Ventilator Associated Pneumonia

Snigdha Rashmi¹, Bhanu Kumar², Sudarshan Murthy³, Prasanna Kumar⁴, Tejashwi. S. Hosmani⁵

¹PG Resident, ⁵Department of Medicine, Affiliated to JSS Medical College, A constituent of JSS University- Mysore, Karnataka, South India

²Associate Professor, Department of Medicine, Affiliated to JSS Medical College, A constituent of JSS University- Mysore, Karnataka, South India

³Professor and HOD Department of Medicine, Affiliated to JSS Medical College, A constituent of JSS University- Mysore, Karnataka, South India

⁴Assistant Professor Department of Medicine, Affiliated to JSS Medical College, A constituent of JSS University- Mysore, Karnataka, South India

⁵PG resident Department of Medicine, Affiliated to JSS Medical College, A constituent of JSS University- Mysore, Karnataka, South India

Abstract: Ventilator Associated Pneumonia (VAP) refers to a type of pneumonia that occurs more than 48-72 hours after end tracheal intubation .This is one of the most common nosocomial infections in ventilated individuals.^{1,2} VAP occurs in 9-27% of intubated atients.^{3,4} Delay in initiating appropriate antibiotic therapy can increase the mortality associated with VAP.^{5,6} Initial empirical therapy can be modified based on the knowledge of local microbiological data, patient characteristics, and sensitivity pattern of expected pathogens at the institution. One of the consequences of increasing antimicrobial resistance is greater probability of inappropriate empiric antibiotic therapy.⁷ The aim of this study was to find the incidence of VAP and mortality associated with it. We also aimed to find frequency of pathogens isolated from the tracheal aspirate of patients diagnosed with VAP and the correlation of pathogen with duration of ICU stay.

Keywords: Ventilatory associated pneumonia; antibiotic sensitivity; pathogens; mortality.

1. Objectives

To find out Microbiological pattern of VAP in patients on ventilator along with Correlation of pathogen with duration of stay and mortality

a) Inclusion Criteria

Patients with VAP, more than 18yrs with consent were included

b) Exclusion Criteria

Patients having prior pneumonia and ARDS ,less than 18 yrs were excluded.

2. Methodology

a) Source of Data

Patients admitted in the critical care areas of JSS Hospital

b) Method of Collection of Data

This was a prospective cohort study done over a period of 1 year. Total of 86 who met the criteria were included. VAP was diagnosed using clinical pulmonary infection score which was evaluated on day2, 4, 7, and 10. CPIS> 6 was used diagnostic. Early onset VAP within 4 days of ventilation and late onset VAP after 4 days of ventilation. Endotracheal aspirate was preferred over protected specimen brush (PSB) sampling and bronchi-alveolar lavage(BAL), as these techniques are more invasive without mortality benefit over endotracheal aspirate

c) Endotracheal Aspirate:

Endotracheal aspirate was obtained under aseptic precautions using a 22-inch Ramson's 12F suction catheter with a mucus extractor as per standard protocol. After this, 2ml of 0.9% saline was injected into ET to flush the exudates into sterile container and immediately taken to the laboratory.

3. Statistical Methods

3.1 Descriptives

The Descriptive procedure displays univariate summary statistics for several variables in a single table and calculates standardized values (z scores). Variables can be ordered by the size of their means (in ascending or descending order), alphabetically, or by the order in which you select the variables (the default).

3.2 Crosstabs (Contingency table analysis)

The Crosstabs procedure forms two-way and multi-way tables and provides a variety of tests and measures of association for two-way tables. The structure of the table and whether categories are ordered determine what test or measure to use.

3.3 Chi square test

The Chi-Square Test procedure tabulates a variable into categories and computes a chi-square statistic. This goodness-of-fit test compares the observed and expected frequencies in each category to test either that all categories contain the same proportion of values or that each category contains a user-specified proportion of values.

3.4 One way ANOVA and Scheffe's post hoc test

The One-Way ANOVA procedure produces a one-way analysis of variance for a quantitative dependent variable by a single factor (independent) variable. Analysis of variance is used to test the hypothesis that several means are equal. This technique is an extension of the two-sample t test.

4. Results



Graph 1: Demographics

The maximum number of patients were in the age group 51-60, includes a total of 11 patients (36.7), followed by 5 (16.7%) in the age group 71-80 years. From the above table and graph we infer that there is no significant relation between the development of VAP and age groups

Table 2: Sex Distribution					
		Frequency	Percent		
Valid	Male	19	63.3		
	Female	11	36.7		
	Total	30	100.0		



Graph 2: Sex Distribution between Patients

Out of 30 patients diagnosed with VAP, 19(63.3%) were males and 11(36.7%) were females. In this study, there was an inclination towards male gender but the difference was statistically not significant.



Polymicrobials

Out of 30 with VAP, 18(60%) had monobacterial and 12(40%) polybacterial infections. In this study monomicrobial infections were more than the polymicrobials infections but no statistical significance.

Table 5: Outcomes of Patients with VAP, with
Monomicrobials and Olymicrobials

			TYPE		Total	
			Mono	Poly	Total	
OUTCOME	Ext	Count	6	1	7	
		% of TYPE	33.3%	8.3%	23.3%	
	Deat	Count	12	11	23	
	h	% of TYPE	66.7%	91.7%	76.7%	
Total		Count	18	12	30	
		% of TYPE	100.0%	100.0%	100.0%	



Graph 4: Outcomes of Patients with VAP, With Monomicrobials and Polymicrobials

Out of 18 patients with monomicrobial VAP, 12 (66.7%) expired where as out of 12 patients with polymicrobial VAP 11(91.7%) expired. Thus in our study the mortality was high with polymicrobial VAP.

International Journal of Science and Research (IJSR) ISSN (Online): 2319-7064 Index Copernicus Value (2013): 6.14 | Impact Factor (2013): 4.438

Monomicrobials and Olymicrobials						
		OUTCO		COME	Total	
			Ext	Death		
	ACR	Count	3	7	10	
		%of OUTCOME	42.9%	30.4%	33.3%	
	KP	Count	1	2	3	
		%of OUTCOME	14.3%	8.7%	10.0%	
	EC	Count	2	0	2	
		%of OUTCOME	28.6%	.0%	6.7%	
	PDM	Count	0	2	2	
		%of OUTCOME	.0%	8.7%	6.7%	
	CPS	Count	0	1	1	
Types		%of OUTCOME	.0%	4.3%	3.3%	
Types	KP+ACR	Count	0	7	7	
		%of OUTCOME	.0%	30.4%	23.3%	
	ACR+EC	Count	0	1	1	
		%of OUTCOME	.0%	4.3%	3.3%	
	EC+PDM	Count	1	0	1	
		%of OUTCOME	14.3%	.0%	3.3%	
	EC+KP	Count	0	1	1	
		%of OUTCOME	.0%	4.3%	3.3%	
	ACR+PD	Count	0	2	2	
	М	%of OUTCOME	.0%	8.7%	6.7%	
]	Fotal	Count	7	23	30	
		%of OUTCOME	100.0%	100.0%	100.0%	





Graph 5: Association of Outcomes And Microbes

A. Pattern of monomicrobial infection and its outcome

Out of 18 patients who developed monomicrobial VAP, the commonest organism isolated was A. baumannii amounting to10 (55.5%) followed by Klebsiella pneumonia 3(16.67%), E.coli2 (11.11%), P.aeruginosa2 (11.11%), coagulase positive S.aureus 1(5.55%). Of the 10 patients with Acinetobacter infection 7 expired (58.3%) and 3 were extubated (50%).

B. Pattern of polymicrobial infection and its outcome

Out of 12 patients who developed polymicrobial VAP ,the most common combination of organisms was K.pneumoniae

with Acinetobacter baumannii amounting to 7 isolates(58.3%) followed by combination of Acinetobacter baumanii and Pseudomonas aeruginosa amounting to 2 (16.6%). Out of 11 deaths in polymicrobial VAP, mortality was maximum with K.pneumoniae with Acinetobacter baumannii (63.63%). 15(65.2%) patients expired and 4(57.1%) patients were extubated. Of the patients who developed late VAP 8(34.8%) patients expired and 3(42.9%) patients were extubated

Organism N		Mean	Std.	Std. Error	
			Deviation		
ACR	10	8.3000	6.20125	1.96101	
KP	3	9.6667	2.51661	1.45297	
EC	2	10.0000	7.07107	5.00000	
PDM	2	7.0000	2.82843	2.00000	
CPS	1	20.0000			
KP+ACR	7	6.4286	.97590	.36886	
ACR+EC	1	12.0000	•	-	
EC+PDM	1	7.0000	•	-	
EC+KP	1	6.0000			
ACR+PDM	2	7.5000	3.53553	2.50000	
Total	30	8.3667	4.67188	.85296	

 Table 8: Association Between Duration of Stay and

 Date a sume

From the above table, we interpret that the correlation between the duration of stay and the microbiological pattern is statistically not significant.

Note:

Im-Imipenem, Tig-Tigecycline, Col-Colistin, Am-Amikacin, Azi-Azithromycin, Cefo-Cefoperazone, Ceft-Ceftriaxone, Cip-Ciprofloxacin, Clin-Clindamycin, Do-Doxycycline, Gati-Gatifloxacin, Gent-Gentamycin, Mer-Meropenem, Lin-Linezolid, PPN+TZ-peracillin+Tazobactam, Van-

Vancomycin, Cotri-Cotrimoxazole, Azt-Aztreonam, Ticar-Ticarcillin, Ert-Ertapenem



Graph 6: A. Drug Sensitivity Of Acinetobacter Baumannii



Graph 6: B. Sensitivity Ofklebsiella Pneumoniae

Organism	No.of.isolates	Sensitive	Resistant
A.baumannii	20	Im(11),Tig(20),Col(4)	Am(20), Azi(20), Azt(20), Cefo(20), Ceft(20),
			Cip(20), Clin(20), Do(20), Gati(20), Gent(20)
			Im(9),Col(16),Mer(20), Lin(20), PPN+TZ(20),Van(20)
K.pneumoniae	11	Am(5),Cip(2),Ert(1),Im(7),	Am(6),Cip(9), Ert(10),Im(4),PPN+TZ(9),
		PPN+TZ(2),Tig(11),Col(7)	Col(4),Azi(11),Azt(11), Cefo(11) Ceft(11),
			Clin(11),Do(11),Gati(11),Gent(11), Mer(11), Lin(11),
			Van(11)
P.aeruginosa	5	Gati(1),Gent(1),Im(1),	Gati(4),Gent(4) Im(4),Ticar(4)PPN+TZ(4),
		Ticar(1),PPN+TZ(1),Col(5)	Am(5),Cip(5),Ert(5),Azi(5), Azt(5),Cefo(5)
			Ceft(5),Clin(5) Do(5),Mer(5),Lin(5),Van(5)
E.coli	5	Am(4),Cefo(1),Cip(2),Cotri(2,	Am(1),Cefo(4)
		Ert(3),Im(3),Mer(3)Tig(4)	,Cip(3),Cotri(3),Ert(2),Im(2),Mer(2),Tig(1),Gati(5),Gent(5),Ti
			car(5),
			Azi(5)Azt(5),Cefo(5) Ceft(5),Clin(5)Do(5),Lin(5)
			,Van(5)PPN+TZ(5)
Coagulase Positive	1	Cip(1),Clin(1),Cotri(1),Ery(1),	Am(1),Cefo(1)
Staphylococci		Lin(1),Pris(1)	,Cip(0),Cotri(0),Ert(1),Im(1),Mer(1),Tig(1),Gati(1),Gent(1),
			Ticar(1),Azi(1),Azt(1),Cefo(1),Ceft(1),Clin(0),Do(1),Lin(0),V
			an(1),PPN+TZ(1)



Graph 6: C Sensitivity of Pseudomonas Aeruginosa



Graph 6: D.Drug Sensitivity Pattern Of Escherichia Coli



Graph 6: E. Sensitivity Of P. Aeruginosa



Graph 6: f. Sensitivity of E. Coli



Graph 9: E. Sensitivity of Coagulase Positive S. Aureus

From the above tables, all the organisms isolated were resistant to all antibiotics that are being routinely used in ICU setting. The acetinobacter isolates were most sensitive to tigecycline and imipenem. The isolates of Klebsiella were sensitive to tigecycline, colistin and imipenem. The isolates of pseudomonas were sensitive to tigecycline and imipenem. The isolates of Escherichia coli were sensitive to amikacin and tigecycline. The coagulase positive staphylococcus was sensitive to ciprofloxacin and clindamycin.

5. Discussion

The incidence of VAP in our study was 34.8%.this correlates with other studies in which the incidence of VAP was 15.5 to 47%, depending on the diagnostic criteria used.^{48, 49, 50, and 51}

There was no statistically significant association between the age of the patient and development of VAP, indicating that age is neither predisposing nor protective factor for the development of VAP. There was no statistically significant association between the gender of the patient and development of VAP, and both males and females are equally predisposed to VAP.⁵²

Table 9	Drug	Sensitivity	Patterns
---------	------	-------------	----------

		Diug Schshivity I			
	Fagon and colleagues,		Panwar and	Gupta and	Our study
	1984 ⁵⁰	colleagues,1990 ⁴⁸	colleagues,2005 ⁵¹	colleagues, 2010	
Incidence of VAP(%)	27.5	24	47	28.04	34.8
Mortality rate (%)	53	33	37	32.71	76.6
Technique used	Protected specimen	Protected specimen	Tracheal aspirate	Tracheal aspirate	Tracheal
	brush	brush, BAL			aspirate
P.aeruginosa	16(31%)	7(28%)	11(46%)	9(30%)	5(16.66%)
S.aureus	17(33%)	5(20%)	6(25%)	8(26.67%)	-
Acinetobacter	8(15%)	6(24%)	2(8%)	6(20%)	20(66.66%)
K.pneumoniae	2(4%)	3(12%)	7(29%)	7(23.33%)	11(36.66%)
Proteus	8(15%)	-	3(13%)	-	-
E.coli	4(8%)	3(12%)	3(13%)	-	5(16.66%)
S. pneumoniae	3(6%)	1(4%)	-	1(3.33%)	-
Coagulase positive staph. Aureus	-	-	-	-	1(3.33%)

The most common organism isolated was Acinetobacter baumannii which was the common pathogen in study by Arindam Dey and Indira Bairy.⁵⁴ Out of 20 isolates, 14 were from patients with early onset VAP and 6 from late onset VAP.The next common organism was Klebsiella pneumoniae with total of 11isolates.Out of these 11 isolates, 7 isolates were in early onset VAP, followed by E.coli 5 isolates and pseudomonas with 5 isolates. The least common was of Coagulase positive Staphylococcus aureus with only 1. There is no significant association between the microbiological patterns of VAP in our hospital with duration of stay.

Analysis of antibiotic sensitivity pattern suggests that these organisms were highly resistant to commonly used antibiotics in our hospital. All 20(100%) isolates of Acinetobacter were sensitive to tigecycline, 11(55%) isolates were sensitive to imipenem 4(20%) to colistin. VAP due to multidrug resistant acetinobacter sensitive to tigecycline is common in our setting.^{35,38,39,40,46,47} Out of 11 isolates of all 11(100%) Klebsiella were sensitive to tigecycline,7(63.63%) were sensitive to imipenem and 5(45.45%) to amikacin and 2 (18.18%) to ciprofloxacin and piperacillin and tazobactam, suggesting a high prevalence of beta-lactamase(ESBL) extended spectrum producing organisms in our setting. All the 5(100%) isolates of Pseudomonas aeruginosa were sensitive to colistin. 4(80%) out of 5 isolates E.coli was found sensitive to tigecycline and amikacin.3 isolates(60%) were sensitive to carbapenems This observation suggests that the organisms isolated in etiology of VAP in our hospital were resistant to commonly used antibiotics like fluoroquinolones, cephalosporins and aminoglycosides but showed variable sensitivity to tigecycline, colistin and carbapenems. Mortality of VAP group was 76.6% which is very high. A similar high mortality was found in other studies.53

6. Conclusions and Summary

Thus, we conclude that most of the VAP cases in our setting were early onset VAP & majority of these are caused by highly resistant strains.VAP due to multidrug resistant acetinobacter sensitive to tigecycline is common in our setting. And local epidemiological data should be connected at all centres, as the information can help in guiding the initial empirical antibiotic therapy, which would be more rationale and help in decreasing mortality and morbidity. An effective empirical antibiotic therapy would also help in preventing development of more resistant strains. CLINICAL UTILITY: To choose the most appropriate empirical antibiotics

References

- [1] Singh N, Rogers P, Atwood CW, et al. Short-Course Empiric Antibiotic Therapy for Patients with Pulmonary Infiltrates in the Intensive Care Unit. A Proposed Solution for Indiscriminate Antibiotic Prescription. Am J Respir Crit Care Med., 2000; 162(2 Pt 1): 505-11.
- [2] Namias, samiian L, Nino D, Shirazi E, O'Neill K, Kett DH.Incidence and susceptibility of pathogenic bacteria vary between intensive care units within a single

hospital: implications for empiric antibiotic strategies.J Trauma 2000; 49:638-45.

- [3] Rello J,Quintana E,Ausina V,Castella J,Luquin M,Net A,Prats G:Incidence,etiology,and outcome of nosocomial pneumonia in mechanically ventilated patients.
- [4] Estes RJ, Meduri GU. The pathogenesis of ventilatorassociated pneumonia: I. Mechanisms of bacterial transcolonization and airway inoculation. Intensive Care Med 1995; 21(4):365–383.
- [5] de la Torre FJ, Pont T, Ferrer A, Rossello J, Palomar M, Planas M. Pattern of tracheal colonization during mechanical ventilation. Am J Respir Crit Care Med 1995;152(3):1028–1033.
- [6] Niederman MS, Mantovani R, Schoch P, Papas J, Fein AM. Patterns and routes of tracheobronchial colonization in mechanically ventilated patients. The role of nutritional status in colonization of the lower airway by *Pseudomonas* species. Chest 1989; 95(1):155–161.
- [7] Zeiher BG, Hornick DB. Pathogenesis of respiratory infections and host defenses. Curr Opin Pulm Med 1996;2(3):166–173.
- [8] Koerner RJ. Contribution of endotracheal tubes to the pathogenesis of ventilator-associated pneumonia. J Hosp Infect 1997;35(2):83–89.
- [9] Valles J, Artigas A, Rello J, Bonsoms N, Fontanals D, Blanch L, etal. Continuous aspiration of subglottic secretions in preventing ventilator-associated pneumonia. Ann Intern Med 1995; 122(3):179–186.
- [10] Mahul P, Auboyer C, Jospe R, Ros A, Guerin C, el Khouri Z, et al. Prevention of nosocomial pneumonia in intubated patients: respective role of mechanical subglottic secretions drainage and stress ulcer prophylaxis. Intensive Care Med 1992; 18(1):20–25.
- [11] Kyle UG, Genton L, Heidegger CP, et al. Hospitalized mechanically ventilated patients are at higher risk of enteral underfeeding than non-ventilated patients. Clin Nutr. 2006; 25 (5):727 - 735.
- [12] Shorr AF, Duh MS, Kelly KM, Kollef MH. Red blood cell transfusion and ventilator-associated pneumonia: A potential link? Crit Care Med 2004; 32: 666-674.
- [13] Zur KBMandell DLGordon REHolzman IRothschild MA Electron microscopic analysis of biofilm on endotracheal tubes removed from intubated neonates. *Otolaryngol Head Neck Surg* 2004; 130407-414.
- [14] Protera C forging a link between biofilms and diseases. Science 1999;2831837-1839
- [15] Stewart PS Costerton JW Antibiotic resistance of bacteria in biofilms.Lancet 2001; 358135-138.
- [16] Fabregas N,Ewig S,Torres A,EL-Ebiary M,Ramirez J,de La Bellacasa JP,Bauer T,Cabello H:Clinical diagnosis of ventilator-associated pneumonia.
- [17] Marquette CH, Copin MC, Wallet F et al.Diagnostic tests for pneumonia in ventilated patients: prospective evaluation of diagnostic accuracy using histology as a diagnostic gold standard. Am J Respir Crit Care Med 1995; 151:1878-88.
- [18] Marquette CH, Georges H, Wallet F, et al. Diagnostic efficiency of endotracheal aspirates with quantitative bacterial cultures in intubated patients with suspected pneumonia: comparison with the protected specimen brush. Am Rev Respir Dis1993; 148:138-44.

- [19] Torres A, Martos A, Puig de la Bellacasa J, et al Specificity of endotracheal aspiration, protected specimen brush, and bronchoalveolar lavage in mechanically ventilated patients. Am Rev Respir Dis 1993; 147:952-7.
- [20] Fagon JY, Chastre J, Wolff M, et al. Invasive and noninvasive strategies for management of suspected ventilator-associated pneumonia: a randomized trial. Ann Intern Med 2000; 132:621-30.
- [21] Souweine B, Veber B, Bedos JP, et al. Diagnostic accuracy of protected specimen brush and bronchoalveolar lavage in nosocomial pneumonia: impact of previous antimicrobial treatments. Crit Care Med 1998; 26:236-44.
- [22] Fujitani S, Yu VL Diagnosis of ventilator-associated pneumonia: focus on non bronchoscopic techniques (non bronchoscopic bronchoalveolar lavage including mini-BAL, blinded protected specimen brush, and blinded bronchial sampling) and endotracheal aspirates. J Intensive Care Med 2006; 21:17-21.
- [23] Torres A, Puig de la Bellacasa J, Rodriguez Roisin R, Jimenez de Anta MT, Agusti-Vidal A. Diagnostic value of telescoping plugged catheters in mechanically ventilated patients with bacterial pneumonia using the Metras catheter. Am Rev Respir Dis1988; 138:117-20.
- [24] Fartoukh M, Maitre B, Honore S, Cerf C, Zahar JR, Brun-Buisson C (2003) Diagnosing pneumonia during mechanical ventilation: the Clinical Pulmonary Infection Score revisited. Am J Respir Crit Care Med 168:173– 179
- [25] Luyt CE, Chastre J, Fagon JY. Value of the clinical pulmonary infection score for the identification and management of ventilator-associated pneumonia. *Intensive Care Med.* 30(5), 844–852(2004).
- [26] Zilberberg MD, Shorr AF. Ventilator-associated pneumonia: the clinical pulmonary infection score as a surrogate for diagnostics and outcome. *Clin. Infect. Dis.* 51 (Suppl. 1), S131–S135 (2010).
- [27] Alvarez-Lerma F.Modifications of empiric antibiotic treatment in patients with pneumonia acquired in the intensive care unit:ICU-Acquired Pneumonia Study Group. Intensive Care Med 1996;22:387-394.
- [28] Heyland DK,Cook DJ,Griffith L et al.The attributable mortality and morbidity of ventilator-associated pneumonia in the critically ill patient.AMJ Respir Crit Care Med 1999;159:1249-1256.
- [29] Trouillet JL, Chastre J, Vuagnat A, et al. Ventilator associated pneumonia caused by potentially drug resistant bacteria. AM J Respir Crit Care Med 1998;157:531-539.
- [30] Kollef MH.The Prevention of Ventilator associated pneumonia.N Engl J Med 1999; 340:627-634.
- [31] Coffin, S, et al. Strategies to Prevent Ventilator-Associated Pneumonia in Acute Care Hospitals. Infect Control Hosp Epidemiol 2008; 29:S31-S40.
- [32] Tigecycline for the treatment of multidrugresistant(including Carbapenem-resistant) Acinetobacter infections:a review of the scientific evidence.J antimicrob chemother .july 2008 62:145-55
- [33] Acinetobacter baumanii:Epidemiology,Antimicrobial Resistance,and treatment options.Clinical Infectious Diseases.April 2008 46:8 1254-1263

- [34] Wates KB, Duffy LB, Dowzicky MJ.Antimicrobial susceptibility among pathogens collected from hospitalized patients in the United States and in vitro activity of tigecycline,a new glycylcycline antimicrobial.Antimicrob Chemother.2006 Agents oct;50(10):3479-84
- [35] Drakulovic MB, Torres A, Bauer TT, Nicolas JM, Nogué S, Ferrer M: Supine body position as a risk factor for nosocomial pneumonia in mechanically ventilated patients: a randomised trial. *Lancet* 1999, 354:1851-1858.
- [36] Li Bassi G, Torres A: Ventilator-associated pneumonia: role of positioning.*Curr Opin Crit Care* 2011, 17:57-63.
- [37] Muscedere J, Rewa O, McKechnie K, Jiang X, Laporta D, Heyland DK: Subglottic secretion drainage for the prevention of ventilator-associated pneumonia: a systematic review and meta-analysis. *Crit Care Med* 2011, 39:1985-1991.
- [38] Griffiths J, Barber VS, Morgan L, Young JD: Systematic review and meta-analysis of studies of the timing of tracheostomy in adult patients undergoing artificial ventilation.*BMJ* 2005, 330:1243.
- [39] Terragni PP, Antonelli M, Fumagalli R, *et al.*: Early vs late tracheotomy for prevention of pneumonia in mechanically ventilated adult ICU patients: a randomized controlled trialJAMA 2010, 303:1483-1489.
- [40] Baraibar J et al.Risk factors for infection for Acinetobacter baumanii in intubated patients with nosocomial pneumonia.Chest.1997 Oct;112(4):1050-4.
- [41] Husni RN,Goldstein LS et al.Risk factors for an outbreak of multi-drug-resistant Acinetobacter nosocomial pneumonia among intubated patients.Chest.1999 May;115(5):1378-82.
- [42] Torres A,Puig de la Bellacasa J,Xaubet A,Gonzalez J,Rodriguez-Roisin R,Jimenez de Anta MT,et al.Diagnostic value of quantitative cultures of bronchoalveolar lavage and telescoping plugged catheters in mechanically ventilated patients with bacterial pneumonia.Am Rev Respir Dis 1989;140:306-10
- [43] Kollef MH.Ventilator-associated pneumonia:A multivariate analysis.JAMA 1993;270:1965-70.
- [44] Fagon JY, Chastre J, Domart Y, Trouillet JL, Pierre J, Darne C, et al. Nosocomial pneumonia in patients receiving continuous mechanical ventilation: Prospective analysis of 52 episodes with use of protected specimen brush and quantitative culture techniques. Am Rev Respir Dis 1989;139:884
- [45] Panwar R,Vidya SN,Alka KD.Incidence,clinical outcome and risk stratification of ventilator-associated pneumonia:A prospective cohort study.Indian J Crit Care Med 2005;9:211-6.
- [46] Apostolopoulou E,Bakakos P,Katostaras T,Gregorakos L.Incidence and risk factors for Ventilator- associated pneumonia in 4 multidisciplinary intensive care units in Athens,Greece.Respir Care 2003;48:681-8.
- [47] Pennigton JE.nosocomial respiratory infection. In : mandell GL,douglas RG Jr,bennet JE,editors. Principles and practice of infectious disease. St.louis,MO:Churchill livingstone;1990.p.2199-2205.
- [48] Arindam Dey,Indira Bairy.Incidence of multidrugresistant organisms causing ventilator-associated

pneumonia in a tertiary care hospital: A nine months prospective study. Ann Thorac Med. 2007 Apr-Jun; 2(2):52-57.

Author Profile



Dr. M. Bhanukumar, MD in medicine working as a associate professor in the medicine department-Affiliated JSSmedical collage of JSS University- Mysore, Karnataka-India



Prof. (Dr.) Sudershan Murthy, MD in medicine. Head of the department of medicine - Affiliated JSS medical collage of JSS University-Mysore, Karnataka-India