Evaluation of *Adenanthera pavonina* Bark Extracts for Antioxidant Activity and Cytotoxicity against Cancer Cell Lines

Renilda Sophy A. J.\(^1\), Albin T. Fleming\(^2\)

\(^1\)PG & Research Department of Advanced Zoology and Biotechnology, Loyola College, Chennai, India -600034

\(^2\)Associate Professor, PG & Research Department of Advanced Zoology and Biotechnology, Loyola College, Chennai, India-600034

Abstract: Plant materials were used for the treatment of malignant diseases for centuries. Various studies on plant extract which have a suitable history of use in folklore for the treatment of cancer had often resulted in the isolation of principles with anti-tumour activity. In the present study, the anti-proliferative effect of acetone and methanol bark extracts of *Adenanthera pavonina* on three cancer cell lines using sulforhodamine-B assay and their antioxidant activities using 1-diphenyl-2-picrylhydrazyl radical and its reducing ability were investigated. NCI-H460, U251, and MCF7 cancer cell lines were used in this assay. The G150 value of methanol extract showed a lowest value of 27µg/mL against NCI-H460. The antioxidant and the reducing power assay showed a significant dose dependant activity. The study also suggests that antioxidant activity of these extracts might be one of the reasons for the anticancer potential of the extracts.

Keywords: Extracts, anti-tumor, cell lines, antioxidant, cytotoxicity

1. Introduction

It is being realized that there is an increase in the incidence of cancer worldwide in spite of cancer awareness, life style modification and new drugs. Bioactive principles were sought in the very early part of the nineteenth century, during which investigation of well known medicinal plants lead to the discovery of a number of biologically active alkaloids. Some of them like morphine, atropine, papavarine and codeine became cornerstones of many aspects of drug discovery. Majority of the cancer drugs originated from natural products like Taxol, Vinceristine and Camptothecin which saw the light by traditional in vitro cell based cytotoxic assays before their real molecular biological targets were identified. Today, with the advent of genomic research and new molecular biology tools for developing bioassays, more sophisticated biological assays in addition to cell-based assays are being employed routinely in the drug discovery paradigm. As a result, a large number of plant derived drugs continue to be discovered on the basis of traditional knowledge like ayurveda, siddha etc.

Oxidative stress is an important contributor to the pathophysiology of a variety of conditions including carcinogenesis, diabetes, cardiovascular dysfunctions, atherosclerosis, inflammation, drug toxicity, reperfusion injury and neurodegenerative diseases (Arumugam, 1998). Antioxidants are believed to play an important role in preventing or alleviating chronic diseases by reducing the oxidative damage to cellular components caused by reactive oxygen species (ROS) (Ceriello, 2003; Vinson, Dabbag, Serry, & Jang, 1995). Numerous plant constituents have shown free radical scavenging or antioxidant activity (Sunil & Ignacimuthu, 2011). There is a growing interest in natural phenolic antioxidants, present in medicinal and dietary plants, that might help attenuate oxidative damage (Rice-Evans, Miller, & Paganga, 1997; Silva, Ferreres, Malva, & Dias, 2005).

*Adenanthera pavonina* is a tree commonly called as red sandalwood, also known as the coral-wood or Barbados’ pride. It is largely populous in India, China, and in Costa Rica. The red sandalwood has been part of the alternative Ayurvedic medicine, as the tree is endemic in India. It is reported to have a rich amount of flavanoids (Rastogi and Mehrotraa, 1991) mainly gallic acid, terpenoids, tannis, sterols (beta-sitosterol, beta-sitosterol-3β –D-glucoside), triterpinoids (nonacosane and hentriacontane) and saponins (sapogenins) (Hussain Arshad and Hussain Sarfaraj, 2010). These phytochemicals are known to have many biological activities. In vitro studies show that *Adenanthera pavonina* leaf extract has antibacterial activity against the intestinal pathogen *Campylobacter jejuni* (Dholvitayakhun et al., 2012). Also, high doses of seed extract have an anti-inflammatory effect in studies in rats and mice (Olajide et al., 2004). We have used acetone and methanol extracts of *A. pavonina* to evaluate the anticancer potential and to determine the antioxidantive and free radical scavenging capabilities of the extract.

2. Materials and Method

Chemicals and reagents

DPPH (1,1-diphenyl,2-picrylhydrazyl), TCA (trichloro acetic acid), ferric chloride and BHT (butylated hydroxyl toluene) were obtained from Sigma chemical co., USA. Ascorbic acid was obtained from SD fine chem. Ltd., Biosar, India. All the other chemicals were of analytical grade.

Collection

Bark of *Adenanthera pavonina* were collected from the suburbs of Chennai, Tamil Nadu, India. The taxonomic
authentication was done by Dr. P. Jayaraman, Plant Anatomy Research Centre, Tambaram, Chennai. The voucher specimen number is PARC/2014/2022.

Preparation of Crude Extracts
The collected bark pieces were washed with running tap water, again washed with distilled water, air dried, homogenized to a fine powder and stored in air-tight bottles. Dried bark of *A. pavonina* was extracted with petroleum ether to remove wax using soxhlet extraction method. The bark powder was dried again and then extracted with acetone and methanol separately. The soxhlet was run until the solvent colour turned colourless. The extract was evaporated to dryness in a rotary vacuum evaporator. The extract was stored at 4°C in airtight sterile vials for further studies.

**DPPH Radical Scavenging Assay**

DPPH quenching ability of *A. pavonina* acetone and methanol extracts were measured according to Hanato, Kagawa, Yasuhara, and Okuda (1988). The methanol DPPH solution (0.15%) was mixed with different concentrations (200–1000 g/ml) of the extracts. After incubation for 30 min, the absorbance was read at 515 nm. The antiradical activity was expressed as IC50 (g/ml), (the antiradical dose required to cause a 50% inhibition). Ascorbic acid was used as standard. The ability to scavenge the DPPH radical was calculated using the following equation:

\[
\text{DPPH scavenging effect } (\%) = \frac{A_0 - A_1}{A_0} \times 100
\]

where, \(A_0\) is the absorbance of the control at 30 min, and \(A_1\) is the absorbance of the sample at 30 min. All samples were analysed in triplicate.

**Reducing Ability Assay**
The reducing power of *A. pavonina* acetone and methanol extracts were evaluated according to the method of Oyaizu, Kagawa, Yasuhara, and Okuda (1986). Different concentrations of the extracts in distilled water (200–1000 g/ml) were mixed with 2.5 ml of 0.2 M phosphate buffer (pH 6.6), and 2.5 ml of 1% K3Fe(CN)6. The mixture was incubated at 50°C for 20 min. After incubation, 2.5 ml of 10% TCA was added to the mixture and centrifuged at 3000 rpm for 10 min. The upper layer of the solution (2.5 ml) was mixed with distilled water (2.5 ml) and FeCl3 (0.5 ml, 0.1%), and the absorbance was measured at 700 nm. Increase in absorbance of the reaction mixture indicated the ability of reducing power. Butylated hydroxytoluene (BHT) was used as standard.

**Antiproliferative Assay**
The growth inhibitory activity of the acetone and methanol extracts of bark of *A. pavonina* were evaluated against cell line panel consisting of NCI-H460 (lung cancer), MCF7 (breast cancer) and U251 (brain cancer cell) using SRB assay. The cell lines were obtained from the American Type Culture Collection (ATCC). The cell lines were routinely maintained as monolayer cell cultures in Roswell Park Memorial Institute medium containing heat in-activated fetal bovine serum (10%, Gibco), and glutamine, penicillin and streptomycin solution (1%, L-glutamine, penicillin and streptomycin). Briefly, 100µL of cell suspension were plated in each well of 96-well plates, and incubated for 24 h at 37°C in a humidified CO2 (5%) incubator. The stock solutions of the two solvent extracts were prepared in dimethyl sulfoxide (DMSO) as a vehicle and various dilutions of the crude extracts (1, 0.5, 0.25, 0.125, 0.0625, 0.0312, and 0.0156 mg/ml) were added (100 µl) in each well. After 48 h of incubation, cold (4°C) trichloroacetic acid (50%, 100 µl) was added gently and left for 1 hour at 4°C, followed by washing with distilled water and air dried at room temperature. To each well, 100 µl of SRB solution was added and kept in dark for 30 minutes; the unbound stain was washed off with acetic acid (1%) and air-dried at room temperature. The protein bound stain was solubilised with 10mM tris-base (pH 10.2) with shaking for 5 min followed by the measures of the absorbance at 515 nm using a microplate reader. Camptothecin was used as positive control. GI50 which is the concentration of the extract or camptothecin causing 50% growth inhibition of cells was determined.

3. Results and Discussion

**DPPH radical scavenging activity**
The DPPH radical is a stable organic free radical with an absorption maximum band around 515–528 nm and thus, it is a useful reagent for evaluation of antioxidant activity of compounds (Sanchez-Moreno, 2002). In the DPPH test, the antioxidants reduce the DPPH radical to a yellow-coloured compound, diphenylpicrylhydrazine, and the extent of the reaction will depend on the hydrogen donating ability of the antioxidants (Bondent, Brand-Williams, & Bereset, 1997). It has been documented that cysteine, glutathione, ascorbic acid, tocopherol, polyhydroxy aromatic compounds (e.g., hydroquinone, pyrogallol, gallic acid), reduce and decolorize diphenylpicrylhydrazine by their hydrogen donating capabilities (Blois, 1958). The acetone and methanol extracts of *A. pavonina* exhibited a significant dose dependent inhibition of DPPH activity. Methanol exhibited a 50% inhibition (IC50) at a concentration of 400 µg/mL. The results are presented in figure 1.

**Reducing power**

It has been reported that reducing power is associated with antioxidant activity and may serve as a significant reflection of the antioxidant activity (Oktay et al., 2003). Compounds with reducing power indicate that they are electron donors, and can reduce the oxidized intermediates of lipid peroxidation processes so that they can act as primary and secondary antioxidants (Yen and Chen, 1995). For the measurements of the reductive ability, we studied the Fe3+ to Fe2+ transformation in the presence of *A. pavonina* extracts. Figure 2 shows the reducing capabilities of acetone and methanol extracts of *A. pavonina* compared to butylated hydroxyl toluene. The plant extract could reduce Fe3+ ions, and the reducing power was comparatively lesser than the standard of butylated hydroxyl toluene. The reducing power of methanol extract was very potent than acetone extract and the reducing power of the both extracts increased with quantity of sample.
Antiproliferative Activity

The lower GI50 values of methanol extract reflected a higher antiproliferative activity. The acetone extracts showed a comparatively less antiproliferative activity than methanol extract (Figure 3). The GI50 value of methanol was lowest against NCI-H460 with a GI50 value of 27µg/mL. The acetone extract showed highest GI50 value of 130µg/mL against U251. According to a previous study, the maximum growth inhibition shown by the crude extract of *Caralluma tuberculata* against MCF-7 cell line was 82% at a concentration of 500 µg/mL (Ahmed et al., 2009). Valko et al.,(2006) investigated the cytotoxicity effects of water extracts from leaves and branches of *Philadelphus coronarius* (*Hydrangeaceae*), against A431 cells (human skin carcinoma cell line) and the human breast adenocarcinoma cell line with various doses. Highest toxic effects were observed against MCF-7 cell line. The result of the present study is in accordance with the effect of crude methanol extract of the stem of *Debregeasia salicifolia* on the growth of MCF-7 cell line investigated by the MTT assay. *D. salicifolia* was also highly active against MCF-7 cell line. The maximum percentage inhibition value obtained for *D. salicifolia* was 99%. Minimum percentage inhibition was observed at extract concentration of 10 µg/mL that was 37.9% (Nisa et al.,2011). The present study showed dose dependent response. Other related research also showed that antiproliferative activity or cytotoxicity were in a dose-dependent manner (Yuan and Walsh, 2006). In accordance with the present study, the ethanol extract of peel and flesh of *Coleus tuberosus* showed both antioxidant and cytotoxic activity against cancer cell lines (Nugraheni et al., 2011). Moreover, the antioxidant rich sea weeds *Dictyota dichotoma*, *Hormophysa triquerta*, *Spatoglossum asperum*, *Stoechospermum marginatum* and *Padina tetrastromatica* comprehensively promoted anti pancreatic tumor potential, as evidently demonstrated by inhibitions of cell viability, proliferation and induced apoptotic cell death. With increasing concentrations of the algal polyphenol fractions, a dose dependent inhibition of cell viability in MiaPaCa-2 cells exposed to dichloromethane or ethyl acetate fractions was observed. (Aravindan et al.,2013).

4. Conclusion

This study, using in vitro assay systems suggested that *A. pavonina* extracts possessed anticancer potential. The antioxidant potential evaluated based on DPPH scavenging activity and the reducing power of the extracts confirmed the antioxidant and free radical scavenging activity of the extract. The antioxidant property of the extracts showed a correlation with the anticancer property of the extracts. This study suggests that the antioxidant activity of these extracts might be helpful in preventing or slowing the progress of oxidative stress-related diseases like cancer. Identification of the antioxidant constituents of the plant which are helpful for the anti-cancer properties are yet to be studied.

References


Author Profile

Renilda Sophy A. J.  PG & Research Department of Advanced Zoology and Biotechnology, Loyola College, Chennai, India-600034.

Albin T. Fleming.  Associate Professor,  PG & Research Department of Advanced Zoology and Biotechnology, Loyola College, Chennai, India - 600034.