

Evaluation of Acute Toxicity of Chromium to Freshwater Air Breathing Teleosts, *Channa gachua* (Bloch)

Komal

Department of Zoology, Patna Science College, Patna University, Patna, Bihar, India

Email: drkomal080321[at]gmail.com

Abstract: Heavy metals are naturally occurring elements that have a density at least five times that of water and a high atomic weight. These metallic elements are considered systemic toxicants because they are known to harm different organs, even at lower exposure levels. Water, the most valuable natural resources, is essential to all life. During the experiment, the freshwater fishes *Channa gachua* were exposed separately to different doses of chromium ($K_2Cr_2O_7$) for 24 hrs, 48 hrs, 72 hrs and 96 hrs. At first, we determined median lethal concentration (LC_{50}) of $K_2Cr_2O_7$ to fish using graphical interpolation method and confirmed it by probit analysis. Changes in locomotory and behavioral attributes were observed carefully and monitored properly. The treated fishes showed restlessness and rapid random movements. Rapid opercular movements were also observed.

Keywords: *Channa gachua*, lethal concentration, graphical interpolation method, probit analysis, chromium

1. Introduction

Nowadays, we live in an environment where the purity of water, air, and all heavy metals and other components is in question. Our changing human habits are exacerbating the problem of environmental pollution and contamination to such an extent that the survival of plants and animals is becoming increasingly difficult over time. The anthropogenic causes of environmental degradation are being ignored, which are directly or indirectly harming existing civilization and causing massive damage to human health and the economy at large.

Chromium (Cr) metal is thought to be one of the most prevalent and pervasive contaminants in aquatic environments, it does not occur naturally in its pure metallic mode (Kotas, et al., 2000; Saha, et al., 2011; Shadreck, et al., 2013; Prasad, et al., 2021). In its pure metallic form, chromium exists in three oxidation states: Cr (II), Cr (III), and Cr (VI). Its most unstable form is Cr (II) while Cr (III), and Cr (VI) are described to be in stable oxidation states (Espenson, J. H., 1970; Shupack, S. I. 1991; Apte, et al., 2006; Jiang, et al., 2019; Ao, M. et al., 2022). Notably, chromium metal is frequently utilized by many industrialists for his desirable purpose as electroplating, ore mining, tanneries, textile dyeing and printing, photography in addition with medical sectors and its waste particulates enter into aquatic medium (Narayani, et al., 2013; Siddiquee, et al., 2015; Dasan, et al., 2020; Kerur, et al., 2021).

Exposure of water pollutant through aquatic animal and their health issues are global concern. Of all aquatic animals, fish are the most sensitive to the pollutants because they are the main organisms in the system (Cooper, C. M., 1993; Camargo, et al., 2006; Jayaswal, et al., 2018; Qadri, et al., 2020; Zahoor, et al., 2023). Heavy metals can enter the fish body through contaminated food through the gills, skin or digestive tract. Various research have shown that fish are particularly sensitive to aquatic pollutants because they absorb and store heavy metals such as nickel and chromium

in their tissues (Kousar, et al., 2014; Javed, et al., 2019; Garai, et al., 2021; Naeem, et al., 2021; Agbugui, et al., 2022).

Metal bioaccumulation must be monitored in order to determine the process's temporal and spatial scope and to evaluate any possible effects on the health of the organism (Has-Schön et al., 2008). In fish exposed to acute and sub-lethal metal concentrations, bioaccumulation of numerous metals has been documented in the literature (Uysal et al., 2009; Kamunde and MacPhail, 2011). Whenever, fish exposed to various hazardous compounds can often exhibit sub-lethal effects that can be detected or diagnosed using biochemical and physiological biomarkers (Al-Ghanim, K.A. 2012; Keiter, S. 2013). Biochemical variables like antioxidants and enzyme activities and physiological variables like serum levels of metabolites, ions, and hormones are prominent among these biomarkers used as stress indicators (Speich, et al., 2001; Pasupathi, et al., 2009; Firat and Kargin, 2010; Atli, et al., 2015).

The morphological alterations that can be induced by Cr (VI) include epithelial hyperplasia, epithelial lifting, secondary gill lamellae degeneration, lamellar cell hyperplasia, and atrophy of the central axis in tissues. There is a lack of knowledge regarding the genetic and histological toxicity of Cr (VI) to aquatic species (Roberts, et al., 2004). The most significant industrial application for nickel is in the manufacturing of stainless steel. Because to its exceptional strength and durability, ability to resist corrosion, appealing appearance, advantageous thermal characteristics, electrical conductivity, and alloying ability. The production of alloys accounts for around 75% of all nickel consumption (Wilson et al., 1986, Nriagu and Pachana, 1988, Nicolaidou and Nott, 1989). The LC_{50} of chromium for common carp (*Cyprinus carpio*) was found to be 191 mg/L by Tayyabah et al. (2005). The dosage and time-dependent LC_{50} of Cr to *Channa marulius* was reported by Mallesh et al. (2015); Naz and Jawed (2015). As fish became older, the toxicity of chromium to them diminished.

Metal accumulation in the tissue and organs of fish (*Oreochromis aureus*) depends on the kind of metals and their concentrations, exposure duration, water physico-chemical characteristics, such as temperature, salinity, and hardness (Allen, 1995), metabolic processes, species type, and age (Pelgrom et al., 1995).

2. Materials and Methods

All of the purchased live *Channa gachua* (*C. gachua*) fish was cleaned by using 0.1% KMNO₄ to remove cutaneous infections. Fish that were healthy (average length 20-25 cm, weight 45-60 g) were chosen and transferred to large glass tank (aquariums) for stocking and acclimatization to various test settings, including light, temperature ($28 \pm 2^\circ\text{C}$), and water physiochemical characteristics, for a period of 15 days.

Water was recycled on alternate day. Tanks always covered with fish netting to prevent the escape of fishes.

Fish were fed with ad libitum, provided with a formulated dite and natural food. Many of the arteficial food as wheat powder, prawn meal, artemia eggs, vitamin B complex, and vitamin A was fed to the fish. To rectify the deficiencies caused by arteficial diet, natural foods like earth worm, and chopped soft meat was provided thrice a week. After feeding the remaining food particles were immediately removed from the tank bottom. Food was stopped one day before the experimental animal (*C. guchua*) which was sacrificed at the end of the experimental period.

Sample water (aquarium's water that were filled with ground/Tap water) were analysed as per the stander method of APHA (2020) for the physiochemical parameters as pH, TDS, TH, alkalinity, chloride, and dissolve oxygen (DO).

Results to the determination of LC₅₀ of chromium aquarium setup was arranged in laboratory premises of the Department of Zoology, Patna Science College, Patna University, Patna.

During the analysis of LC₅₀, stock solution of Potassium dichromate (K₂Cr₂O₇) was dissolved in deionized water and the stock solution (1000ppm) were prepared. The bioassay of acute toxicity for the chemical of this experiment, potassium dichromate (K₂Cr₂O₇) was carried out on the static system to determine the 24, 48, 72, and 96 hours LC₅₀ in the experimental fish, *C. gachua* for the definitive test, the scale finding experiment was executed first to measure the concentration of the experimental solution.

In the definitive test, ten fishes were used for every group. Group 1, 2, 3, 4, 5, and 6 were exposed to rising concentration of, potassium dichromate of (K₂Cr₂O₇) whereas the group 1 was retained, potassium dichromate (K₂Cr₂O₇) fresh water to serve as a control. The minimal concentrations of, potassium dichromate (K₂Cr₂O₇) tested were: 25ppm, 40ppm, 90ppm, 150ppm, and 180ppm. The exposure duration for the experiment was 24, 48, 72, and 96 hours. The test was replicated three times for the accuracy of this experiment.

By using probit analysis method; above experimental data represents the level of LC₅₀, along with toxicity level of potassium dichromate (K₂Cr₂O₇) upon the *C. gachua*.

Test Chemical

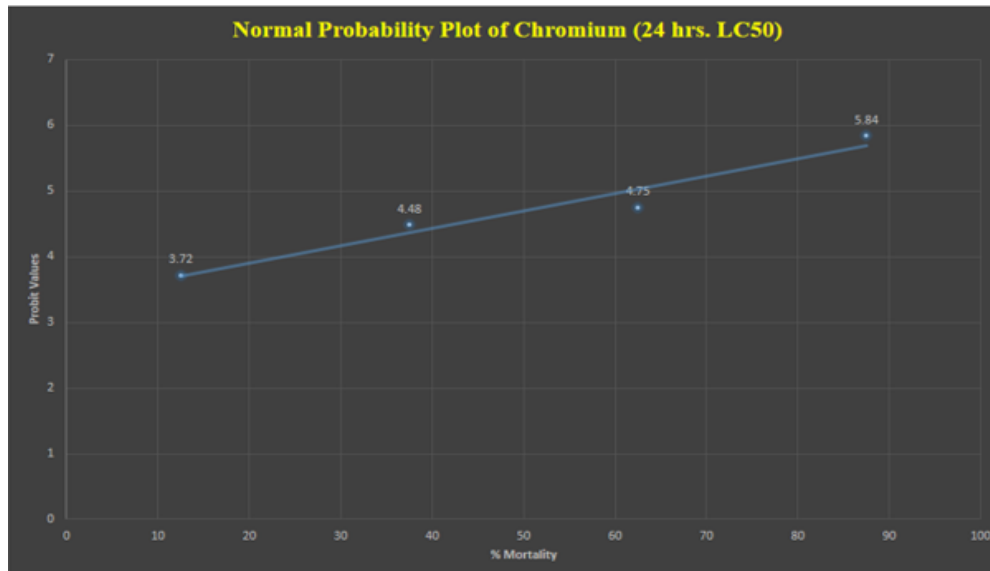
Analytical-grade Potassium dichromate (K₂Cr₂O₇) was used to prepare a stock solution in dechlorinated water, from which the required nominal test concentrations were obtained by serial dilution.

Table 1: Data shows LC₅₀ value of chromium as K₂Cr₂O₇, for a period of 24, 48, 72, and 96 hrs. in freshwater fish *C. gachua*.

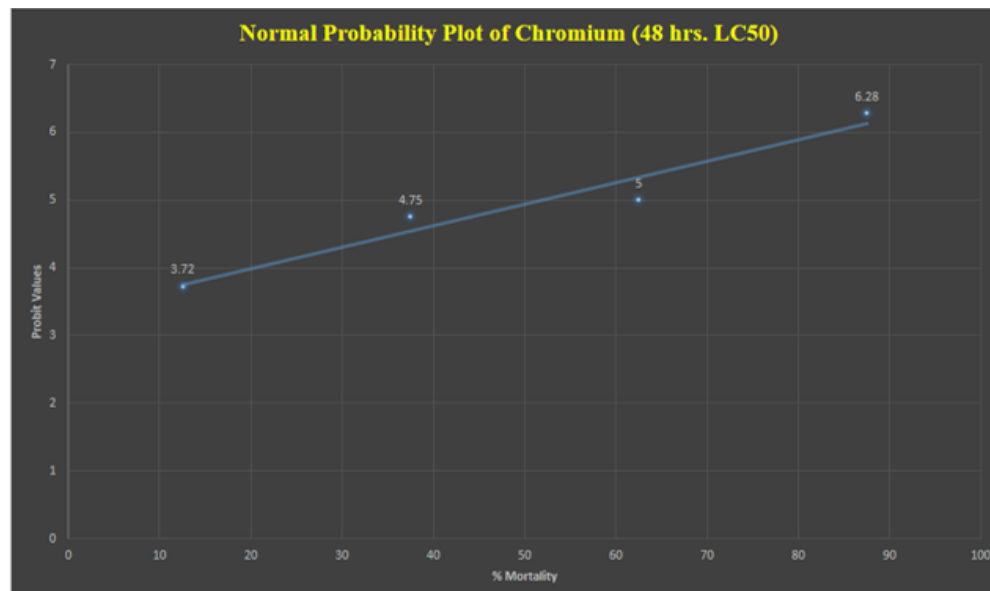
Aquarium No.	No. of test fish C.Gachua	Chromium Concentration	Mortality During							
			24hrs.		48 hrs.		72 hrs.		96 hrs.	
			No.	%	No.	%	No.	%	No.	%
Gr. I	10	Control	Nil	Nil	Nil	Nil	Nil	Nil	Nil	Nil
Gr. II	10	180ppm	8	70	9	90	10	100	-	-
Gr. III	10	150 ppm	4	40	5	50	7	70	8	80
Gr. IV	10	90 ppm	3	30	4	40	4	40	5	50
Gr. V	10	40 ppm	1	10	1	10	2	20	3	30
Gr. VI	10	25 ppm	Nil	Nil	Nil	Nil	Nil	Nil	Nil	Nil
Gr. VII	10	10 ppm	Nil	Nil	Nil	Nil	Nil	Nil	Nil	Nil

Probit Regression Analysis (Cr)

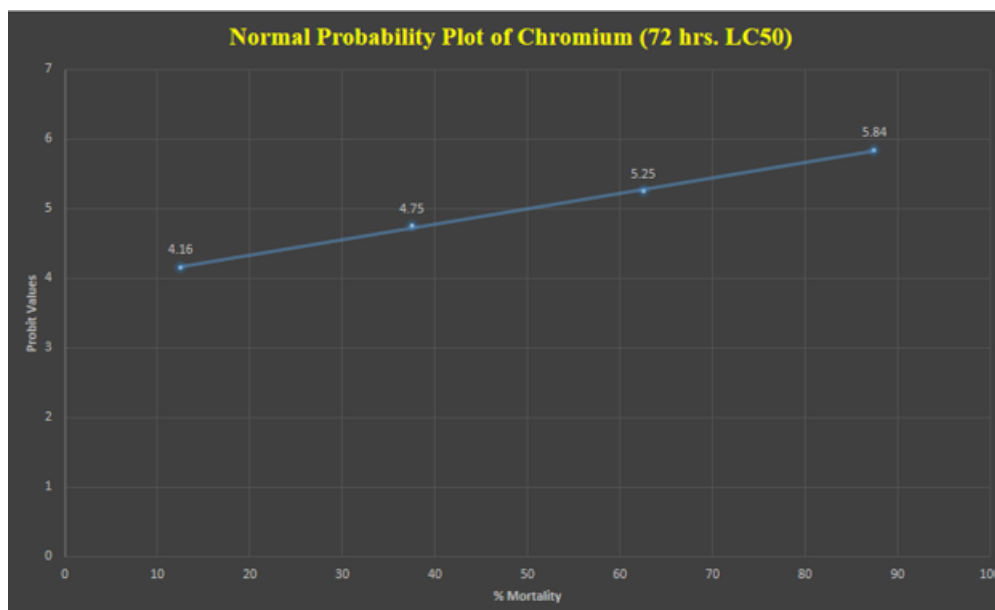
@ 24h



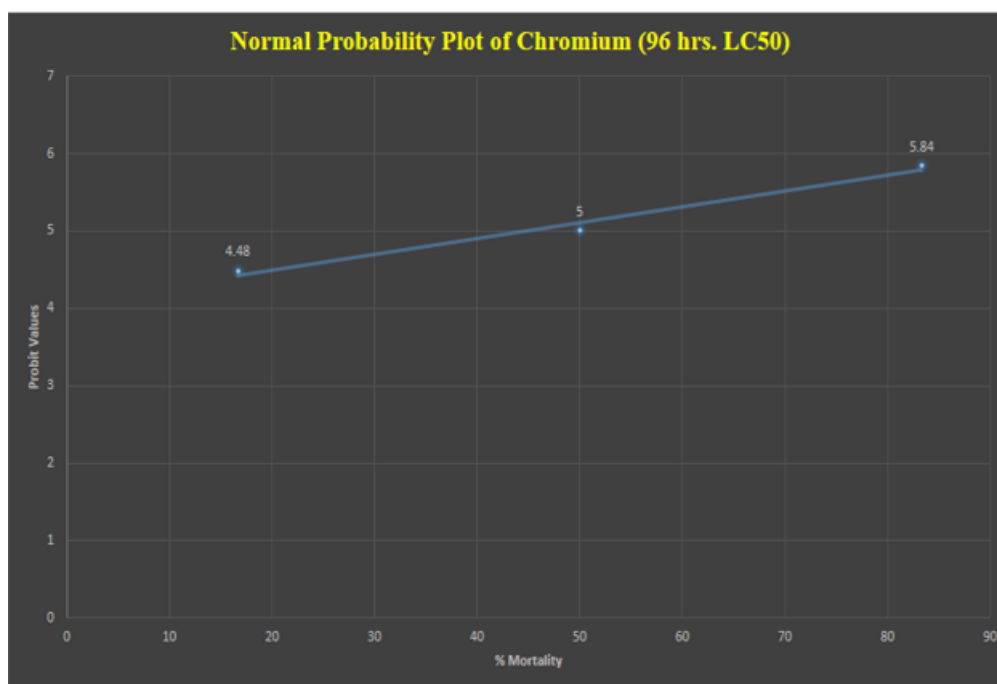
@ 48 h



@ 72 h



@ 96 h



3. Discussion

The present study establishes, for the first time, the acute toxicity profile of potassium dichromate in *Channa gachua*, with a 96-h LC₅₀ of 90 mg/L. Mortality increased in a clear concentration- and time-dependent manner, consistent with the classical acute toxicity pattern reported for cr across freshwater teleosts.

The concentration-dependent behavioural alterations noted in the present study– including surfacing/air-gulping changes, altered opercular movement, and abnormal swimming, Increased mucus secretion and surface discoloration, when observed, are consistent with a generalized stress and epidermal irritation response reported across chromium exposed fish species.

Because *C. gachua* relies on periodic surfacing for aerial respiration through its suprabranchial chamber, an ecological consideration previously highlighted in comparative toxicity work on this species⁵. This physiological trait strengthens the ecological relevance of *C. gachua* as a sentinel species for surface-water metal contamination in shallow wetland habitats, even though its acute LC₅₀ in the present study did not indicate exceptional sensitivity relative to its congener.

4. Conclusion

The fish *Channa gachua* were exposed to different concentrations of potassium dichromate. The clear concentration- and time-dependent mortality of this air-breathing species as a bioindicator for chromium contamination in freshwater The LC₅₀ values were calculated at 24 hours, 48 hours, 72 hours and 96 hours. We found here high values of the LC₅₀ which might have been due to various factors. Further studies are needed to understand the key factors that are playing vital role in variations in the LC₅₀ values in *C. gachua* 50%

References

- [1] Kori-Siakpere, O. and V.E. Egor., 1997. Haematological characteristics of the African mudfish *Clarias buthupogon* (Pisces: Clariidae). Bull. Sci. Ass. Nig. 21: 177-185.
- [2] Saha, R., Nandi, R., & Saha, B. (2011). Sources and toxicity of hexavalent chromium. Journal of Coordination Chemistry, 64(10), 1782-1806.
- [3] Shadreck, M., & Mugadza, T. (2013). Chromium, an essential nutrient and pollutant: review. African Journal of Pure and Applied Chemistry, 7(9), 310-317.
- [4] A Prasad, S., Yadav, K. K., Kumar, S., Gupta, N., Cabral-Pinto, M. M., Rezania, S., ... & Alam, J. (2021). Chromium contamination and effect on environmental health and its remediation: A sustainable approaches. Journal of Environmental Management, 285, 112174.
- [5] Shupack, S. I. (1991). The chemistry of chromium and some resulting analytical problems. Environmental Health Perspectives, 92, 7-11.
- [6] Siddiquee, S., Rovina, K., Azad, S. A., Naher, L., Suryani, S., & Chaikaew, P. J. J. M. B. T. (2015). Heavy metal contaminants removal from wastewater using the potential filamentous fungi biomass: a review. J Microb Biochem Technol, 7(6), 384-395.
- [7] Speich, M., Pineau, A., & Ballereau, F. (2001). Minerals, trace elements and related biological variables in athletes and during physical activity. Clinica Chimica Acta, 312(1-2), 1-11.
- [8] Pasupathi, P., Chandrasekar, V., & Kumar, U. S. (2009). Evaluation of oxidative stress, enzymatic and non-enzymatic antioxidants and metabolic thyroid hormone status in patients with diabetes mellitus. Diabetes & Metabolic Syndrome: Clinical Research & Reviews, 3(3), 160-165.
- [9] Firat, O., & Kargin, F. (2010). Individual and combined effects of heavy metals on serum biochemistry of Nile tilapia *Oreochromis niloticus*.

- Archives of environmental contamination and toxicology, 58, 151-157, 187 Anukriti (An International Peer Reviewed Multidisciplinary Refereed Research Journal) Vol. 15, No. 10, October 2025 Impact Factor: 8.0 ISSN: 2250-1193
- [10] Atli, G., Ariyurek, S. Y., Kanak, E. G., & Canli, M. (2015). Alterations in the serum biomarkers belonging to different metabolic systems of fish (*Oreochromis niloticus*) after Cd and Pb exposures. *Environmental Toxicology and Pharmacology*, 40(2), 508-515.
 - [11] Nriagu, J. O., & Pacyna, J. M. (1988). Quantitative assessment of worldwide contamination of air, water and soils by trace metals. *nature*, 333(6169), 134-139.
 - [12] Nicolaidou, A., & Nott, J. A. (1989). Heavy metal pollution induced by a ferro-nickel smelting plant in Greece. *Science of the total environment*, 84, 113-117.
 - [13] Naz, S., & Javed, M. (2015). Determination of 96-hr LC50 and lethal responses of *Wallago attu* to water-borne chromium.
 - [14] Kotaš, J., & Stasicka, Z. J. E. P. (2000). Chromium occurrence in the environment and methods of its speciation. *Environmental pollution*, 107(3), 263-283.
 - [15] Espenson, J. H. (1970). Oxidation of transition metal complexes by chromium (VI). *Accounts of Chemical Research*, 3(10), 347-353.
 - [16] Apte, A. D., Tare, V., & Bose, P. (2006). Extent of oxidation of Cr (III) to Cr (VI) under various conditions pertaining to natural environment. *Journal of Hazardous Materials*, 128(2-3), 164-174.
 - [17] Ao, M., Sun, S., Deng, T., Zhang, F., Liu, T., Tang, Y., ... & Qiu, R. (2022). Natural source of Cr (VI) in soil: The anoxic oxidation of Cr (III) by Mn oxides. *Journal of Hazardous Materials*, 433, 128805
 - [18] Jiang, B., Gong, Y., Gao, J., Sun, T., Liu, Y., Oturan, N., & Oturan, M. A. (2019). The reduction of Cr (VI) to Cr (III) mediated by environmentally relevant carboxylic acids: state-of-the-art and perspectives. *Journal of Hazardous Materials*, 365, 205-226.
 - [19] Kerur, S. S., Bandekar, S., Hanagadakar, M. S., Nandi, S. S., Ratnamala, G. M., & Hegde, P. G. (2021). Removal of hexavalent Chromium-Industry treated water and Wastewater: A review. *Materials Today: Proceedings*, 42, 1112-1121.
 - [20] Cooper, C. M. (1993). Biological effects of agriculturally derived surface water pollutants on aquatic systems-a review. *Journal of environmental quality*, 22(3), 402-408.
 - [21] Camargo, J. A., & Alonso, Á. (2006). Ecological and toxicological effects of inorganic nitrogen pollution in aquatic ecosystems: a global assessment. *Environment international*, 32(6), 831-849.
 - [22] Javed, M., & Usmani, N. (2019). An overview of the adverse effects of heavy metal contamination on fish health. *Proceedings of the National Academy of Sciences, India Section B: Biological Sciences*, 89, 389-403.
 - [23] Jayaswal, K., Sahu, V., & Gurjar, B. R. (2018). Water pollution, human health and remediation. *Water remediation*. 11-27,
 - [24] Zahoor, I., & Mushtaq, A. (2023). Water pollution from agricultural activities: A critical global review. *Int. J. Chem. Biochem. Sci*, 23(1), 164-176.
 - [25] Naeem, S., Ashraf, M., Babar, M. E., Zahoor, S., & Ali, S. (2021). The effects of some heavy metals on some fish species. *Environmental Science and Pollution Research*, 28, 25566-25578.
 - [26] Uysal, K., Köse, E., Bülbül, M., Dönmez, M., Erdoğan, Y., Koyun, M., ... & Özmal, F. (2009). The comparison of heavy metal accumulation ratios of some fish species in Enne Dame Lake (Kütahya/Turkey). *Environmental monitoring and assessment*, 157, 355-362.
 - [27] Firat, Ö., & Kargin, F. (2010). Individual and combined effects of heavy metals on serum biochemistry of Nile tilapia *Oreochromis niloticus*. *Archives of environmental contamination and toxicology*, 58, 151-157.
 - [28] Mallesh, B., Pandey, P. K., Kumar, K., Vennila, A., & Kumar, S. (2015). Bioconcentration of hexavalent chromium in *Cirrhinus mrigala* (Ham 1822): effect on haematological parameters. *Journal of Earth Science*, 5(1), 59-67.
 - [29] Naz, S., & Javed, M. (2015). Determination of 96-hr LC50 and lethal responses of *Wallago attu* to water-borne chromium
 - [30] Pelgrom, S. M. G. J., Lock, R. A. C., Balm, P. H. M., & Bonga, S. W. (1995). Integrated physiological response of tilapia, *Oreochromis mossambicus*, to sublethal copper exposure. *Aquatic Toxicology*, 32(4), 303-320.
 - [31] Tayyab, S., Tanveer, A., & Muhammad, A. Acute toxicity studies of hexavalent Cr in the common carp, *Cyprinus carpio*. *Pakistan J. Zool.* 2005; 20: 209-214.