

# Hypolipidemic and Antiatherogenic Effects of Aqueous Extract of Libyan Propolis in Lead Acetate Intoxicated Male Albino Mice

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**Abstract:** *Hyperlipidemia is a major cause of atherosclerosis and atherosclerosis-associated conditions. Coronary artery disease is the epidemic of modern civilization in which dyslipidemia contributes significantly to its pathogenesis. Flavonoids and various phenolics are the most important pharmacologically active constituents in propolis capable of scavenging free radicals and thereby protecting lipids from being oxidized or destroyed during oxidative damage. The aim of this study was to investigate the hypolipidemic and antiatherogenic effects of aqueous extract of Libyan propolis in lead acetate intoxicated male albino mice. In this study, Thirty two adult male albino were used for this study and divided into four groups. The first group was control group, the 2nd was the propolis group orally received propolis (200 mg/kg body wt), the 3rd was the experimental and received lead acetate (500 mg /kg diet), the 4th one co-administered lead acetate (500 mg/kg diet) with propolis (200 mg/kg body wt) daily for 30 days. Blood samples were obtained for assessment of serum cholesterol, triglycerides, HDL, LDL, parameters. In lead treated animals, the serum cholesterol, triglycerides, HDLc, LDLc, VLDL, Castelli's Risk Index I (TC/HDLc), Castelli's Risk Index II (LDLc/HDLc), Atherogenic Coefficient {(TC-HDLc)/HDLc} and Atherogenic Index of Plasma{ (AIP)= log(TG/HDLc)} parameters were increased and serum HDLc was decreased. Co-administration of propolis significantly improved of lipids profile parameters the ratios based on these parameters. Serum cholesterol, triglycerides, non HDLc, LDLc, VLDL, Castelli's Risk Index I (TC/HDLc), Castelli's Risk Index II (LDLc/HDLc), Atherogenic Coefficient {(TC- HDLc)/HDLc} and Atherogenic Index of Plasma{ (AIP)= log(TG/HDLc)} parameters were significantly declined and serum HDLc was elevated. It can be concluded that, the lead had adverse effects on serum lipids profile parameters and the ratios based on these parameters. Propolis showed hypolipidemic and antiatherogenic effects in lead acetate intoxicated male albino mice. So, the populations of high risk to lead should be advised to take propolis.*

**Keywords:** Antiatherogenic, Male albino mice, Hypolipidemic effect, Lead acetate, Libyan propolis.

## 1. Introduction

Heavy metals like lead, cadmium etc. have very long half life and are severely toxic at a very low dose [1]. Lead is a natural stable element and is bioaccumulative in nature. It is an environmental poison of significance to the grazing livestock and a potential public health hazard, as it is excreted in milk [2]. It represents an exclusive case (among cumulative metal contaminants) because of its ubiquitous presence in the environment and easy recognition of its major sources, which give rise to environmental pollution [3]. It has been used in medicines, paintings, pipes, ammunition and in more recent times in alloys for welding storage materials for chemical reagents [4]. It is an environmental pollutant that causes damage to biological systems [5]. Several reports have indicated that lead can cause neurological, histopathological, hematological, gastro-intestinal, reproductive, circulatory and immunological pathologies, all of them related to the dose and the duration of time of lead exposure [6 - 9]. It also produces high blood pressure that increases the risk of heart attack [10]. Toxicities due to lead exposure have been attributed to the ability of lead to induce oxidative stress through the generation of reactive oxygen species [11]. Elevation of total cholesterol, triglycerides and lipoproteins such as (LDL, VLDL) levels and reduction in HDL - CHOL level were recorded in serum of lead intoxicated rats [5 & 12].

Hyperlipidemia is a condition which characterized by abnormal elevation of lipid such as (triglyceride and cholesterol) and lipoproteins such as (LDL, VLDL) levels in the blood [13]. Scientific evidence indicates that oxidation of low density lipoprotein (LDL), which carry cholesterol in the blood stream plays an important role in the development of atherosclerosis, the underlying disorder leading to heart attacks and ischemic strokes [14 & 15]. Hyperlipidemia is a major cause of atherosclerosis and atherosclerosis-associated conditions, such as coronary heart disease (CHD) [15], ischemic cerebrovascular disease, and peripheral vascular disease [16]. Coronary artery disease is the epidemic of modern civilization in which dyslipidemia contributes significantly to its pathogenesis [17]. The basic pathogenesis of atherosclerosis involves an insult to the endothelial and smooth muscle cells of the arterial wall by various harmful factors such as viral infection, mechanical damage and dislipidemia, especially abnormal oxidized low density lipoproteins.[16]. It is important to reduce excessive cholesterol and LDL-cholesterol oxidation to low levels, which represent adequate mechanisms for maintenance of normal body functions [18].

Several experimental studies in various laboratories are underway, to study the prophylactic effect of various natural antioxidant compounds against toxic metals. Herbs are generally considered safe and proved to be effective against various human ailments and their medicinal uses have been

gradually increasing in developed countries [19]. Natural antioxidants strengthen the endogenous antioxidants defenses from reactive oxygen species and restore the optimal balance by neutralizing the reactive species [20].

Propolis is resinous natural product collected from cracks in the bark of trees and leaf buds which are enriched with salivary enzymes of honey bees. It has more than 180 compounds including polyphenols, flavonoids, phenolic acids and its esters [21-23]. Melatonin and caffeic acid phenethyl ester are compounds of honey bee propolis, that were recently found to be potent free radical scavengers and antioxidants [24]. Many flavonoids are known to be antioxidants, and several of these, such as quercetin which has been identified as constituents of propolis have been shown to be inhibitors of low density lipoprotein oxidation [25]. It is believed that propolis exerts a therapeutic or preventive effect in inflammation, heart disease, and even diabetes mellitus and cancer and there have been several reports indicating various biological activities of propolis and its constituents, such as anticancer [26 & 27], antioxidant, anti-inflammatory and antibiotic activities [28]. The actual ingredients in individual propolis products may differ significantly, according to a number of variables including the type of bees that produced the propolis, time of the year and the geographic location of the hives [29]. Flavonoids and various phenolics are the most important pharmacologically active constituents in propolis capable of scavenging free radicals and thereby protecting lipids from being oxidized or destroyed during oxidative damage [30]. The evidence reporting the hypolipidemic and anti-atherogenic effects of propolis in lead acetate intoxicated male albino mice are hardly found. So, the present work aimed to evaluate hypolipidemic and antiatherogenic effects of propolis in lead acetate intoxicated male albino mice.

## 2. Materials and Methods

### 2.1. Chemicals

Lead acetate was purchased from Sigma Chemical Co., USA. Lead acetate was given in diet as 500 mg/kg diet daily [31] for 30 days. Propolis samples were collected from different localities of Surman city, west Libya. Aqueous propolis extract was prepared according to the method of El-khayat *et al.* [32]. Briefly, propolis was kept dry and freeze-dried (-40°C) until used. Propolis samples were mixed with distilled water, heated gently and filtered through Whatman No:1 filter paper. The choice of the dose of propolis was based on the results of the previous studies, where the antioxidant effect of this agent was confirmed. Propolis was freshly prepared and administered to animals orally by gavage at a dose of 200 mg/kg body wt [33] once daily for 30 days.

### 2.2. Animals

Thirty two adult male albino mice (*Mus musculus*) weighting 25-30 g were used for this study. The animals were obtained from animal house unit in the Faculty of Pharmacy, Tripoli University, Libya. The animals were housed in plastic cages measuring about (29×15×12) cm, with about four mice per

cage. Floors of cages were covered with soft crushed wood shaving; all cages were washed two times per week with 70% alcohol throughout the period of the study. The animals were provided with tap water *ad libitum* and fed with the standard commercial chow. The animals were kept in the animal house of Faculty of Science, Alejelat, Zawia University in an air conditioned room with an optimum temperature of 25±2 °C, humidity (60-70%) and light/dark condition (12/12). The animal procedures were performed in accordance with Guide Lines for Ethical Conduct in the Care and Use of Animals.

### 2.3 Experimental Design

After one week of acclimation, the animals were randomized and divided into four groups (8 albino mice for each) as follow:

Group I (control group): provided with tap water and fed with normal diet.

Group II (propolis group): The animals received propolis (200 mg/kg body wt/day) orally by gavage daily for 30 days.

Group III (lead acetate treated group): The animals received 500 mg lead acetate/kg diet daily for 30 days.

Group IV (lead acetate/propolis co-administered): The animals received 500 mg lead acetate/kg diet concurrently with propolis (200 mg/kg body wt/day) orally by gavage daily for 30 days.

At the end of the experimentation and 24 hours after the last dose, all animals were sacrificed under light ether anesthesia, then rapidly dissected and subjected to the following examinations:

### 2.4 Biochemical Analysis

Blood samples were drawn by cardiac puncture. The sample was collected in clean dry tube and centrifuged at 3000 rpm for 15 minutes then, serum was separated and kept in a deep freezer at -20°C until biochemical measurements were carried out. Total cholesterol concentration was estimated according to Allain *et al.* [34], triglycerides concentration also by the method of Fossati and Prencipe [35] and HDL-cholesterol by Burstein *et al.* [36]. VLDL-cholesterol and LDL-cholesterol concentrations were estimated by using the Friedewald equation [37]. The atherogenic ratios were calculated as follows: Castelli's Risk Index (CRI-I) = TC/HDLc, Castelli's Risk Index (CRI-II) = LDLc/HDLc, Atherogenic Coefficient (AC)=(TC- HDLc) /HDLc and Atherogenic Index of Plasma (AIP)= log TG/HDLc.

### 2.5 Statistical Analysis

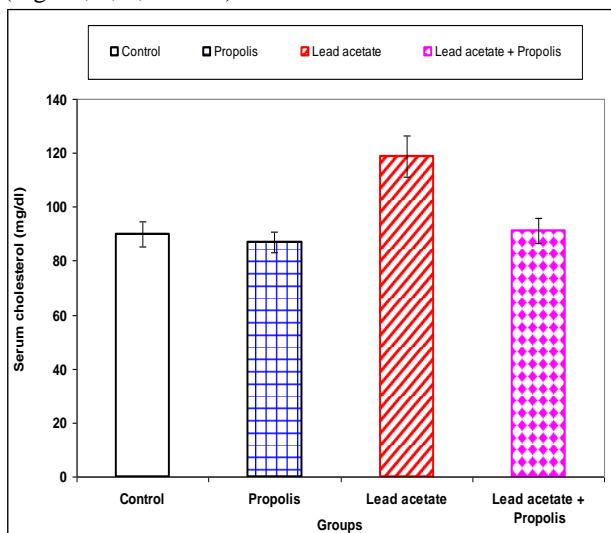
The values were presented as means ± SD of different groups. Differences between the mean values were estimated using one way ANOVA. The results were considered statistically significant when  $p < 0.05$ .

## 3. Results

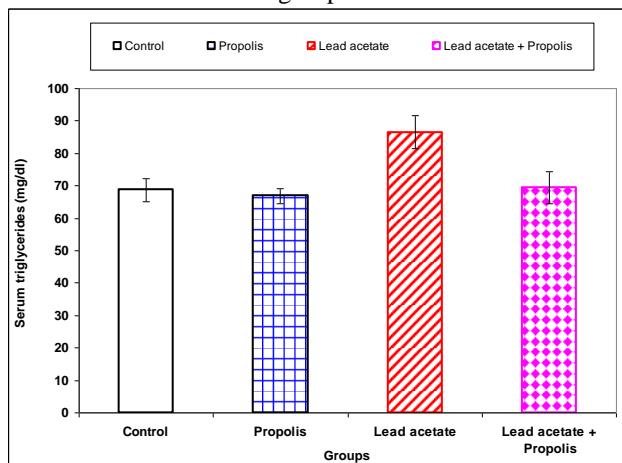
Lipid profile parameters in serum of the different groups are shown in Table 1. Mice that received lead acetate (500 mg/kg diet) daily for 30 days had significantly ( $p < 0.05$ ), increased the serum cholesterol, triglycerides, non HDLc, LDLc and

VLDL concentrations. Co-administration of lead acetate with propolis were significantly ( $p < 0.05$ ) prevented the changes recorded in serum cholesterol, triglycerides, non HDLc, LDLc and VLDL concentrations as compared with control group (Fig. 1, 2, 4 & 5). On the other hand, serum HDL cholesterol concentration of lead acetate treated mice was significantly ( $p < 0.05$ ) decreased as compared to the control mice (Fig. 3). Co-administration of lead acetate with propolis were significantly ( $p < 0.05$ ) prevented the changes recorded in serum HDLc concentration as compared with control group.

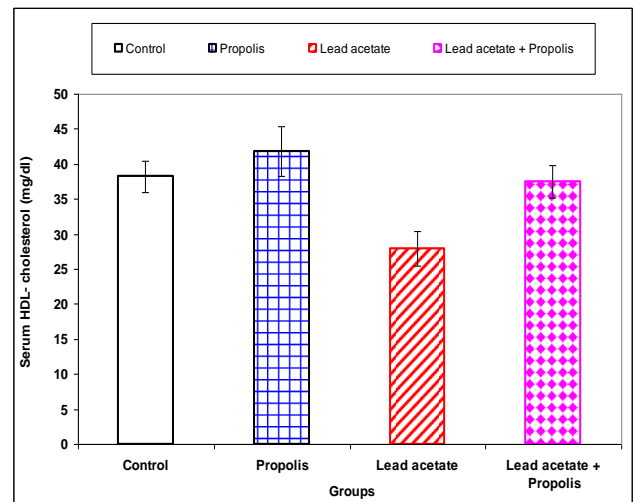
Table 2 showed the means and standard deviations for Castelli's Risk Index I (TC/HDLc), Castelli's Risk Index II (LDLc/HDLc), Atherogenic Coefficient{(TC-HDLc)/HDLc} and Atherogenic Index of Plasma{(AIP)= $\log(TG/HDLC)$ } in control group, propolis group, lead acetate treated group and albino mice group co-administrated of lead acetate with propolis. These ratios were elevated in lead acetate treated male albino mice group compared with the control group with statistically significant differences ( $p < 0.05$ ). Co-administration of lead acetate with propolis were declined these ratios with statistically significant differences ( $p < 0.05$ ), when compared with lead acetate group (Figs. 6, 7, 8, 9 & 10)



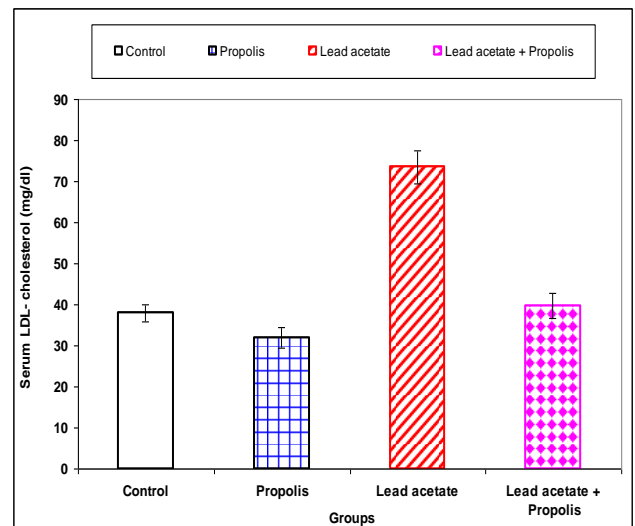
**Figure 1:** Serum cholesterol concentration in different groups.



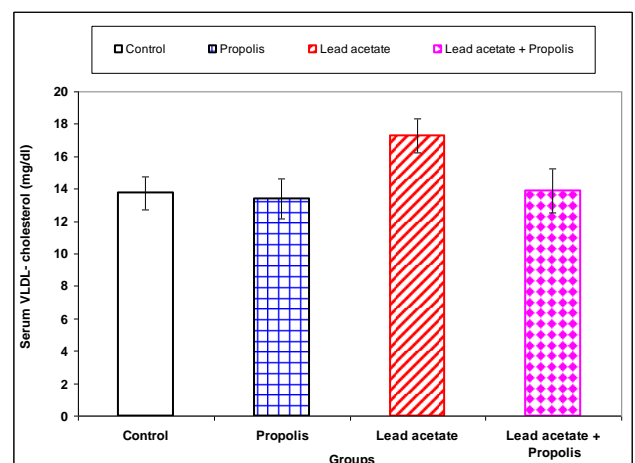
**Figure 2:** Serum triglycerides concentration in different groups.



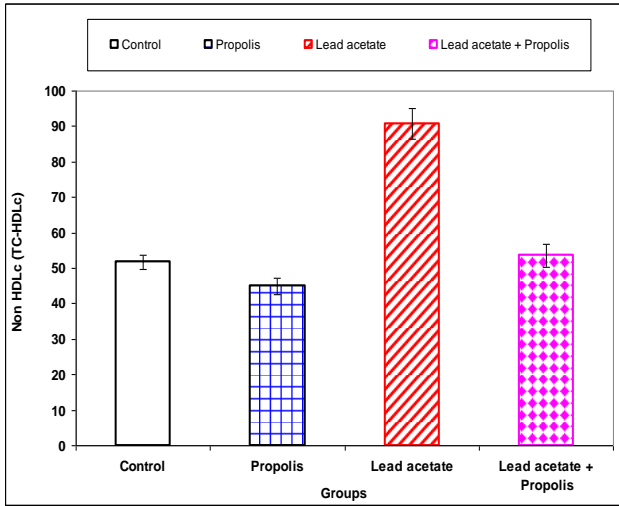
**Figure 3:** Serum HDL-cholesterol concentration in different groups.



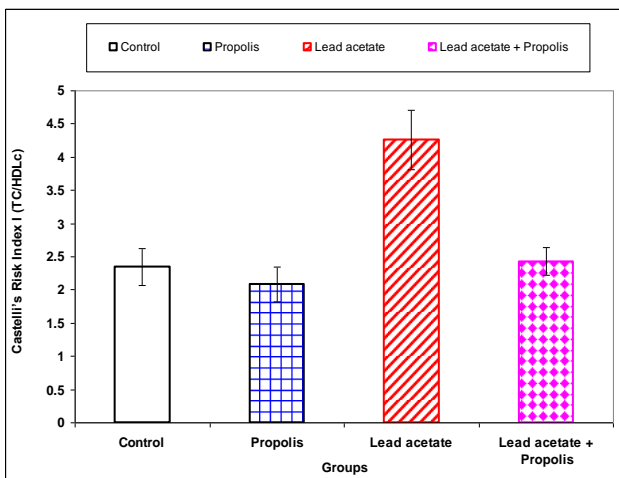
**Figure 4:** Serum LDL-cholesterol concentration in different groups.



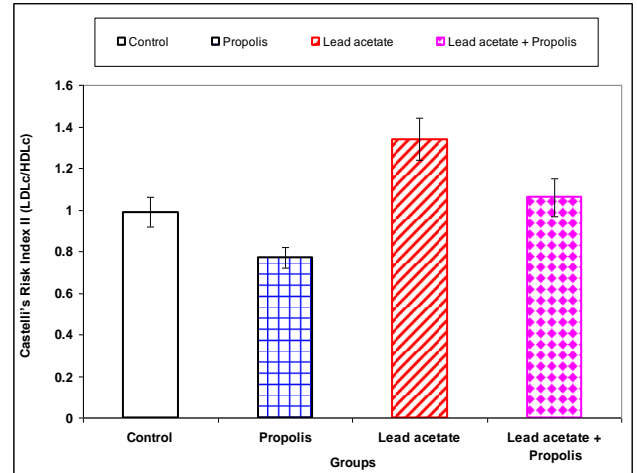
**Figure 5:** Serum VLDL-cholesterol concentration in different groups.



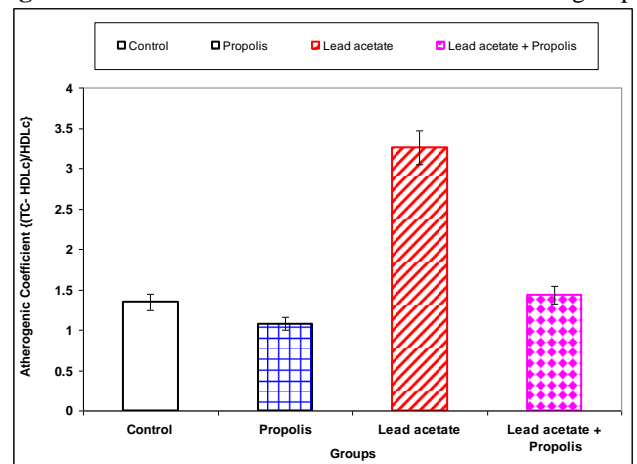
**Figure 6:** Serum non HDL-cholesterol( TC-HDLc) concentration in different groups.



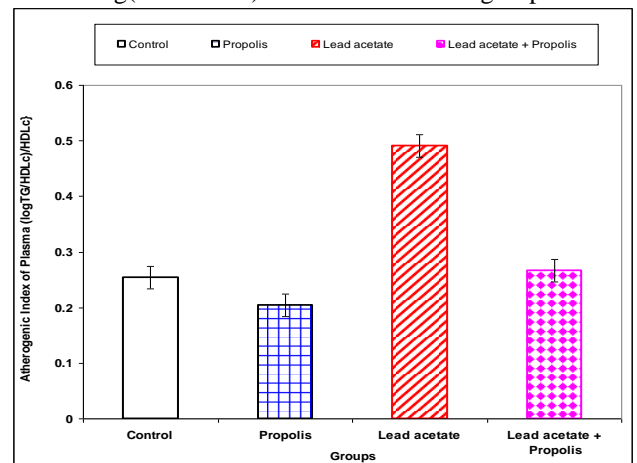
**Figure 7:** Cardiac Risk Ratio (Castelli's Risk Index I) TC/HDLC (LDLc/HDLC) in different animals groups



**Figure 8:** Castelli's Risk Index II in different animals groups.



**Figure 9:** Atherogenic Coefficient {(TC- HDLc)/HDLc} in log(TG/HDLC) in different animals groups



**Figure 10:** Atherogenic Index of Plasma(AIP)= different animals groups

**Table 1:** Effect of aqueous extract of propolis on lipid profile parameters of lead acetate treated male albino mice in different groups.

Parameters	Groups			
	Control	Propolis	Lead acetate	Lead acetate + Propolis
	Mean + SD	Mean + SD	Mean + SD	Mean + SD
Serum cholesterol (TC, mg/dl)	89.90 ±4.78	86.80 ±3.86	118.70 ±7.52 <sup>a</sup>	91.12 ±4.71 <sup>b</sup>
Serum triglycerides (TG, mg/dl)	68.70 ±3.49	66.90 ±2.38	86.50 ±5.13 <sup>a</sup>	69.40 ±4.88 <sup>b</sup>
Serum HDL- cholesterol (mg/dl)	38.20 ±2.28	41.80 ±3.60	27.90 ±2.47 <sup>a</sup>	37.50 ±2.37 <sup>b</sup>
Serum LDL- cholesterol (mg/dl)	37.96 ±2.01	31.96 ±2.40	73.50 ±4.10 <sup>a</sup>	39.74 ±3.11 <sup>b</sup>
Serum VLDL- cholesterol (mg/dl)	13.74±1.03	13.38±1.22	17.3±1.06 <sup>a</sup>	13.88±1.37 <sup>b</sup>
Non HDLc (TC-HDLc) (mg/dl)	51.7 ±2.00	45.00 ±2.30	90.80 ±4.30 <sup>a</sup>	53.62 ±3.20 <sup>b</sup>

a: Significant differences as compared with control group ( P < 0.05 ) .  
 b: Significant differences as compared with lead acetate treated group ( P < 0.05).  
 All data are mean of 6 individuals.

**Table 2:** Effect of aqueous extract of propolis on the ratios based on lipid profile parameters of lead acetate treated male albino mice in different groups

Parameters	Groups			
	Control	Propolis	Lead acetate	Lead acetate + Propolis
	Mean + SD	Mean + SD	Mean + SD	Mean + SD
Cardiac Risk Ratio (Castelli's Risk Index I) TC/HDLc	2.35±0.21	2.08±0.11	4.2±0.30 <sup>a</sup>	2.43±0.20 <sup>b</sup>
Castelli's Risk Index II (LDLc/HDLc)	0.99 ±0.07	0.77 ±0.05	1.34 ±0.10 <sup>a</sup>	1.06 ±0.09 <sup>b</sup>
Atherogenic Index of Plasma(AIP)= log(TG/HDLc)	0.254±0.019	0.204±0.018	0.491±0.022 <sup>a</sup>	0.267±0.021 <sup>b</sup>
Atherogenic Coefficient {(TC- HDLc)/HDLc}	1.35 ±0.10	1.08 ±0.08	3.26 ±0.21 <sup>a</sup>	1.43 ±0.11 <sup>b</sup>

a : Significant differences as compared with control group ( P < 0.05 ) .  
 b : Significant differences as compared with lead acetate treated group ( P < 0.05).  
 All data are mean of 6 individuals.

#### 4. Discussion

The present data indicated that cholesterol, triglycerides, LDLc and VLDL concentrations were significantly increased by lead acetate treatment, while HDL-c concentration was decreased in the serum. Several studies have shown that lead exposure induces alterations in serum lipid profiles [5, 12 & 38-40]. These results run parallel to those reported by Ghosh *et al.*, [38] who found that treatment of rats with lead acetate at a dose of 15 mg / kg body weight intraperitoneally (i.p) for a period of seven consecutive days caused alterations in the total cholesterol, triglyceride, HDLc, LDLc. Also, There was significant increase (p<0.05) in the serum total cholesterol, LDL CHOL and triglycerides in oral treated albino rats with lead acetate(740mg/kg body weight) daily for 28 days group compared to the normal control group [5]. Lowering levels of high density lipoprotein (HDL) was a contrary effect because high HDL levels have been shown to bear an inverse correlation with risks for atherosclerosis [41].

Cholesterol is an essential part of every cell in the body. It is necessary for formation of new cells and for older cells to repair themselves after injury. It is also used by the adrenal glands in the synthesis of some hormone, such as cortisol, by the testicles to form testosterone, and by the ovaries to form estrogen and progesterone [42]. The high cholesterol level in plasma may be due to increased uptake of exogenous cholesterol and subsequent deposition and decreased cholesterol catabolism as evidenced by a reduction in bile acid production and turnover of bile acids. The metabolism of free and ester cholesterol are impaired in liver, spleen and thymus tissue and the rate of turnover was specifically

decreased in all tissues of hyperlipidemic rats [15]. Lead nitrate-mediated development of hypercholesterolemia involves the activation of cholesterol biosynthetic enzymes (i.e. 3-hydroxy- 3methylglutaryl-CoA reductase, farnesyl diphosphate synthase, and squalene synthase, CYP51) and the simultaneous suppression of cholesterol-catabolic enzymes such as 7a-hydroxylase [43]. Increase in LDL, VLDL levels are increase the risk of cardiovascular diseases [44 & 45].

Oxidative stress, specifically the oxidation of low density lipoprotein (LDL), has long been suspected of having a critical role in the development of atherosclerosis, in consequence of which antioxidants have been expected to have potential as antiatherogenic agents. Such agents would be able, in theory, to inhibit the oxidative modification of LDL that leads to the accumulation of cholesterol in atherosclerotic lesions [46 & 47].

Results of the present study have shown that Castelli's Risk Index I (TC/HDLc), Castelli's Risk Index II (LDLc/HDLc), Atherogenic Coefficient{(TC-HDLc) /HDLc} and Atherogenic Index of Plasma{(AIP)=log(TG/HDLc)} were elevated in lead acetate treated male albino mice group compared with the control group with statistically significant differences (p<0.05). These results run parallel to those reported by Ghosh *et al.*, [38] who reported that treatment of rats with lead acetate at a dose of 15 mg/kg body weight intraperitoneally (i.p) for a period of seven consecutive days caused alterations in the Cardiac Risk Ratio (Castelli's Risk Index I) TC/HDLc and Castelli's Risk Index II (LDLc/HDLc). Bhardwaj et al. [17] reported that lipid ratios like Atherogenic Index of Plasma, Castelli risk index and

Atherogenic coefficient could be used for identifying individuals at higher risk of cardiovascular disease in Indian population in the clinical setting especially when the absolute values of individual lipoproteins seem normal and in individuals with elevated triglycerides concentrations. Thus, the use of these indexes should be encouraged to complement the existing profile of tests for identifying high risk individuals for Coronary Artery Disease (CAD) and effective drug management.

Results of the present study which have shown that co-administration of propolis with lead acetate induced significant reduction in serum cholesterol, triglycerides, LDLc and VLDL concentrations and elevation in serum HDL-cholesterol. These results are in concordance with those of Maimuna *et al.*, [5] who, reported that the deleterious effects caused by lead intoxication were prevented in albino rats given lead concomitantly with aqueous extract of *Capsicum annum L.* fruits, suggesting that the extract offered protection against lead-induced organ damage in albino rats. Co-administration of propolis to chlorpyrifos treated rats restored serum total cholesterol, triglycerides and LDL-cholesterol parameters to normal levels [48]. Also, Abdel-Wahab [49] reported that pretreatment with melatonin in AlCl<sub>3</sub>-treated rats alleviated the elevation of total cholesterol and triglycerides in the plasma and restored their values toward the normal value of the control group. This anti-hyperlipidemic effect of melatonin may be primarily attributed to its antioxidant activity and the protection of cellular membrane integrity from Al-induced oxidative damage [50]. Another possible mechanism for the effect of melatonin on lipid profile may be its action on the gastrointestinal tract and the inhibition of cholesterol and triglycerides uptake, the augmentation of endogenous cholesterol clearance mechanisms through increasing the activity of cholesterol degrading enzymes and/or its effect on thyroid hormones which in turn affect lipid metabolism [51 & 52].

Treatment of male albino mice with lead acetate plus propolis decreased triglyceride level compared to the male albino mice treated with lead acetate only. Similar results were obtained by Cetin *et al.*, [53] who found that treatment of rats with propetamphos plus propolis decreased triglyceride levels compared to the rats treated with propetamphos. This suggests that propolis can modulate lipid metabolism. Fuliang *et al.*, [54] reported propolis to cause decrease in triglyceride level when administered to rats with diabetes mellitus. In addition, Kolankaya *et al.*, [55] reported that propolis caused a decrease in triglyceride level of rats treated with alcohol.

Oral ethanolic extracts of propolis (EEP) caused a significant decrease in plasma levels of total cholesterol, triacylglycerol, LDL-cholesterol and VLDL-cholesterol and significant increase in HDL-cholesterol in rabbits fed cholesterol diet. The data suggest that EEP may be protective against atherosclerosis and cardiovascular disease, particularly because they also decreased plasma LDL-cholesterol level [56]. Flavonoids supplementation significantly increased HDL-cholesterol and HDL-

cholesterol/total-cholesterol ratio [57]. The favorable lipid profile indicates a possible antiatherogenic property of the flavonoids [58]. Bok *et al.*, [18] suggest that flavonoids reduce cholesterol biosynthesis by means of inhibition of hepatic 3-hydroxy-3-methyl-glutaryl-CoA (HMG-CoA) reductase and acyl CoA: cholesterol o-acyltransferase (ACAT). Reduced ACAT activity may lead to lower availability of cholesterol ester for VLDL cholesterol packing, thereby resulting in a reduction of VLDL-cholesterol secretion from the liver, as suggested by Carr *et al.*, [59]. Diets containing flavonoids reduced the VLDL [60].

Increases of HDL have cardioprotective effect and it was proved by various studies. [44 & 45]. The increase in HDL-C observed in the present study, might be due to stimulation of pre- $\beta$  HDL-C and reverse cholesterol transport as demonstrated by previous studies [15 & 61]. High HDL-C levels could potentially contribute to its anti-atherogenic properties, including its capacity to inhibit LDL oxidation and protect endothelial cells from the cytotoxic effects of oxidized LDL [62]. The ethanol extract of propolis resulted in decreased serum levels of total cholesterol, triacylglycerol, LDL-cholesterol, VLDL-cholesterol of fasting rats; and to increased serum levels of HDL-cholesterol. This suggests that propolis can modulate the metabolism of blood lipid [54].

In the present study, co-administration of lead acetate with propolis were reduced Castelli's Risk Index I (TC/HDLc), Castelli's Risk Index II (LDLc/HDLc), Atherogenic Coefficient{(TC-HDLc)/HDLc} and Atherogenic Index of Plasma{(AIP)=log(TG/HDLc)} with statistically significant differences ( $p < 0.05$ ), when compared with lead acetate group.

In our study hypolipidemic and antiatherogenic effects of aqueous extract of propolis may be due to the antioxidant actions of the extract. Some antioxidant compounds identified in propolis include ferulic acid, quercetin and caffeic acid [63]. Some propolis is made bioactive by the presence of prenylated compounds [64]. Russo *et al.*, [65] studied a propolis and determined the antioxidant properties that are conferred by galangin, caffeic acid, ferulic acid, *p*-cumaric and CAPE. The antioxidant activities of propolis are related to its ability to scavenge singlet oxygen, superoxide anions, peroxy radicals, hydroxyl radicals and peroxy nitrite [66]. The primary mechanism of the effect of propolis may involve the scavenging of free radicals that cause lipid peroxidation. The other mechanism may comprise the inhibition of xanthine oxidase, which is known to cause free radicals to be generated [67].

## 5. Conclusion

From the previous discussion, it can be concluded that, the lead had adverse effects on lipid profile parameters and the ratios based on these parameters. Aqueous extract of Libyan propolis showed hypolipidemic and anti-atherogenic effects in lead acetate intoxicated male albino mice. So, the populations of high risk to lead should be advised to take

propolis. Further studies are necessary to elucidate exact mechanism of hypolipidemic and anti-atherogenic effects and potential usefulness of propolis as a hypolipidemic and antiatherogenic agent against heavy metals toxicity in clinical trials.

## References

- [1] Gurer H and Ercal N: Can antioxidants be beneficial in the treatment of lead poisoning? *Free Radic. Biol. Med.*, 2000;29: 927-945.
- [2] Sahayaraj, A P and Ayyadurai, K: Concentration of lead in the bioecosystem of buffaloes reared near Cooum river in Chennai. *Anim Sci Repor* 2007; 1 (2): 48 -55.
- [3] Hymavathi V and Rao LM: Effect of sublethal concentration of lead on the haematology and the biochemical constitution of *Channa punctata*. *Bulletin of pure and applied sciences* 2000;19:1-15.
- [4] Garaza A, Vega R and Soto E: Cellular mechanisms of lead neurotoxicity. *Med Sci Monitor* 2006;12(3): 57-65.
- [5] Maimuna Z , Ismail U, Mfonobong A and Balarabe S: Oxidative stress status and lipid level of rats co-administered with lead acetate and aqueous extract of sweet bell pepper (*Capsicum annum* L.) Fruits. *Inter J Sci Res* 2015,4(1):934 - 941.
- [6] Park SK, Schwartz J, Weisskopf M, Sparrow D, Vokonas PS, Wright RO, Coull B, Nie H and Hu H: Low level lead exposure, metabolic syndrome, and heart rate variability: The VA normative aging study. *Environ Health Perspect* 2006;114: 1718-1724.
- [7] Ademuyiwa O, Ugbaja RN, Rotimi SO, Abam E, Okediran BS, Dosumu OA and Onunkwor BO: Erythrocyte acetyl-cholinesterase activity as a surrogate indicator of lead-induced neurotoxicity in occupational lead exposure in Abeokuta, Nigeria. *Environ Toxicol Pharm* 2007;24:183-188.
- [8] Azab AE: Hepatoprotective effect of sesame oil against lead induced liver damage in albino mice: Histological and biochemical studies. *Amer J BioSci* 2014; 2(6-2): 1-11.
- [9] Azab AE, El-Dakhly AT, Alrawi QK and Albasha MO: Protective effects of sesame oil against lead acetate induced haemato-biochemical toxicity in albino mice. *Inter J Sci Res* 2015;4 (2): 2053 – 2063.
- [10] Vaziri ND, Liang K and Ding Y: Increased nitric oxide inactivation by reactive oxygen species in lead – induced hypertension. *Kidney Int* 1999; 56: 1492–1498.
- [11] Adikwu E, Deo O, Geoffrey, OBP and Enimeya DA: Lead organ and tissue toxicity: Roles of mitigating agents (Part 1). *British J Pharm Toxicol* 2013;4(6): 232-240.
- [12] Newairy AA and Abdou HM: Protective role of flax lignans against lead acetate induced oxidative damage and hyperlipidemia in rats. *Food Chem Toxicol* 2009;47(4): 813 – 818.
- [13] Anuradha CV, Ravikumar P: Restoration on tissue antioxidants by fenugreek seeds (*Trigonella Foenum Graecum*) in alloxan diabetic rats. *Indian J Physiol Pharmacol.* 2001; 45: 408-420.
- [14] Rao AV and Shen H: Effect of low dose lycopene intake on lycopene bioavailability and oxidative stress. *Nutr Res* 2002;22: 1125-1131.
- [15] Barakat LAA and Mahmoud RH: The antiatherogenic, renal protective and immunomodulatory effects of purslane, pumpkin and flax seeds on hypercholesterolemic rats. *North Amer J Med Sci* 2011; 3(9): 351 – 357.
- [16] Jaiswal J, Bhardwaj H, Rao CV and Sharma N: Hypolipidemic effect of *Calotropis gigantea* seeds extract on high fat diet induced atherogenic rats. *World J Pharm Pharmaceut Sci* 2014;3(6): 1139-1147.
- [17] Bhardwaj S, Bhattacharjee J, Bhatnagar MK and Tyagi S: Atherogenic index of plasma, castelli risk index and atherogenic coefficient- new parameters in assessing cardiovascular risk. *Int J Pharm Bio Sci* 2013; 3(3):359-364.
- [18] Bok SH, Lee SH, Park YB, Bae KH, Son KH, Jeong TS and Choi MS: Plasma and hepatic cholesterol and hepatic activities of 3-hydroxy-3-methyl-glutaryl-CoA reductase and acyl CoA: Cholesterol transferase are lower in rats fed citrus peel extract or a mixture of citrus bioflavonoids. *J Nutr* 1999; 129(6): 1182-1185.
- [19] Heeba GH, and Abd-Elghany MI: Effect of combined administration of ginger (*Zingiber Officinale* Roscoe) and atorvastatin on the liver of rats. *Phytomed* 2010;17(14): 1076-1081.
- [20] Ho C, Ferrara T, Chen Q, Rosen R and Huang M: Phytochemicals in teas and rosemary and their cancer preventive properties in: *Food phytochemicals for cancer prevention.* Amer Chem Soc Washington 1994: 2-19.
- [21] Marquele FD, Di Mambro VM, Georgetti SR, Casagrande R, Valim YML and Fonseca MJV: Assessment of the antioxidant activities of Brazilian extracts of propolis alone and in topical pharmaceutical formulation. *J Pharmacol Biomed Anal* 2005;39: 455-462.
- [22] Li YJ, Lin JL, Yang CW and Yu CC: Acute renal failure induced by a Brazilian variety of propolis. *Am J Kid Dis* 2005;46(6): 125-129.
- [23] Gunduz C, Biray C, Kosova B, Yilmaz B, Eroglu Z, Sahin F, Omay SB and Cogulu O: Evaluation of Manisa propolis effect on leukemia cell line by telomerase activity. *Leuk Res* 2005;29(11): 1343-1346.
- [24] Ozguner F, Bardak Y and Comlekci S: Protective effects of melatonin and caffeic acid phenethyl ester against retinal oxidative stress in long-term use of mobile phone. A comparative study. *Mol Cell Biochem* 2006;282(1-2): 83-88.
- [25] Frankel EN, Kanner J, German JB, Parks E and Kinsella JE: Inhibition of oxidation of human low-density lipoprotein by phenolic substances in red wine. *Lancet* 1998; 341: 454-457.
- [26] Abd El-Rahman SS: West-Libyan propolis and rosemary have synergistic anti-tumor effect against 12-O-tetradecanoylphorbol 13-acetate-induced skin tumor in BULB/C mice previously initiated with 7,12-dimethylbenz[a]anthracene. *Basic Appl Pathol* 2010; 3: 46 -51.
- [27] Valente MJ, Baltazar AF, Henrique R, Estevinho L and Carvalho M: Biological activities of Portuguese Propolis: Protection against free radical-induced

- erythrocyte damage and inhibition of human renal cancer cell growth in vitro. *Food Chem Toxicol* 2011;49:86-92.
- [28] Akao Y, Maruyama H, Matsumoto K, Ohguchi K, Nishizawa K, Sakamoto T, Araki Y, Mishima S and Nozawa Y: Cell growth inhibitory effect of cinnamic acid derivatives from propolis on human tumor cell lines. *Biol Pharm Bull* 2003; 26: 1057–1059.
- [29] Banskota AH, Tezuka Y, Midorikawa K, Matsushige K and Kadota S: Two novel cytotoxic benzofuran derivatives from Brazilian propolis. *J Nat Prod* 2000; 63: 1277– 1279.
- [30] Nieva Moreno M I, Isla M I, Sampietro AR and Vattuone MA: Comparison of the free radical scavenging activity of propolis from several regions of Argentina. *J. Ethnopharmacol* 2000;71:109-114.
- [31] Shalan MG, Mostafa MS, Hassouna MM, Hassab El-Nabi SE and El-Refaeia A : Amelioration of lead toxicity on rat liver with Vitamin C and silymarin supplements. *Toxicol* 2005; 206: 1-15.
- [32] El-Khayat Z, Ezzat AR, Arbid MS, Rasheed WI, and Elias TR: Potential effects of bee honey and propolis against the toxicity of ochratoxin A in rats. *Macedonian J Med Scie* 2009;2(4): 1-8.
- [33] Ashry MA, Abd Ellah HF and Gheth EMM: The Possible Ameliorative Effect of Propolis in Rat's Liver Treated with Monosodium Glutamate (MSG). *Nature Sci* 2012;10(12): 209 - 219 .
- [34] Allain CC, Poon LS, Chan CSG, Richmond W and Fu PC: Enzymatic determination of total serum cholesterol. *Clin Chem* 1974;20(4): 470-475.
- [35] Fossoti, P and Prencipe L: Serum triglycerides determination calorimetrically with an enzyme that produces hydrogen peroxide. *Clin Chem* 1982;28(10):2077-2080.
- [36] Burstein M, Scholnik H and Morfin R: Rapid method for the isolation of lipoproteins from human serum by precipitation with polyphenols. *J lipid Res* 1970; 11: 583-595.
- [37] Friedwald W, Levy R and Fredrichson D: Estimation of the concentration of low density lipoprotein cholesterol in plasma without use of the preparative ultracentrifuge. *Clin Chem* 1972; 226: 499-502.
- [38] Ghosh D, Dey M, Ghosh AK, Chattopadhyay A and Bandyopadhyay D: Melatonin protects against lead acetate induced changes in blood corpuscles and lipid profile of male Wistar rats. *J Pharm Res* 2014;8(3): 336-342.
- [39] Kasperczyk S, Birkner E, Kasperczyk A and Kasperczyk J: Lipids, lipid peroxidation and 7-ketocholesterol in workers exposed to lead. *Hum Exp Toxicol* 2005; 24: 287- 295.
- [40] Bashandy SA: Beneficial effect of combined administration of vitamin C and vitamin E in amelioration of chronic lead hepatotoxicity. *Egypt J Hosp Med.* 2006; 23: 371- 384.
- [41] Miller NE: Associations of high-density lipoprotein subclasses and apolipoproteins with ischemic heart disease and coronary atherosclerosis. *Am. Heart J.* 1987;113: 589-597.
- [42] Jefcoate C R, McNamara BC, Artemenko I and Yamazaki T: Regulation of cholesterol movement to mitochondrial cytochrome P450sc in steroid hormone synthesis. *J Steroid Biochem Mol Biol.* 1992;43(8):751-767.
- [43] Kojima M, Nemoto K, Murai U, Yoshimura N, Ayabe Y and Degawa M: Altered gene expression of hepatic lanosterol 14 $\alpha$ -demethylase (CYP51) in lead nitrate-treated rats. *Arch Toxicol.* 2002; 76: 398 – 403.
- [44] Roy S: Antioxidant and protective effect of latex of *Calotropis procera* against alloxan induced diabetes in rats. *J Ethnopharmacol.* 2005; 102(3): 470-473.
- [45] Jaiswal J, Bhardwaj H, Srivastava S, Gautam H, Sharma S and Rao C: Anti-diabetic activity of methanolic extract of *Calotropis gigantea* seeds on STZ induced diabetic rats. *Int J Pharm Pharm Sci.* 2013; 6(1): 254-257.
- [46] Brown MS and Goldstein JL: Lipoprotein metabolism in the macrophage: implications for cholesterol deposition in atherosclerosis. *Annu. Rev. Biochem.* 1983;52:223–261.
- [47] Suzuki H, Kurihara Y, Takeya M, Kamada N, Kataoka M, Jishage K, Ueda O, Sakaguchi H, Higashi T, Suzuki T, et al: A role for macrophage scavenger receptors in atherosclerosis and susceptibility to infection. *Nature* 1997;386(6622): 292–296.
- [48] El Mazoudy RH, Attia AA and El-Shenawy NS: Protective role of propolis against reproductive toxicity of chlorpyrifos in male rats. *Pest Biochem Physiol* 2011;101:175–181.
- [49] Abdel-Wahab WM: AlCl<sub>3</sub>-induced toxicity and oxidative stress in liver of male rats: Protection by melatonin. *Life Sci J* 2012; 9(4): 1173-1182.
- [50] Carla M, Alice S, Ramos A, Azevedo MF, Lima CF, Fernandes-Ferreira M and Wilson C P: Sage tea drinking improves lipid profile and antioxidant defenses in humans. *Int J Mol Sci* 2009; 10: 3937-3950.
- [51] Chan TY and Tang PL: Effect of melatonin on the maintenance of cholesterol homeostasis in the rat. *Endocr Res.* 1995; 21(3):681-96.
- [52] Wakatsuki A, Okatani IY, Kaneda C and Fukaya T: Effect of short-term melatonin administration on lipoprotein metabolism in normolipidemic postmenopausal women. *Maturitas* 2001; 38:171- 177.
- [53] Cetin E, Kanbur M, Silici S and Eraslan G: Propetamphos induced changes in haematological and biochemical parameters of female rats: Protective role of propolis. *Food Chem Toxicol* 2010;48 : 1806 -1810.
- [54] Fuliang HU, Hepburn HR, Xuan H, Che M, Daya S and Radloff SE: Effects of propolis on blood glucose, blood lipid and free radicals in rats with diabetes mellitus. *Pharmacol Res* 2005;51: 147-152.
- [55] Kolankaya D, Selmanoglu G, Sorkun and K, Salih B: Protective effects of Turkish propolis on alcohol induced serum lipid changes and liver injury in male rats. *Food Chem.* 2002;78:213-217.
- [56] Fernandes AAH, Novelli ELB and Junior A F: Beneficial effects of propolis on experimental hypercholesterolaemia in rabbits. *J Brazilian Soc Food Nutr* 2006;31(1): 65-78.
- [57] Yousef MI, Kamel KI, Esmail AM, and Baghdadi HH: Antioxidant activities and lipid lowering effects of isoflavone in male rabbits. *Food Chem Toxicol* 2005; 42: 1497-1503.
- [58] Adaramoye OA, Nwaneri VO, Anyanwu KC, Farombi EO and Emerole GO: Possible antiatherogenic effect of



- kolaviron (*Garcinia kola* seed extract) in hypercholesterolaemic rats. Clin Exper Pharmacol Physiol 2005; 32(1/2): 40-46.
- [59] Carr TP, Parks JS and Rudel LL: Hepatic ACAT activity in African green monkeys in highly correlated to plasma LDL cholestewrol enrichment and coronary artery atherosclerosis. Arteriocler Thromb1992;12(11): 1274-1283.
- [60] Kurowska EM and Manthey JA: Hypolipidemic effects and absorption of citrus polymethoxylated flavones in hamsters with diet-induced hypercholesterolemia. J Agr Food Chem 2004; 52(10): 2879-2886.
- [61] Daniel M: Medicinal Plants: Chemistry and Properties. Science Publishers, Enfield, NH, 2006 p. 184.
- [62] Assmann G and Nofer J: Anthropometric protective effects of high density lipoproteins. Ann Rev Med 2003; 54: 321-341.
- [63] Usami E, Kusano G, Takayose T, Wachi H and Seyama Y: Assessment of antioxidant activity of natural compounds by water and lipid –soluble antioxidant factor. Yakugaku Zasshi. 2004;124:847-850.
- [64] Chen CN, Weng MS, Wu CL and Lin J K: Comparison of radical scavenging activity, cytotoxic effects and apoptosis induction in Human melanoma cells by Taiwanese propolis from different sources. Evidenced-based Compl Altern Med (eCAM) 2004;1(2):175-185.
- [65] Russo A, Troncoso N, Sanchez F, Garbarino J and Vanella A: Propolis protect human spermatozoa from DNA damage caused by benzo[a]pyrene and exogenous reactive oxygen species. Life sci 2006;78:1401-1406.
- [66] Ferrali M, Signorini C and Caciotti B : Protection against oxidative damage of erythrocytes membrane by the flavinoid quercetin and its relation to iron chelating activity. FEBS Lett 1997; (416):123- 129.
- [67] Kanbura M, Eraslan G and Silici S: Antioxidant effect of propolis against exposure to propetamphos in rats. Ecotoxicol Environ Safety 2009; (72): 909-915.