Hemocyte and Biochemical Changes of Antheraea mylitta D. Infected with Antheraea mylitta Cytoplasmic Polyhedrosis Virus (AmCPV)

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Abstract: The hemocyte and biochemical changes in Antheraea mylitta after per oral inoculation of Cytoplasmic Polyhedrosis Virus (AmCPV) was observed. The total haemocyte count (THC) increased gradually from day 1 (13650/mm³) to 8 (16625/mm³) in uninoculated silkworms kept as control. In AmCPV treated silkworm larvae the THC increased up to day 2 (12700 - 14480/mm³) of inoculation. Thereafter, a decrease was noticed from day 3 - 8 (14350 - $3812/mm^3$). The differential haemocyte count (DHC) was different in inoculated silkworms then the control lot of silkworms. In the haemolymph of control larvae the prohaemocyte, plasmatocytes and granulocytes were more in number whereas oenocytoids were less in number. The number of degenerated cells increased among treated lot up to 8^{th} days of post inoculation. The total protein content in the haemolymph of treated larvae during the first and second day was similar to that of control. On third day, the protein content increased among treated larvae. But from fourth day onwards, the protein content decreased (20.25 mg/ml) against control (33.73 mg/ml). The total carbohydrate content gradually increased during fifth instar in control lot larvae, whereas, it steadily increased up to 4^{th} day and then showed a down ward trend from 5^{th} day treated larvae. The total lipid content in the haemolymph decreased among treated larvae day 2, whereas in control larvae, an increasing trend was noticed.

Keywords: Antheraea mylitta, AmCPV, Hemocyte and Biochemical

1. Introduction

Immune system includes certain types of blood cells. It also includes chemicals, carbohydrates, lipids and proteins. In insects, several types of haemocytes are observed in the haemolymph (Butt & Shields, 1996). Saran *et al.* (2002) classified the blood cells in the silkworm, *B. mori* L. and *A. mylitta* in to six types *viz.* prohaemocytes, plasmatocytes, granulocytes, spherulocytes, imaginal spherulocytes and oenocytes. Proteins are the derivatives of high molecular weight polypeptides. They play a vital role in the formation of structures in organisms. Like carbohydrates and fats proteins also can be utilized for energy purpose. In silkworm, blood glucose level can be correlated to their level of metabolism, and is comparable with mammalian blood glucose. Lipids play an important role in the biochemical processes delaying growth and development of insects.

To date, reports on hemocyte and biochemical response in *AmCPV* infected silkworm, *A. mylitta* are scanty. Hence, present investigation was conducted to study hemocyte and biochemical changes of *Antheraea mylitta* D. infected with *Antheraea mylitta* Cytoplasmic Polyhedrosis Virus (*AmCPV*).

2. Materials and Methods

Cytoplasmic Polyhedrosis Virus (AmCPV) Inoculum

Cytoplasmic polyhedrosis virus was purified from diseased silkworm as suggested by Aizawa, (1971).

Inoculation of AmCPV

AmCPV (1 x 10⁵ PIB/ml.) was smeared on to the *Terminalia* tomentosa (assan) leaves and fed to 4th instar of Daba B.V eco race after 24 hr of moult. The treated and controlled batches were reared in indoor rearing condition.

Samples collection

Every day from 0 to 8th day 6 larvae/day were collected from each replication, the haemolymph from all the 6 larvae was collected in to three eppendoff tubes (2 larvae haemolymph/tube) on ice and stored at 4°C. A total of 6 tubes represented 3 replication collections.

Estimation of haemocytes count

Every day total haemocyte count (THC) estimation in the haemolymph of treated and control batches was determined following the method described by Tauber and Yeager (1935) using haemocytometer. The THC per mm³ of haemolymph was estimated according to the formula suggested by Jonesh (1962). Different haemocytes were identified based on the morphological features as described by Nittono (1960). The observations were made on THC and DHC counts.

Estimation of biochemicals

The total protein, total carbohydrate and total lipid content in hemolymph was estimated by the method of Lowry *et al.* (1951), phenol-sulphuric acid method - Dubois *et al.*, 1956 and Folch *et al.*, 1951, respectively.

Annalysis of data

Data recorded for THC and DHC counts, total protein, total carbohydrate and total lipid content were statistically analyzed using Completely Randomized Design (Snedecor and Cockron, 1971).

3. Results and Discussion

Total Haemocyte Counts (THC)

The total hemocyte count in the control silkworm increased from 1^{st} day to 6^{th} day and decreased on 7^{th} and 8^{th} day. In the control, total hemocyte counts was $13650/\text{mm}^3$ and

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increased to $17702/\text{mm}^3$ by 6th day (Table 1). On 7th day total hemocyte count was $16878/\text{mm}^3$ and 8th day the count was decreased to $16625/\text{mm}^3$. While in *AmCPV* treated silkworm the count was increased after inoculation up to 2nd day of infection and then there was a decrease for a period ranging from 3-8 days.

(Balavenkatasubaiah *et al*, 2001) and decrease (Gillium and Shimanuki, 1967) to counter foreign body when infected. The cellular responses to infection have been worked out in many insect by earlier workers (Horohove and Dunn, 1983). On the basis of the above findings of the earlier workers it is evident that CPV induce the defense response through multiplication of haemocytes as is indicated by the increase in total haemocyte counts of the hemolymph of the worms.

The present observations are in agreement with the earlier investigation that the number of haemocytes may increase

Table 1. Total haemocyte count in AmCPV treated and healthy silkworm, A. mylitta
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Treatment				Days post	inoculation			
Heatment	1	2	3	4	5	6	7	8
AmCPV treated	12700	14480	14350	12671	9851	6203	5009	3812
control	13650	12530	12982	14455	16670	17702	16878	16625
S.E. \pm	131.26	141.51	126.07	105.82	155.98	1823.12	191.76	191.23
C. D. at 5%	137.18	152.26	177.43	124.28	256.75	308.45	289.45	331.87

Differential Haemocyte Counts (DHC)

In control hemocytes showed a gradual increasing trend during developmental period. The number of prohemocyte, plasmatocytes, granulocytes and spherulocytes *viz.* 22-35, 32-38, 35-38 and 17-28, respectively in control (Table 2). While during progressive infection, the gradual decrease in number of prohemocyte, spherulocytes and oenocytoids *viz.* 28-12, 24-9 and 13-3 respectively was noticed in *AmCPV* treated batch. The plasmatocytes and granulocytes increased up to 3^{rd} day and decresing trend was observed from 5^{th} day onwards. The number of oenocytoid was less in both the

treatments and ranged from 3 to 15. Few vermiform cells, synonyms of plasmatocyte were recorded in both treatment and control. The number of degenerated cells was comparatively less in control than the treated batch.

The number of prohemocyte decreased due to the conversion of prohemocyte to other types of haemocyte during course of infection and number of plasmatocytes and granulocytes increased as both are involved in defense mechanism against entry of pathogens.

Table 2. Differential baemocyte count in	AmCPV treated and healthy silkworm, A. m	vlitta
Table 2. Differential fractiocyte count fi	Amer v treated and heating sikworm, A. m	упца

Treatment	Haamaarta		Days post inoculation								
	Haemocyte	1	2	3	4	5	6	7	8		
	PR	28	25	24	23	18	17	15	12		
	PL	33	39	44	42	38	35	32	25		
	GR	35	45	48	45	44	41	32	27		
AmCPV treated	SP	24	20	18	17	15	14	12	9		
	OE	13	11	10	8	8	5	4	3		
	VER	1	-	-	-	-	-	-	-		
	DEG	22	24	25	26	30	37	39	42		
	PR	22	25	27	29	30	33	34	35		
	PL	32	33	35	37	39	41	37	38		
	GR	35	36	38	39	42	40	38	38		
Control	SP	17	18	20	22	25	26	27	28		
	OE	5	6	8	10	10	12	13	15		
	VER	-	-	-	-	4	1	-	-		
	DEG	10	12	13	14	16	9	7	8		
PR = Prohae	emocyte	GR = Gr	anulocyte	cyte OE = Oenocytoid			= Degenerat	ed cell			
PL = Plasma	atocyte	SP = Sph	erulocyte	VER = Ve	rmiform cell						

Biochemical changes in *AmCPV* treated and control batches

Total protein content

The hemolymth protein in control silkworm increased gradually from 16.31 mg/ml on 1^{st} day to 33.73 mg/ml on day 8^{th} (Table 3). In treated silkworm, the total hemolymph protein have shown increasing trend from 1^{st} (16.26 mg/ml) to 3^{rd} day (24.22 mg/ml) and decreasing trend from 4^{th} day onwards and reached 20.25 mg/ml by 8^{th} day from the inoculation.

The results indicated that changes occured in the hemolymph protein, during the course of AmCPV infection. The difference in hemolymph protein healthy silkworm and treated silkworm becames more pronounced as the diseases progresses. This would probably indicate that during infection the synthesis and release of proteins from fat bodies are greatly increased. There are reports of production of antimicrobial substances such as lectin, defensin and attacin with the entry of foreign bodies (Wago, 1995).

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Treatment			To	otal protein	content mg/1	nl		
				Days post i	noculation			
	1	2	3	4	5	6	7	8
AmCPV treated	16.26	20.14	24.22	23.58	22.16	21.59	20.98	20.25
Control	16.31	20.19	23.54	26.74	28.53	30.34	32.83	33.73
S.E. \pm	0.21	0.28	0.19	1.37	0.48	0.52	0.41	0.56
C. D. at 5%	1.92	1.27	1.21	1.52	1.68	1.93	2.64	2.15

Table 3: Total protein content in AmCPV treated and healthy silkworm, A. mylitta

Total carbohydrate and total lipid contents

The hemolymph carbohydrate in control silkworm increased gradually from 6.75 mg/ml on 1^{st} day to 11.12 mg/ml on day 8^{th} (Table 4). In inoculated silkworm, the total hemolymph

carbohydrate have shown increasing trend from 1^{st} (6.73 mg/ml) to 6^{th} day (8.05 mg/ml) and decreasing trend from 7^{th} day onwards and reached 7.56 mg/ml by 8^{th} day from the treatement.

Table 4: Total carbohydrate content in AmCPVtreated and healthy silk	worm, A. <i>mylitta</i>
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Treatment			Total	carbohydra	ate content n	ng/ml				
	Days post inoculation									
	1	2	3	4	5	6	7	8		
AmCPV treated	6.73	6.98	7.83	8.31	8.2	8.05	7.93	7.56		
Control	6.75	7.02	7.89	8.42	9.86	10.51	10.69	11.12		
S.E. ±	0.19	0.27	0.19	1.36	0.46	0.51	0.42	0.57		
C. D. at 5%	1.91	1.26	1.21	1.51	1.64	1.93	2.61	2.14		

The hemolymph lipid in control silkworm increased gradually from 10.22 mg/ml on 1^{st} day to 12.85 mg/ml on day 8^{th} (Table 5). In inoculated silkworm, the total hemolymph lipid have shown increasing trend from 1^{st}

(10.18 mg/ml) to 4^{th} day (10.12 mg/ml) and decreasing trend from 5^{th} day onwards and reached 9.20 mg/ml by 8^{th} day from the treatment.

Table 5: Total lipid content in AmCPV treated and healthy silkworm, A. mylitta

			Г	Cotal lipid co	ontent mg/m	1		
Treatment				Days post i	noculation			
	1	2	3	4	5	6	7	8
AmCPV treated	10.18	10.59	10.25	10.12	9.88	9.73	9.51	9.20
Control	10.22	10.63	11.15	11.84	12.38	12.65	12.72	12.85
S.E. \pm	0.21	0.27	0.19	1.36	0.49	0.52	0.42	0.57
C. D. at 5%	1.93	1.27	1.21	1.52	1.71	1.94	2.62	2.13

The total carbohydrate and lipid contents increased to certain level and decreased steadily as the disease developed and it is reasonable to assume that as the disease progresses the number of pathogens increases and carbohydrates and lipids were utilized as a source of energy required for the growth and development of *AmCPV*.

4. Inference

The total haemocyte count (THC) increases gradually in healthy silkworm where as in AmCPV infected silkworm larvae in initial stages THC increases there after decreases. In the healthy larvae the prohaemocyte, plasmatocytes and granulocytes will be more in number whereas oenocytoids were less in number. The number of degenerated cells increases as the intensity of the disease increases. In initial infection of AmCPV, total protein, carbohydrate and lipid contents will be improved as like healthy silkworm but, when disease enhances it shows down ward trend.

References

- [17] Aizawa, K. (1971) Structure of polyhedra and virus particles of cytoplasmic polyhedrosis, In the cytoplasmic polyhedrosis of the silkworm (Eds: Aruga, H. and Tanada, Y.) Univ. Tokyo press, Tokyo, pp. 23-36.
- [18] Balavenkatasubbaiah, M., Natraju, B., Thiagrajan, V. and Datta, R.K. (2001) Haemocyte counts in different breeds of silkworm, *Bombyx mori* L., and their changes during progressive infection of BmNPV. *Indian J. Seric.*, 40(2), 158-162
- [19] Butt, T.M. and Shields, S.K. (1996) The structure and behaviour of gypsy moth (*Lymantria dispar*) haemocytes. J. Invertebr. Pathol., **68**, 1-14.
- [20] Dubois, M., Gilles, K. E., Hamilton, J.K., Rebers, P.A and Smith, F (1956) Calorimetric method of determination of sugars and related substances, Anal. Chem., 28, 350-356.
- [21] Folch, I., Ascoli, I., Less, M., Meath, J.A. and Le Baron, F.N (1951) Preparation of lipid extracts from brain tissue. J. Biol. Chem. 191, 833-841.

- [22] Gilium, M. and Shimanuki H. (1967) In vitro phagocytosis of Nosema apis spores by honey bee haemocytes. J. Invertebr. Pathol., 9, 387-389.
- [23] Horohov, D.W. and Dunn, P.E. (1983) Phagocytosis and nodule formation by hemocytes of *Manduca sexta* larvae following injection of *Pseudomonas aeruginosa*. *J. Invertebr. Pathol.*, **41**, 203-213
- [24] Jonesh, J.C. (1962) Current concepts concerning insect hemocytes Am. Zool., 2, 209-246
- [25] Lowry, O.H., Roserbrough, N.J., Farr, A.L. and Randal, R.J. (1951) Protein measurement with the folin-phenol reagent. J. Biol. Chem. 193, 265-275.
- [26] Nittono, Y. (1960) Studies on the blood cells in the silkworm, *Bombyx mori* L. Bull. Seric. Expt. Stn., 16, 261-266.
- [27] Saran, S.K., Mishra, P.K., Kumar, D., Singh, B.M.K., Sinha, B.R.R.P., Rai, S. and Pandey, P.N. (2002) Effect of total and differential haemocyte counts due to infection of *Nosema* species in *Antheraea mylitta* D.(Lepidoptera: Saturniidae) larvae. XIXth Congress of the International Sericultural Commission Proceedings. 21st – 25th Sept., Bangkok, Thailand. pp 308 – 311.
- [28] Snedecor, G.W. and Cockron, W.E. (1971) Statistical Methods. Oxford IHB Publishing Co. New Delhi, pp. 339 – 361.
- [29] Tauber, O.E. and Yeager, J.E (1935) On the total blood counts of insects. I. Orthoptera, Odonata, Hemiptera and Homoptera. Ann. Entomol. Soc. Am., 28, 229-240.
- [30] Wago, H. (1995) Host defense reaction of insects. Appl. Ent. Zool. 39.