









**Table 2:** Solubilization zone produced on modified Aleksandrov medium amended with KCl, K<sub>2</sub>SO<sub>4</sub>, mica powder are the potassium sources

Bacterial isolate	KCl Solubilization zone diameter (mm)	K <sub>2</sub> SO <sub>4</sub> Solubilization zone diameter (mm)	Mica powder solubilization zone (mm)
JS-17	13.2	9.2	4.0
JS-7	12.6	8.1	3.5
JS-3	12.2	7.2	3.1
JS-16	11.4	6.4	2.6
JS-31	10.5	5.2	2.2
JS-53	10.0	4.1	1.8

### 3.5 Zinc Solubilization Activity

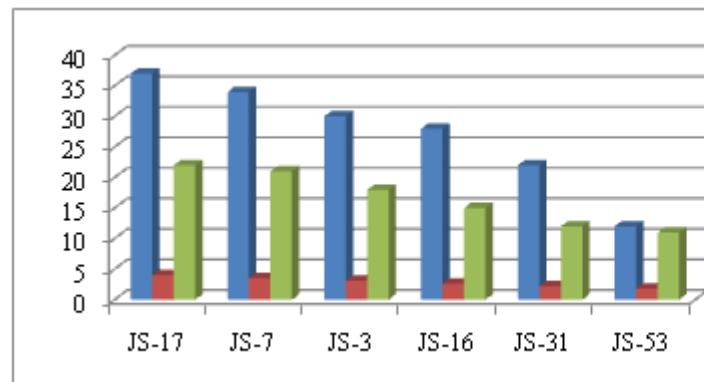
Zinc is one of the eight essential micronutrients required for healthy growth and reproduction of crop plants. From the data, it is obvious that the strains varied in their ability to solubilize different forms of insoluble zinc. Out of 55 strains, we could identify 14 strong Zn solubilizers. In the past also, efforts were made to identify zinc solubilizers with varying abilities. The demonstrated variation in the ability of solubilizing given zinc sources could be due to metabolic activity of a given strain [47]. There are different mechanisms of solubilization which have been identified with proton excretion, production of organic acids and other chelating metabolites [48]. Organic acid production by microbial strains plays a major role in solubilization [49]. The zinc solubilization in our studies could be due to production of organic acids, like gluconic acids that is increased by the fall in pH of culture media noted in all cases. The transformation of glucose to gluconic acid by the glucose oxidative external pathway in *Pseudomonas* spp. and other bacteria has been interpreted as a competitive approach by microorganisms [50]. Zinc phosphate solubilization by a strain of *P. fluorescens* was also reported [51]. The supplementing medium with zinc sulphate resulted in more gluconic acid production. Plant growth parameters of legume crop increase when seedlings were inoculated with zinc solubilizing bacteria compared to uninoculated controls [52]. The potential of rhizobacteria to promote nutrient uptake and plant growth could be due to synergistic action of the growth promoting traits than individual effect [53]. This solubilization property is important in nutrient cycling. Zinc phosphate solubilization by a strain of *Pseudomonas fluorescens* was also investigated [54].

All the selected strains of *Pseudomonas* used could effectively solubilize the insoluble Zn compounds used namely ZnCO<sub>3</sub> and ZnO. It is apparent from the zinc solubilization data that the solubilization potential varied with each isolate. The zone of solubilization was comparatively high in ZnO amended medium as compared to ZnCO<sub>3</sub>. Size of the solubilization zone ranged from 7 to 22mm in ZnCO<sub>3</sub> and from 9 to 33mm in ZnO incorporated medium. Among the cultures JS-16, and JS-17 showed the highest solubilization zone in ZnCO<sub>3</sub>, where as JS-17 and JS-7 showed 31mm zone in ZnO amended medium (Table 3)

**Table 3:** Solubilization zones produced on modified PVK medium amended with ZnO, ZnCO<sub>3</sub>

Bacterial isolate	ZnO Solubilization zone diameter (mm)	ZnCO <sub>3</sub> Solubilization zone diameter (mm)
JS-17	31	22
JS-7	31	21
JS-3	26	18
JS-16	22	15
JS-31	13	12
JS-53	12	11

Most of the soils are rich in total Zn, K and P. However, their availability to the plants when needed is very limited. Pseudomonads able to solubilize P, K and Zn and thus, offer best possible nutrient recycling mechanism at a low input cost where expensive inorganic fertilizers are becoming too expensive to small and marginal farmers. In the present study, bacteria when characterized by P, K and Zn solubilization was observed that some strains have high potential to solubilize the said essential nutrients. *Pseudomonas* isolate JS-17 showed TCP solubilization, and were also able to solubilize insoluble Zn sources. Interestingly, isolate JS-16 that could solubilize maximum Zn could not show any Pi solubilization. In this study we could find 6 isolates that could solubilize P, K and Zn. There are some PGPR that can fix nitrogen, solubilize mineral nutrients and mineralize organic compounds [55]. As we could screening for a potential PGPR strains on the basis of direct plant growth promoting traits viz., IAA, solubilization of Phosphate, Zinc solubilization, Potassium solubilization.

**Graph 5:** Comparative solubilization zones of P, K & Zn minerals

### 4. Conclusion

The experiment performed under *invitro* condition showed that these isolates may promote plant growth promoting potentials. PGPR include facilitating the uptake of certain nutrient like nitrogen, phosphorus, potassium and zinc forms for nutrient availability from the soil and root environment by producing plant hormone indole-3-acetic acid and multinutrient solubilization. Such multiple positive PGP Traits isolates can be further explored as potential biofertilizer for the sustainable agriculture. The direct promotions of plant growth by such situation warrants for compatibility between such strains for formulating consortia and use in the field. However, if strains that possess ability to mobilize multi-nutrients could be a benefit as they could be deployed directly with minimum efforts without going through the formulation.

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## References

- [1] Gyaneshwar, P.; Kumar, N.J.; Pareka, L.J. & Podle, P.S. (2002). Role of Soil Microorganisms in Improving P Nutrition of Plants. *Plant and Soil*, Vol. 245, No. 1, (August 2002), pp. 83-93.
- [2] Brady, N.C., 1990. The nature and properties of soils. *Macmillan*, New York, USA, pp.351-380.
- [3] Goldstein A.H., Bacterial solubilization of mineral phosphates: historical perspective and future prospects. *American Journal of Alternative Agriculture*1, 1986, pp.51-57.
- [4] Kumar and Kumar, 2000, Plant growth promoting activity of Pseudomonads in Rice crop, *Int.J.Curr.Microbiol.App.Sci* Volume 2 Number 11 (2013) pp. 152-157.
- [5] Richardson, 2001. Measures of skill and value of ensemble prediction systems, their inter relationship and the effect of ensemble size. *Q. J. R. Meteoml. SOC.* (2001). 127, pp. 2473-2489.
- [6] Aleksandrov, V.G., Blagodyr, R.N. and Iiev, I.P., 1967, Liberation of phosphoric acid from apatite by silicate bacteria. *Mikrobiologi Zh (Kiev)*, 29, 111-114.
- [7] Ullman, W.J., Kirchner, D.L. and Welch, S.A., 1996, Laboratory evidence by microbially mediated silicate mineral dissolution in nature. *Chemistry and Geology*, 132, 11-17.
- [8] Bennett, P.C., Choi, W.J. and Rogera, J.R., 1998, Microbial destruction of feldspars. *Mineral Management*, 8(62A), 149-150.
- [9] Groudev, S.N., 1987, Use of heterotrophic microorganisms in mineral biotechnology. *Acta Biotechnology*, 7, 299-306.
- [10] Rogers, J.R., Bennett, P.C. and Choi, W.J., 1998, Feldspars as a source of nutrients for microorganisms. *American Mineralogy*, 83, 1532-1540.
- [11] P. N. Sharma, C. Chatterjee, C. P. Sharma, and S. C. Agarwala, "Zinc deficiency and anther development in maize," *Plant and Cell Physiology*, vol. 28, no. 1, pp. 11-18, 1987.
- [12] M. N. Hughes and R. K. Poole, "Metal speciation and microbial growth—the hard (and soft) facts," *Journal of General Microbiology*, vol. 137, no. 4, pp. 725-734, 1991.
- [13] F. L. Crane, I. L. Sun, and M. G. Clark, "Transplasma-membrane redox systems in growth and development," *Biochimica et Biophysica Acta*, vol. 811, no. 3, pp. 233-264, 1985.
- [14] M. N. Hughes and R. K. Poole, "Metal speciation and microbial growth—the hard (and soft) facts," *Journal of General Microbiology*, vol. 137, no. 4, pp. 725-734, 1991.
- [15] T.Wakatsuki, "Metal oxidoreduction by microbial cells," *Journal of Industrial Microbiology*, vol. 14, no. 2, pp. 169-177, 1995.
- [16] Pramer, D. and Schmidt, E.L. (1956). Experimental Soil Microbiology. *Burter Publ. Co. Minneapolis*. 107 pp.
- [17] Rangaswami, G.1993. Diseases of crop plants in India. *Preitice hall of India* pvt. Ltd, New Delhi, p.498.
- [18] Kreig NR and Holf JG. 1984. *Bergeys Manual of Systematic Bacteriology*. William and Wilkins, Baltimore, USA.
- [19] Pikovskaya, R.I. (1948) Mobilization of phosphorus in soil in connection with the vital activity of some microbial species. *Mikrobiologiya* 17, 362-370.
- [20] Olsen, S.R. and L.E. Sommers. 1982. Phosphorus. p 416-418. In: A.L. Page *et al.* (eds.) *Methods of soil analysis, part 2. Agron. Mongr. 9. 2nd ed. ASA and SSSA*, Madison, WI.
- [21] Aleksandrov, V. G., Blagodyr, R. N. and Iiev, I. P. (1967). Liberation of phosphoric acid from apatite by silicate bacteria. *Mikrobiyol Zh.* (Kiev), 29: 111-114.
- [22] Amit Sagervanshi, Priyanka Kumari, Anju Nagee and Ashwani Kumar.2012. Isolation and Characterization Of Phosphate Solublizing Bacteria from Anand Agriculture Soil. *International Journal of Life Sciences and Pharma Research*. 23:256-266.
- [23] M.A. Whitelaw, Growth Promotion of Plants inoculated with phosphate solubilizing fungi, *Adv. Agron.* 2000, 69: 99-151.
- [24] R.M.N. Kucey, H.H. Janzen and M.E. Legett, 1989, Microbially mediated increases in plant available phosphorus, *Adv. Agron.* 42: 199-228.
- [25] C.S. Nautiyal, S. Bhadauria, P. Kumar, H. Lal, R. Mondal and D. Verma, Stress induced phosphate solubilization in bacteria isolated from alkaline soils. *FEMS Microbiol.Lett.* 2000, 182: 291-296.
- [26] Nirmala Jyothi Lukkani, EC. Surendranatha Reddy, Sulphur oxidation by fluorescent pseudomonads isolated from ground nut rhizospheric soil and their variability in sulphate production , *International Journal of Current Research* Vol. 7, Issue, 01, pp.11373-11377, January, 2015.
- [27] J.C. Biswas, J.K. Ladha and F.B. Dazzo, Rhizobia Inoculation Improves Nutrient Uptake and Growth of Lowland Rice. *Soil Sci. Soc. Amer. J.* 2000, 64: 1644-1650.
- [28] Chen, Y.P., Rekha, P.D., Arun, A.B., Shen, F.T., Lai, W.A. and Young, C.C. 2006b. Phosphate solubilizing bacteria from subtropical soil and their tricalcium phosphate solubilizing abilities. *Applied Soil Ecology*, 34: 33-41.
- [29] Deshwal, V.K., Reena, Sharma, P., Gupta, S., Chakraborty, M. and Chatterji, T. 2011d. Phosphorous Solubilizing *Pseudomonas Aeruginosa* PMV-14 Enhance Productivity in Rice Crop. *Int. J. Appl. Agri. Res.* 6(1): 29-33.
- [30] Johri J K, Surange S & Nautiyal C S, (1999) Occurrence of salt, pH and temperature tolerant phosphate solubilizing bacteria in alkaline soils, *Curr .Microbiol.* 39 89-93.
- [31] Gyaneshwar, P., L. J. Parekh, G. Archana, P. S. Podle, M. D. Collins, R. A. Hutson and K. G. Naresh. 1999. Involvement of a phosphate starvation inducible glucose dehydrogenase in soil phosphate solubilization by

- Enterobacter asburiae*. *FEMS Microbiol. Lett.* 171:223-229.;
- [32] Venkateswarlu, B.; Rao, A.V. & Raina, P.(1984). Evaluation of Phosphorous Solubilization by Microorganisms Isolated from Arid Soils. *Journal of the Indian Society of Soil Science*, Vol. 32, No. 3, (September 1984), pp. 273-277.
- [33] Gyaneshwar, P.; Kumar, N.J.; Pareka, L.J. & Podle, P.S. (2002). Role of Soil Microorganisms in Improving P Nutrition of Plants. *Plant and Soil*, Vol. 245, No. 1, (August 2002), pp. 83-93.
- [34] Puente ME, Bashan Y, Li CY, Lebsky VK (2004) Microbial populations and activities in the rhizoplane of rock-weathering desert plants. I. Root colonization and weathering of igneous rocks. *Plant Biol* 6:629–642.
- [35] Rodriguez, H., Fraga, R. 1999. Phosphate solubilizing bacteria and their role in plant growth promotion. *Biotechnol. Adv.* 17, 319-339.
- [36] Hameeda, B., Rupela, O.P., Reddy, G., Satyavani, K., 2006. Application of plant growth-promoting bacteria associated with composts and macrofauna for growth promotion of Pearl millet (*Pennisetum glaucum* L.). *Biol. Fertil. Soils*. in press.
- [37] Nirmala Jyothi Lukkani and EC. Surendranatha Reddy, 2014, Evaluation of plant growth promoting attributes and Biocontrol potential of native fluorescent *Pseudomonas* spp. against *Aspergillus niger* causing collar rot of ground nut. Volume-4, issue-4, Oct-Dec-2014, *International Journal of Plant, Animal and Environmental Sciences*.
- [38] Han H.S., Supanjani, Lee K.D. (2006) Effect of co-inoculation with phosphate and potassium solubilizing bacteria on mineral uptake and growth of pepper and cucumber, *Plant Soil Environ.* 52, 130–136.
- [39] Mikhailouskaya, N. and Tcherhysh, A., 2005, K-mobilizing bacteria and their effect on wheat yield. *Latvian Journal of Agronomy*, 8, 154-157.
- [40] Hu, X.F., Chen, J. and Guo, J.F., 2006, Two phosphate and potassium solubilizing bacteria isolated from Tiannu mountain, Zhejiang, China. *World Journal of Microbiology and Biotechnology*, 22, 983-990.
- [41] Li, Y.F., 1994, The characteristics and function of silicate dissolving bacteria fertilizer. *Soil Fertilizers*, 2, 48-49.
- [42] Zeng, X., Liu, X., Tang, J., Hu, S., Jiang, P., Li, W. and Xu, L., 2012, Characterization and potassium-solubilizing ability of *Bacillus circulans* Z1-3. *Advanced Science Letters*, 10, 173-176.
- [43] H.L.S. Tandon, and G.S. Sekhon, Potassium research and agricultural production in India, *Fertilizer development and consultation organization*, New Delhi. pp. 144, 1988.
- [44] Liu, G.Y., 2001, Screening of silicate bacteria with potassium releasing and antagonistic activity. *Chinese Journal of Applied Environmental Biology*, 7, 66-68.
- [45] Yakhontova, L.K., Andreev, P.I., Ivanova, M.Y. and Nesterovich, L.G., 1987, Bacterial decomposition of smectite minerals. *Doklady Akademii Nauk, USSR*, 296, 203-206.
- [46] Liu, W., Xu, X., Wu, S., Yang, Q., Luo, Y. and Christie, P., 2006, Decomposition of silicate minerals by *Bacillus mucilaginosus* in liquid culture. *Environmental Geochemistry and Health*, 28, 133-140.
- [47] Sadaf and Nuzhat (2008). Effect of various parameters on the efficiency of zinc phosphate solubilization by indigenous bacterial isolates, *African Journal of Biotechnology*. Vol. 7 (10), pp. 1543-1549, 16 May, 2008.
- [48] Agnihorti VP (1970). Solubilization of insoluble phosphates by some soil fungi isolated from nursery seedbeds. *Can. J. Microbiol.* 16: 877-880.
- [49] Nguyen C, Yan W, Le Tacon F, Lapyire F (1992). Genetic variability of phosphate solubilizing activity by monocaryotic and dicaryotic mycellia of the ectomycorrhizal fungus *laccaria bicolor* (Maire) PD Orton. *Plant Soil* 143: 193-199.
- [50] PAUL H. WHITING,\* MELVIN MIDGLEY and EDWIN A. DAWES (1975). The Regulation of Transport of Glucose, Gluconate and 2-Oxogluconate and of Glucose Catabolism *Pseudomonas aeruginosa*. *Biochem. J.* (1976) 1M4, 6S9-669.
- [51] Di Simine, CD; Sayer, JA; Gadd, GM (1998). Solubilization of zinc phosphate by a strain of *Pseudomonas fluorescens* isolated from a forest. *Soil. Biol Fertil Soils* 28:87–94.
- [52] Iqbal, U. and Jamil, N. and Ali, I. and Hasnain, S. 2010. Effect of zinc-phosphate-solubilizing bacterial isolates on growth of *Vigna radiata*. *Ann. Microbiol.* 60: 243–248.)
- [53] Glick B.R., Patten C.L., Holguin G., Penrose D.M., 1999. Biochemical and Genetic Mechanisms Used by Plant Growth Promoting Bacteria. *Imperial College Press*, London, UK.)
- [54] C. D. Di Simine, J. A. Sayer, and G. M. Gadd, “Solubilization of zinc phosphate by a strain of *Pseudomonas fluorescens* isolated from a forest soil, *Biology and Fertility of Soils*, vol. 28, no. 1, pp. 87–94, 1998.
- [55] Martínez-Viveros, O., Jorquera, M.A., Crowley, D.E., Gajardo, G., Mora, M.L. 2010. Mechanisms and practical considerations involved in plant growth promotion by rhizobacteria. *Journal of Soil Science and Plant Nutrition* 10, 293–319.

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