrial DNA mutation affected organelle number and protein synthesis. Is the tRNA_{Leu}(UUR) gene an etiologocal hot spot?. *Journal of Clinical Investigation* 1993;92: 2906-2915.
- [20] Goto, Y., I. Nonaka, and S. Horai. A mutation in the tRNA Leu(UUR) gene associated with the MELAS subgroup of mitochondrial encephalomyopathies. *Nature* 1990;348: 651-653.
- [21] Takashi K, Kodawari H, Mori Y, Tobe K, Sakuta R, Suzuki Y, et. al. A subtype of diabetes mellitus associated with a mutation of mitochondrial DNA. *N Engl J Med* 1994;330: 962-968.
- [22] Ohkubo K, Yamano A, Nagashima M, Mori Y, Anzai K, Akehi Y, et al. Mitochondrial gene mutation in the tRNA Leu(UUR) Region and diabetes: Prevalence and Clinical Phenotypes in Japan, *Clinical Chemistry*, 2001;47:1641-1648.
- [23] Duraisamy P, Elango S, Vishwananadha VP, Balamurugan R. Prevalence of mitochondrial t RNA gene mutation and their association with specific clinical phenotypes in patients with type 2 diabetes mellitus of Coimbatore. *Genetic testing and Molecular Biomarkers* 2010;14: 49-55.
- [24] Aebi H, wyss SR, Scherz B, Gross J. Properties of erythrocyte catalase from homozygotes and heterozygotes for Swiss type acatalasemia. *Biochemical Genetics* 1976;14:791-807.

- [25] Crouch RK, Gandy SE, Kimsey G, Galibraith RA, Galibraith GMP, Buse MG. The inhibition of islets superoxide dismutase by daibetogenic drugs. *Diabetes* 1981;30:235-241.
- [26] Ken CF, Lee CC, Duan KJ, Lin CT. Unusual stability of manganese superoxide dismutase from a new species, *tatumella Ptyseas* CT: its gene structure, expression and enzymatic properties. *Protein Expression and Purification* 2004;40:42-50.
- [27] Marklund S, Marklund G. Involvement of the superoxide anion radical in the autoxidation of pyrogallol and a convenient assay for superoxide dismutase. *Eur. J. Biochem* 1974;47:469-474.
- [28] J. Stocks, T.L.Dormandy. Auto oxidation of Human red cell lipids induced by hydrogen peroxide, *British Journal of Heamatology* 1971, 20:95-111.
- [29] Green LC, Wagner DA, Glogowski J, Skipper PL, Wishnok JS, Tannenbaum SR. Analysis of nitrate, nitrite and [15N] nitrate in biological fluids. *Anal Biochem* 1982;126:131-138.
- [30] Lowery O.H., N.J. Rosebrugh, A.L. Farr and R.J. Randall. Protein measurement with folin-phenol reagent. *J. Biol Chem* 1951: 265.
- [31] Van Den Ouweland JMW, Lunkes HHPJ, Ruitenbeek W, Sandkuij LA, De Vijlder MF, Struyvenberg PAA, et.al. Mutation in mitochondrial tRNA Leu (UUR) gene in a large pedigree with maternally transmitted type 2 diabetes mellitus and deafness. *Nature Genetics* 1992;1: 368-371.
- [32] Cadenas E, Davies KJ. Mitochondrial free radical generation, oxidative stress and aging. *Free Radical Biology and Medicine* 2000;3: 222-230.
- [33] Griesmacher A, Kindhouser M, Andert S E, Schreiner W, Toma C, Knoebl P, et. al. Enhanced serum levels of thiobarbituric acid reactive substance in diabetes mellitus. *American Journal of Medicine* 1995;98:469-475.
- [34] Giron MD, Salto R, Gonzalez Y, Giron J A, Nieto N, Periago J, et. al. Modulation of hepatic and intestinal glutathione s transferase and other antioxidant enzyme by dietary lipids in streptozotocin diabetic rats. *Chemosphere* 1999;38: 3003-3013.
- [35] Ramanathan M, Jiaswal Ak and Bhattacharya SK. Superoxide dismutase, catalase and glutathione peroxidase activities in the brain of strptozotocin induced diabetic rats. *Ind. J. Exp. Biol* 1999;37:182-183.
- [36] Merzouk S, Hichami A, Madani S, Merzouk H, Berrouiguet AY, Prost J, Moutairou K, Chabane-Sari and Khan NA. Antioxidant status and levels of different vitamins determined by high performance liquid chromatography in diabetic subjects with multiple complications. *Gen. Physiol. Biophys* 2003;22:15-27.
- [37] Kangralkar V.A., Patil SD, Bandivadekar RM. Oxidative stress and diabetes: A review. *International Journal of Pharmaceutical applications* 2010;1: 38-45.
- [38] Godin DV, Wohaeib SA, Garnett ME, Goumeniouk AD. Antioxidant enzyme alterations in experimental and clinical diabetes. *Mol Cell Biochem* 1988; 84:223-233.
- [39] Noda Y, Mori A, Packer L. Gliclazide scavengers hydroxyl, superoxide and nitric oxide radicals: an ESR study. *Mol Pathol. Pharmacol* 1999; 96:115-124.
- [40] P. Chelikani, I Fita and P.C. Loewen. Diversity of structures and properties among catalases. *Cellular and Molecular Life Sciences* 2004;61:192-208.
- [41] Takemoto K, Tanaka M, Iwata H et.al. Low catalase activity in blood is associated with the diabetes caused by alloxan. *Clinica Chimica Acta* 2009;407:43-46.
- [42] Tiedge M, Lortz S, Modey R, Lenzen S. Complimentary action of antioxidant enzymes in the protection of bioengineered insulin producing RIN m5F cells against the toxicity of reactive oxygen species. *Diabetes* 1998; 47:1578-1585.
- [43] Sundaram RK, Bhaskar A, Vijayalingam S, Vishwanathan M, Mohan R, Shanmugasundaram. Antioxidant status and lipid peroxidation in type 2 diabetes mellitus with and without complications. *Clinical Science* 1996;90:255-260.
- [44] Marica JB, Veljko B, Jadranka B, Zeljko M, Ivan J, Zeljko R. Impact of glycemic control on antioxidant enzyme activity in patients with type 2 diabetes mellitus. *Diabetologia Croatica* 2004;33:131-135.
- [45] Zelko IN, mariani JJ, Folz RJ. Superoxide dismutase multienzyme family: a comparison of the Cu/Zn SOD (SOD1), MnSOD (SOD2) and EC SOD (SOD3) gene structures evolution and expression. *Free Radical Biology and Medicine* 2002; 33:337-349.
- [46] Faraci FM, Didion SP. Vascular protection: Superoxide dismutase isoform I the vessel wall. *Atherosclerosis Thrombosis and Vascular Biology* 2004;24:1367-1373.
- [47] Wang X, Tao L and Hai CX. Redox-regulating role of insulin: the essence of insulin effect. *Molecular and Cellular Endocrinology* 2012;349:111-127.
- [48] Hamed S, Brenner B, Aharon A, Daoud D, Roguin A. Nitric oxide and SOD modulate endothelial progenitor cell function in type 2 diabetes mellitus. *Cardiovascular Diabetology* 2009;8:56.
- [49] Gallou G, Ruelland A, Legars B, MAugender D. Allanic H and Cloarec L. Plasma malonaldehyde in type 1 nad type 2 diabetic patients. *Clin.Chim. Acta* 1993;214:227-234.
- [50] Altomare E, Vendemiale G, Chicco D, Procacci V and Cirelli F. Increased lipid peroxidation in type 2 poorly controlled diabetic patients. *Diabetes metab* 1992;18:264-271.
- [51] Pasaoglu H, Banu S, Bukan N. Lipid peroxidation and resistance to oxidation n patients with type 2 diabetes mellitus. *Tohoku J. Exp. Med* 2004;203:211-218.
- [52] Gupta S, Chari S. Proxidant and antioxidant status in patients of type 2 diabetes mellitus IHD. *Indian Journal of Clinica Biochemistry* 2006;21:118-122.
- [53] Guzik TJ, West NEJ, Pillai R, Taggart DP, Channon KM. Nitric Oxide modulates superoxide release and peroxynitrite formation in human blood vessels. *Hypertention* 2002;39:1088-1094.
- [54] Giugliano D, Ceriello A. Paolisso G. Oxidative stress and diabetic vascular complications. *Diabetes Care* 1996;96:257-267.
- [55] Bhatia S, Shukla R, MAdhu V, Kaur J, Prabhu KM. Antioxidant status, lipid peroxidation and nitric oxide end products in patients of type 2 diabetes mellitus with neuropathy. *Clinical biochemistry* 2003;36:557-562.
- [56] Maejima K, Nakano S, Himeno M, Ichi S T, Makishi H, Ito T, Nakagawa A, et. al. Increased basal level of plasma nitric oxide in type 2 diabetic

subjects:Relationship to microvascular complications.
Journal of Diabetes and its complications 2001;15:135-143.

- [57] Chistiakov DA, Sobenin IA, Bobryshev YV, Orekhov AN. Mitochondrial dysfunction and mitochondrial DNA mutation in atherosclerotic complications in diabetes. World J. Cardiol 2012;5:148:156.
- [58] Lamson, D.W., and S.M. Plaza. Mitochondrial factors in the pathogenesis of diabetes: a hypothesis for treatment. Alternative Medicine Review 2002;7: 94-111.
- [59] Naveed AK, Wahid M, Naveed A. Mitochondrial tRNA Leu (UUR) gene mutation and maternally inherited diabetes mellitus in Pakistani population. International Journal of Diabetes Mellitus 2009;1: 11-15.

