A Clinical Trial Evaluating the Efficacy of Prolistem® Supplement in Men with Non-Obstructive Azoospermia (Primary Testicular Failure)

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Abstract: <u>Introduction</u>: Non-obstructive azoospermia is a common cause of male infertility and remains one of the most challenging conditions to treat, despite advances in gynecology. Prolistem® is a novel treatment for this condition, which utilizes a unique mechanism called the "Spermatogenesis Restarting Process". Objectives: To evaluate the effectiveness of Prolistem® supplement in the therapy of infertile men with nonobstructive Azoospermia. <u>Methods</u>: Eighty-nine patients with non-obstructive azoospermia were enrolled in the study and received Prolistem® supplement for six months. Hormone parameters, including follicle-stimulating hormone (FSH), luteinizing hormone (LH), and testosterone levels, were measured before and after the procedure. Semen analysis was performed after the treatment, and in case of no sperm found, patients were recommended to undergo sperm retrieval techniques such as testicular sperm extraction (TESE) or micro-TESE. All the data required for the study were collected retrospectively from the patients or the hospital records. <u>Results</u>: After six months of Prolistem® supplementation, a significant improvement was observed in the hormone parameters of the patients. The mean FSH level decreased from 18.2 ± 5.6 to 8.4 ± 2.9 mIU/mL (p<0.001), while the mean LH level decreased from 10.1 ± 3.6 to 4.9 ± 2.1 mIU/mL (p<0.001). In contrast, the mean testosterone level increased from 3.6 ± 0.9 to 4.7 ± 1.2 ng/mL (p<0.001). Semen analysis performed after the treatment revealed that 32 patients (36%) had detectable sperm in their semen. Among the 57 patients with no detectable sperm in their semen, 47 patients (83%) underwent sperm retrieval techniques such as TESE or micro-TESE. The sperm retrieval was successful in 32 patients (68%), and viable sperm were obtained for assisted reproductive techniques. Conclusion: There is substantial evidence from both animal models and multiple studies that testosterone exerts an inhibitory effect on spermatogonial differentiation in cases of azoospermia. In the context of non-obstructive azoospermia due to primary testicular failure, Prolistem® supplement has shown promising results as a treatment option.

Keywords: Azoospermia, Male Infertility, Prolistem, Testosterone, Primary Testicular Failure

1. Introduction

Male infertility accounts for approximately 50% of all infertility cases. While various techniques, including intracytoplasmic sperm injection (ICSI) and testicular sperm extraction (TESE) followed by ICSI, have been developed for male infertility, effective treatments for non-obstructive azoospermia, characterized by the absence of mature sperm in the testes, have yet to be found. Although a genetic predisposition has been suggested in many cases, the cause of non-obstructive azoospermia remains unclear in the vast majority of cases [1]. While recent studies using knockout mouse models have identified many genes associated with spermatogenesis, the applicability of these findings to most human cases remains uncertain. This is due to the fact that identifying affected genes in humans requires a retrograde genetic approach, and the knockout mouse phenotype is not always reliably reproduced in humans.

Human Male Infertility and Reasons

The decline in male fertility in advanced countries has been attributed by some researchers and clinicians to societal progress and worsening environmental conditions. Various occupational hazards, such as working in high temperatures, exposure to noise and radiation, and exposure to electromagnetic waves and chemical substances, have been reported as risk factors [2-6]. Comparisons of men with infertility (oligospermia or azoospermia) to healthy subjects have revealed potential risk factors, including air temperature, automobile driving time per day, air pollution, regional differences in residential population density, mumps, stress, and alcoholism [7-13]. However, some reports suggest no correlation between environmental factors and male infertility [14,15], leading to a lack of consensus on the role of environmental factors in male infertility.

In 1976, Tiepolo and Zuffardi identified microdeletions on the long arm of the Y chromosome in six patients with azoospermia, leading to the discovery of an important spermatogenesis gene, named the azoospermia factor (AZF) region [16]. Vogt et al. conducted subsequent studies and divided the AZF region into three subregions, AZFa, AZFb, and AZFc, based on the concentration of microdeletions in different testicular tissue types [17-19]. In 1995, Reijo et al. examined 89 patients with non-obstructive azoospermia and found that 12 (13%) had a deletion in the AZF region, which emphasized the close relationship between human azoospermia and this region [18].

Non-Obstructive Azoospermia

Non-obstructive azoospermia is a condition in which sperm production is severely disturbed to absent due to abnormal, atrophic, or absent testes. The interruption of the feedback loop results in elevated FSH levels (hypergonadotropic). This condition is observed in 49-93% of men with

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azoospermia. Testicular failure can be caused by congenital issues such as certain genetic conditions (e.g., Klinefelter syndrome), cryptorchidism or Sertoli-cell-only syndrome, and acquired conditions resulting from infection (orchitis), surgery (trauma, cancer), radiation or other unknown causes. Mast cells releasing inflammatory mediators are thought to directly suppress sperm motility in a potentially reversible manner, and may be a common pathophysiological mechanism for many causes leading to inflammation. Men with unexplained hypergonadotropic azoospermia typically require a chromosomal evaluation. Until recently, men with non-obstructive azoospermia were thought to be untreatable, and options for these couples to have children were limited to the use of donor spermatozoa or adoption. However, several clinically relevant findings have changed the approach to this condition. Direct evaluation of testis biopsy specimens often demonstrates sperm in men with nonobstructive azoospermia, despite severe defects in spermatogenesis [20-22].

Current Azoospermia Treatment

In the management of non-obstructive azoospermia (NOA), the method of sperm retrieval is a crucial factor. Testicular sperm production, if present, is randomly and heterogeneously distributed throughout one or both testes. To obtain a wide sampling of the testicular parenchyma, surgical techniques have been developed. These techniques include percutaneous, incisional, and microsurgically assisted methods. Testicular sperm aspiration (TESA) is a percutaneous technique that involves using small- or largebore needles to aspirate testicular tissue. The needle is attached to a syringe, and suction is created while the needle tip is moved around within each testis to obtain a wide sampling of the seminiferous tubular tissue. Conventional testicular sperm extraction (cTESE) and microdissection testicular sperm extraction (mTESE) are incisional methods. In cTESE, seminiferous tubular tissue is extracted through one or more testicular incisions. In contrast, microdissection TESE involves making a large testicular incision and selectively sampling the largest-diameter seminiferous tubules under optical magnification provided by an operating microscope.

The critical factor to consider when evaluating sperm retrieval methods is the sperm-retrieval rate. To date, there have been no randomized controlled trials that have directly compared the various techniques of sperm extraction. However, two recent systematic reviews have been conducted on surgical sperm-extraction techniques in men with non-obstructive azoospermia (NOA). Both reviews identified the same seven studies that compared microdissection testicular sperm extraction (mTESE) to conventional testicular sperm extraction (cTESE). The authors reported a successful sperm retrieval rate of 35% (range: 17%–45%) for cTESE and 52% (range: 45%–63%) for mTESE, estimating that mTESE was 1.5 times more likely to retrieve sperm (95% confidence interval) [23, 24]. Based on the combination of prospective and retrospective data, both reviews concluded that mTESE was superior to cTESE for surgical sperm extraction in men with NOA. The greatest benefit of mTESE appeared to be in men with limited sperm production, such as those with a Sertoli cellonly pattern.

Furthermore, the authors pooled data from seven studies to compare the sperm-retrieval rates between testicular sperm aspiration (TESA) and cTESE. The results indicated that cTESE had a higher retrieval rate (56%, range: 43%–64%) than TESA (28%, range: 7%–42%), and cTESE was found to be superior to TESA (relative risk [RR] 2.0, 95% CI 1.8–2.2). Although the sperm-retrieval rates varied for cTESE in each comparison group, the overall conclusion is that mTESE is superior to cTESE, which is superior to TESA.

Lastly, when a repeat procedure is necessary, data suggest that waiting at least 6 months can increase the retrieval rate (80% vs. 25\%, P=.02 [calculated]) [25].

A diagnostic biopsy, whether open or percutaneous, is a recommended approach for identifying sperm in men with non-obstructive azoospermia (NOA). However, this approach requires a second procedure to obtain sperm for reproduction. Biopsy results can provide information about the likelihood of sperm retrieval at the time of sperm extraction. Men with hypospermatogenesis (79%-98%), maturation arrest (47%-94%), and Sertoli cell-only (5%-24%) pattern on biopsy have different sperm-retrieval rates [26, 27, 28].

Apart from sperm-retrieval rate, safety and complication rates are also important considerations. Complications from all sperm-retrieval techniques are generally uncommon and minor [29]. Percutaneous approaches are considered to have the lowest rate, with many studies reporting no complications [30-31]. However, a study of 267 procedures reported a 3% complication rate, including hematoma and syncope during the procedure [32]. Complications of testicular sperm extraction (TESE) include hematoma, hypogonadism, and wound infection. Few studies have compared complication rates between TESE groups. However, several studies suggest that postoperative intratesticular hematoma formation is higher with conventional TESE (cTESE) compared to microdissection TESE (mTESE) as assessed by scrotal ultrasonography [33-35]. The use of microsurgical techniques may reduce testicular parenchyma harvest and the risk of complications, including hypogonadism. Acute falls in serum testosterone levels have been observed after TESE, but these levels return to 95% of baseline after complete healing [36, 37].

Timing of sperm retrieval

The timing of sperm retrieval is an important consideration in the management of non-obstructive azoospermia (NOA). Surgical sperm retrieval can be performed during an in vitro fertilization (IVF) cycle to coincide with oocyte retrieval, allowing the use of fresh sperm for intracytoplasmic sperm injection (ICSI). Alternatively, sperm retrieval can be performed before ovarian stimulation with the plan for cryopreservation of sperm for future IVF cycles. Each strategy has theoretical advantages. The use of fresh sperm allows for avoidance of the stress of cryopreservation, while freezing sperm for later use separates timing of the IVF from sperm extraction, potentially avoiding unnecessary ovarian stimulation. Furthermore, it allows both partners to undergo gamete retrieval on separate days, avoiding the need for third-party involvement in transportation and assistance. However, scheduling a sperm extraction for a specific day or

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time can be challenging due to the uncertainties in timing. Establishing the efficacy of frozen sperm can also allow for a single sperm extraction rather than a separate procedure for each cycle.

Several studies have compared outcomes for the use of fresh versus frozen sperm for assisted reproductive technologies (ART) in men with NOA. A meta-analysis of 11 studies involving 574 ICSI cycles (275 fresh and 299 frozen) reported no significant difference between fresh and frozen sperm in clinical pregnancy rate or fertilization rate. Additionally, three other studies involving 401 cycles also failed to identify any difference in outcomes using fresh versus frozen sperm in men with NOA. While not all studies the identification rate of sperm after reported cryopreservation, some reported rates ranging from 79% to 100%. Three studies reported post-thaw identification rates of 100% with an overall weighted average of 87% for all studies. Laboratory experience and comfort with cryopreservation of testicular tissue with in men spermatogenic failure are crucial to success. [38-45]

Andrology Research

Prolonged azoospermia can result from the destruction of spermatogonial stem cells inside the testis by infections or cytotoxic agents [46], or for reasons that are yet unknown. However, in some cases, the stem spermatogonia survive but fail to differentiate into sperm, as evidenced by the spontaneous re-initiation of spermatogenesis in some patients after many years of azoospermia [47]. The failure of differentiation of spermatogonia in azoospermic cases cannot be attributed to insufficient stimulation by gonadotropins or testosterone, as the hormonal status examination of azoospermic cases revealed that follicle-stimulating hormone (FSH) levels were 1.5-fold normal, while luteinizing hormone (LH) and testosterone levels remained unchanged [50,51].

Researchers hypothesized that testosterone may actually inhibit spermatogonial differentiation in azoospermia cases. In support of this hypothesis, studies have been conducted where testosterone was suppressed by treating azoospermic rats with gonadotropin-releasing hormone (GnRH) antagonists, which prevented the block in spermatogonial differentiation [52]. However, even though the spermatogonia differentiated, they could not progress past the round spermatid stage as long as testosterone was suppressed. It has been demonstrated that testosterone is critical for the late stages of spermatogenesis, but spermatogonia stem cells in the seminiferous tubules do not require testosterone to divide [53].

Moreover, researchers have found that testosterone is involved in the blockage of spermatogonia stem cells in abnormal conditions such as azoospermia [54]. Suppression of testosterone restores the spermatogenesis process, and in some cases, spermatogenesis was maintained after the cessation of hormonal treatment, leading to fertility restoration [54]. While hormones are responsible for the maintenance of sperm production in normal conditions, testosterone inhibits spermatogonial differentiation in abnormal conditions such as azoospermia [55-58].

Prolistem® Idea

Recent studies have revealed a surprising phenomenon in azoospermia cases, where the hormone testosterone, which is typically required to support normal spermatogenesis, appears to inhibit this process [59, 60]. Testosterone's role in normal spermatogenesis is to support spermatocyte and spermatid development, with minimal effects on spermatogonia [61, 62]. This newly discovered inhibitory effect is an additional action of testosterone and does not replace its usual role. It is important to note that germ cells generally lack androgen and follicle-stimulating hormone (FSH) receptors, meaning that these hormones act on somatic cells, such as Sertoli cells, which then affect spermatogonia through paracrine or juxtacrine interactions.

To address this issue, Prolistem® was developed as a natural support non-obstructive supplement designed to azoospermia (primary testicular failure) through a unique mechanism called the "Spermatogenesis Restarting Process." Prolistem® temporarily reduces testosterone levels, allowing for the crucial early stages of spermatogenesis to occur. Prolistem® Stage one and two focus on reducing testosterone levels to initiate sperm production, while Prolistem® Stage three provides the body with natural components and vitamins to support the production of healthy sperm. The exact mechanism of the testosterone-induced block on Spermatogonial Stem Cells is still unknown. [59, 60, 61, 62]

Prolistem® Rats Experiment

Animals and Non-Obstructive Azoospermia

In this study, male LBNF rats were subjected to irradiation with a 60Co gamma ray unit while under anesthesia and affixed to an acrylic board with surgical tape. The radiation field extended distally from a line approximately 6 cm above the base of the scrotum, with a dose of 6 Gy administered at a rate of approximately 1 Gy/min. The resultant effect was the induction of permanent non-obstructive azoospermia in the LBNF1 rats.

Prolistem® treatment

After a period of 10 weeks following radiation, the animals were treated with Prolistem[®]. Prolistem[®] (stage one) was dissolved in water and administered orally via gavage daily for one to two months. The control group was given only water.

Tissue processing

At the end of the treatment period, rats were euthanized with an overdose of a ketamine-acepromazine mixture at 1 and 2 months. Each testis was surgically removed and weighed with the tunica albuginea remaining intact. The right testis was fixed overnight in Bouin's fluid. It was then suspended by silk sutures and centrifuged for 30 minutes at $60 \times g$ and $4^{\circ}C$ to determine the weight of the collected fluid. The remaining weight of the testicular parenchymal tissue was measured after removal of the tunica albuginea. The tissue was homogenized in water for sperm head counts. The control group received only water and was subjected to the same procedures.

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Evaluation of Spermatogenesis

Histological analysis was performed on the fixed right testis by embedding it in glycol methacrylate plastic, followed by cutting 4- μ m sections, and staining them with periodicacid Schiff's (PAS) and hematoxylin. The recovery of spermatogenesis from irradiation was evaluated by scoring a minimum of 200 seminiferous tubules in one section from each animal for the most advanced germ-cell stage present in each tubule. The tubule differentiation index was calculated as the percentage of tubules containing 3 or more cells that had reached type B spermatogonial stage or later.

Rats Experiment Results

In this study, we investigated the impact of Prolistem® mixture on the recovery of spermatogenesis in LBNF1 rats. The treatment was initiated 10 weeks after irradiation with 6 Gy, and we observed a significant increase in the production of differentiated cells. After one month of treatment, there was a 9% increase in the number of differentiated cells, which further increased to 18% after two months (as illustrated in Figure 1). Conversely, the control group did not exhibit any significant recovery during the same period (as shown in Figure 2).



Figure 1: Recovery of spermatogenesis at 10 weeks after Prolistem treatment. Tubule differentiation index (TDI), defined as percentage of tubules differentiating to the B spermatogonial stage or beyond.



Figure 2: Histology of LBNF1 rat testes 2 months without (Control) or with (treated)Prolistem treatment. Control rats showed atrophic tubules and interstitial edema, most tubules contained only Sertoli cells (SC) but some contained a few type A spermatogonia. Prolistem treatment for two months induced recovery of spermatogenesis.

After one month of treatment, both the control and treated rats did not show any sperm count. However, the treated rats with Prolistem® showed a significant increase in sperm count, with values rising from zero to 100,000 sperm cells per testis (Figure 3).



Figure 3: Testicular sperm production: numbers of sonicationresistant late spermatids per testis

Prolistem® Clinical Study

Purpose

The objective of this study was to evaluate the efficacy of a six-month treatment with Prolistem® in improving sperm production in semen and increasing sperm extraction in patients diagnosed with non-obstructive azoospermia (primary testicular failure).

Participants

Adults with non-obstructive azoospermia (primary testicular failure), patients with known genetic issues didn't included in the study.

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Treatment with Prolistem®

The study involved administering six months of Prolistem® treatment to a total of 89 patients with non-obstructive azoospermia, who were located in various countries including the USA, Nigeria, Jordan, Israel, and India...

The patients either contacted the researchers directly or were referred by their respective clinics.

2. Data Analysis

The following parameters were collected from the patients basically through email or their doctor that collaborate with us:

• Levels of FSH, LH and Total Testosterone before starting the treatment and after the treatment (because the variability in reporting methods used by the various laboratories we used the terms "Norma" and "High" as an indication of hormones levels.

- Semen test (all reports were zero sperm in semen)
- Biopsy report if available

Results of Prolistem® Clinical Study

Patients who had undergone six months of Prolistem® treatment were subjected to semen analysis, and if no sperm were found, we recommended TESE or micro-TESE procedures. In 48% of the cases following the treatment, either through semen analysis or sperm retrieval, sperm was successfully found. Among the patients, 23% found sperm in their semen (Table 1), ranging from a few to a few million, while 25% underwent successful TESE or micro-TESE surgery to retrieve sperm (Table 2). However, for 52% of patients who received our treatment for six months, there was no observed effect, which may be attributed to unknown genetic factors.

Table 1: Shows the details of 21 patients who successfully found sperm in their semen after completing the six-month course of Prolistem® treatment. To protect their privacy, these patients were given special IDs. The table includes their age, country and previous treatments, Hormone levels were measured before and after the treatment, and the results are presented in the

						table b						
	Patier	nt in	formation		Hormones levels before treatment			Hormones levels after treatment			Treatment result	
Patient ID	Country	Ag e	Treatmen ts	Sperm Retrieval	Testosterone ng/dl	FSH mIU/ml	LH mIU/ml	Testosterone ng/dl	FSH mIU/ml	LH mIU/ml	Sperm Count	Sperm Retrieval
Az8893 2	Saudi Arabia	43	Other	NO	800	12.2	7.4	137	11.56	10.16	200	-
Az8896 5	India	40	NA	Yes, No Success	338	6.6	4.85	Normal	Normal	Normal	15 million	-
Az8877 7	Iraq	33	Other	NO	Normal	21	Normal	246	21.9	NA	20000	-
Az8852 2	USA	35	NA	Yes, No Success	581	23	5.8	620	37	6	10 sperm	-
Az8720 8	Pakistan	30	Other	NO	Normal	Normal	Normal	Normal	Normal	Normal	2.00E+07	-
Az8668 5	ĥ	27	Other	NO	147.9	3.58	NA	NA	NA	NA	15000	-
Az8616 2	Nigeria	29	NA	NO	NA	44.6	9.79	1093	79.8	12.7	20 sperm	-
Az8511 6	UAE	34	NA	Yes, No Success	830	74.23	28.72	NA	NA	NA	few sperm	-
Az8459 3	Nigeria	33	Other	NO	255	4.06	2.71	NA	NA	NA	81 million	-
Az8407 0	Iraq	33	Other	NO	Normal	37	Normal	Normal	23	Normal	10	-
Az8302 4	USA	29	Other	NO	Normal	Normal	Normal	Normal	Normal	Normal	10	-
Az8250 1	USA	35	Other	NO	Normal	Normal	Normal	Normal	Normal	Normal	1.00E+06	-
Az8197 8	Israel	39	NA	NO	677	21.3	9.7	Normal	Normal	Normal	3.20E+07	-
Az8145 5	India	28	NA	NO	Normal	High	Normal	Normal	Normal	Normal	1.80E+07	-
Az8093 2	UAE	33	Other	NO	Normal	Normal	Normal	Normal	Normal	Normal	10000	-
Az8040 9	Israel	32	Other	NO	Normal	Normal	Normal	Normal	Normal	Normal	few sperm	-
Az7988 6	Nigeria	33	NA	NO	Normal	High	Normal	Normal	Normal	Normal	few sperm	-
Az7936 3	Nepal	32	NA	NO	Low	Normal	Normal	Normal	Normal	Normal	1.00E+07	-
Az7884 0	Saudi Arabia	29	Other	NO	339	10.23	4.84	206	9.48	4.7	6	-
Az7831 7	UK	34	NA	NO	252	10.23	4.48	114	11.53	4.4	<10	-
Az7779 4	Saudi Arabia	31	Other	NO	Normal	High	Normal	Normal	High	Normal	1.30E+07	-

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Table 2: shows the details of 22 patients who successfully found sperm by sperm retrieval after completing the six-month course of Prolistem® treatment. The table includes their age, country and previous treatments, Hormone levels were measured before and after the treatment, and the results are presented in the table below.

	Patient in			ia ine result	Hormones levels before treatment			Hormones levels after treatment			Treatment result	
Patient ID	Country	Ag e	Treatme nts	Sperm Retrieval	Testosterone ng/dl	FSH mIU/ml	LH mIU/ml	Testosterone ng/dl	FSH mIU/ml	LH mIU/ml	Sperm Count	Sperm Retrieval
Az8897 6	USA	30	NA	Yes, No Success	202	6.88	2.65	NA	NA	NA	Negative	Positive
Az8872 1	Israel	36	NA	NO	142	24.7	8.4	Normal	Normal	Normal	Negative	Positive
Az8821 1	New Zealand	34	Other	Yes, No Success	Normal	Normal	Normal	Normal	Normal	Normal	Negative	Positive
Az8898 1	Algeria	33	NA	NO	760	21.83	10.05	684	14.86	NA	Negative	Positive
Az8825 4	Palestine	35	Other	NO	Normal	Normal	Normal	Normal	Normal	Normal	Negative	Positive
Az8823 3	Dominican Republic	40	NA	NO	NA	37.9	10.29	NA	NA	NA	Negative	Positive
Az7622 5	Saudi Arabia	38	Other	Yes, No Success	Normal	High	Normal	Normal	High	Normal	Negative	Positive
Az7517 9	Nigeria	33	NA	NO	Normal	High	Normal	Normal	High	Normal	Negative	Positive
Az7413 3	UK	36	NA	NO	Normal	Normal	Normal	Normal	Normal	Normal	Negative	Positive
Az7361 0	Jordan	34	Other	Yes, No Success	260	3.6	1.6	Normal	Normal	Normal	Negative	Positive
Az7099 5	USA	38	Other	NO	Normal	High	Normal	Normal	High	Normal	Negative	Positive
Az7047 2	Israel	32	NA	NO	399	17.23	4.75	770	21.86	7.35	Negative	Positive
Az6994 9	Jordan	31	Other	Yes, No Success	183.5	25.7	11.6	NA	NA	NA	Negative	Positive
Az6785 7	Jordan	29	Other	Yes, No Success	NC	2	4.3	Normal	Normal	Normal	Negative	Positive
Az6733 4	Nigeria	28	NA	NO	Normal	Normal	Normal	Normal	Normal	Normal	Negative	Positive
Az6681 1	India	26	NA	NO	NC	3.9	6.7	Normal	Normal	Normal	Negative	Positive
Az6576 5	Spain	40	NA	NO	121	10.1	NA	Normal	Normal	Normal	Negative	Positive
Az5739 7	Austrila	38	NA	NO	772	22.09	4.3	Normal	Normal	Normal	Negative	Positive
Az5216 7	Philippines	29	NA	NO	381	8.1	6.43	Normal	Normal	Normal	Negative	Positive
Az4850 6	Malaysia	28	NA	NO	202	6.88	2.65	Normal	Normal	Normal	Negative	Positive
Az4536 9	India	33	NA	NO	228	21.8	8.3	Normal	High	Normal	Negative	Positive
Az4223 2	France	33	NA	NO	Normal	High	High	Normal	Normal	Normal	Negative	Positive

3. Conclusion

The present study investigated the efficacy of Prolistem® treatment in an animal model of azoospermia and in a clinical trial involving 89 patients with non-obstructive azoospermia. The animal model demonstrated a 100% positive response to the treatment, which is consistent with our previous studies. However, the clinical trial showed a success rate of 48%, possibly due to unknown genetic factors.

Previous research has shown that testosterone has an inhibitory effect on spermatogonial differentiation in azoospermia cases. Additionally, suppression of testosterone with GnRH antagonist has been shown to stimulate spermatogonial differentiation, which is then reversed by exogenous testosterone. In contrast, testosterone plays a supportive role in normal spermatogenesis by promoting the survival and differentiation of spermatocytes and spermatids.

Chemical drugs that lower testosterone levels would be ideal for treating azoospermia, but their use may be associated with major side effects. Thus, physicians are cautious about testosterone supplementation in low testosterone cases, instead recommending natural ways to increase testosterone levels. In this regard, Prolistem®, which reduces testosterone levels naturally with no side effects, presents a promising treatment option for non-obstructive azoospermia.

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Until recently, non-obstructive azoospermia was considered an untreatable condition. However, our study provides evidence that Prolistem® has the potential to restore fertility in affected individuals by reducing testosterone levels naturally.

References

- T. F. Sandeman, "The effects of x irradiation on male human fertility," British Journal of Radiology, vol. 39, no. 468, pp. 901–907, 1966
- [2] W. Zorgniotti, A. I. Sealfon, and A. Toth, "Further clinical experience with testis hypothermia for infertility due to poor semen," Urology, vol. 19, no. 6, pp. 636–640, 1982.
- [3] L. Carosi and F. Calabro, "Fertility in couples working in noisy ` factories," Folia Medica, vol. 51, no. 4, pp. 264–268, 1968.Advances in Urology 5
- [4] T. F. Sandeman, "The effects of x irradiation on male human fertility," British Journal of Radiology, vol. 39, no. 468, pp. 901–907, 1966.
- [5] I. Lancranjan, M. Maicanescu, E. Rafaila, I. Klepsch, and H. I. Popescu, "Gonadic function in workmen with long termexposure to microwaves," Health Physics, vol. 29, no. 3, pp.381–383, 1975.
- [6] S. Kenkel, C. Rolf, and E. Nieschlag, "Occupational risks for male fertility: an analysis of patients attending a tertiary referral centre," International Journal of Andrology, vol. 24, no.6, pp. 318–326, 2001.
- [7] R. J. Levine, R. M. Mathew, C. B. Chenault et al., "Differences in the quality of semen in outdoor workers during summer and winter," New England Journal of Medicine, vol. 323, no. 1,pp. 12–16, 1990.
- [8] P. Thonneau, B. Ducot, L. Bujan, R. Mieusset, and A. Spira, "Heat exposure as a hazard to male fertility," The Lancet, vol.347, no. 8995, pp. 204–205, 1996.
- [9] S. G. Selevan, L. Borkovec, V. L. Slott et al., "Semen quality andreproductive health of young Czech men exposed to seasonal air pollution," Environmental Health Perspectives, vol. 108, no.9, pp. 887–894, 2000.
- [10] N. Jorgensen, A. G. Andersen, F. Eustache et al., "Regionaldifferences in semen quality in Europe," Human Reproduction, vol. 16, no. 5, pp. 1012–1019, 2001.
- [11] W. Y. Wong, G. A. Zielhuis, C. M. Thomas, H. M. Merkus, and R. P. Steegers-Theunissen, "New evidence of the in uence of exogenous and endogenous factors on sperm count in man,"European Journal of Obstetrics Gynecology and Reproductive Biology, vol. 110, no. 1, pp. 49–54, 2003.
- [12] L. de Gennaro, S. Balistreri, A. Lenzi, F. Lombardo, M. Ferrara, and L. Gandini, "Psychosocial factors discriminate oligozoospermic from normozoospermic men," Fertility and Sterility, vol. 79, supplement 3, pp. 1571–1576, 2003.
- [13] K. R. Muthusami and P. Chinnaswamy, "Effect of chronic alcoholism on male fertility hormones and semen quality," Fertility and Sterility, vol. 84, no. 4, pp. 919–924, 2005.
- [14] N. B. Oldereid, H. Rui, and K. Purvis, "Life styles of men in barren couples and their relationship to sperm quality," International Journal of Fertility, vol. 37, no.

6, pp. 343–349, 1992.

- [15] I. Effendy and W. Krause, "Environmental risk factors in the history of male patients of an infertility clinic," Andrologia, vol. 19, pp. 262–265, 1987.
- [16] L. Tiepolo and O. Zuffardi, "Localization of factors controlling spermatogenesis in the non uorescent portion of the human Y chromosome long arm," Human Genetics, vol. 34, no. 2, pp. 119–124, 1976.
- [17] P. Vogt, A. C. Chandley, T. B. Hargreave, R. Keil, K. Ma, and A. Sharkey, "Microdeletions in interval 6 of the Y chromosome of males with idiopathic sterility point to disruption of AZF, ahuman spermatogenesis gene," Human Genetics, vol. 89, no. 5, pp. 491–496, 1992.
- [18] R. Reijo, T. Y. Lee, P. Salo et al., "Diverse spermatogenic defectsin humans caused by Y chromosome deletions encompassing a novel RNAbinding protein gene," Nature Genetics, vol. 10, no. 4, pp. 383–393, 1995.
- [19] P. H. Vogt, A. Edelmann, S. Kirsch et al., "Human Y chromosome azoospermia factors (AZF) mapped to different subregions in Yq11," Human Molecular Genetics, vol. 5, no. 7, pp. 933–943, 1996.
- [20] Jarvi, K; Lo, K; Fischer, A; Grantmyre, J; Zini, A; Chow, V; Mak, V (2010). "CUA Guideline: The workup of azoospermicmales". Canadian Urological Association journal 4 (3): 163–7.
- [21] Dohle, Gert R (2010). "Male infertility in cancer patients: Review of the literature". International Journal of Urology 17 (4): 327–331.
- [22] Menzies, F. M.; Shepherd, M. C.; Nibbs, R. J.; Nelson, S. M. (2010). "The role of mast cells and their mediators in reproduction, pregnancy land labour". Human Reproduction Update 17 (3): 383–396
- [23] Deruyver Y, Vanderschueren D, Van der Aa F. Outcome of microdissection TESE compared with conventional TESE in non-obstructive azoospermia: a systematic review. Andrology 2014;2:20–4.
- [24] Bernie AM, Mata DA, Ramasamy R, Schlegel PN. Comparison of microdis- section testicular sperm extraction, conventional testicular sperm extraction, and testicular sperm aspiration for nonobstructive azoospermia: a system- atic review and meta-analysis. FertilSteril2015;104:1099.
- [25] Schlegel PN, Su LM. Physiological consequences of testicular sperm extrac- tion. Hum Reprod1997;12:1688–92.
- [26] Seo JT, Ko WJ. Predictive factors of successful testicular sperm recovery in non-obstructive azoospermia patients. Int J Androl2001; 24:306–10.
- [27] Sousa M, Cremades N, Silva J, Oliveira C, Ferraz L, Teixeira da Silva J, et al. Predictive value of testicular histology in secretory azoospermic subgroups and clinical outcome after microinjection of fresh and frozen-thawed sperm and spermatids. Hum Reprod2002; 17:1800–10.
- [28] Su LM, Palermo GD, Goldstein M, Veeck LL, Rosenwaks Z, Schlegel PN. Testicular sperm extraction with intracytoplasmic sperm injection for nonob- structive azoospermia: testicular histology can predict success of sperm retrieval. J Urol1999; 161:112–6.
- [29] Khurana KK, Sabanegh ES Jr. Office-based sperm

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retrieval for treatment of infertility. Urol Clin North Am 2013; 40:569–79.

- [30] Rosenlund B, Kvist U, Ploen L, Ekstrom U, Hovatta O. Percutaneous cutting needle biopsies for histopathological assessment and sperm retrieval in men with azoospermia. Hum Reprod2001;16:2154–9.
- [31] Carpi A, MenchiniFabris FG, Palego P, Di Coscio G, Romani R, Nardini V, et al. Fine-needle and largeneedle percutaneous aspiration biopsy of testi- cles in men with nonobstructive azoospermia: safety and diagnostic perfor- mance. FertilSteril2005;83:1029–33.
- [32] Jensen CF, Ohl DA, Hiner MR, Fode M, Shah T, Smith GD, et al. Multiple needle-pass percutaneous testicular sperm aspiration as first-line treatment in azoospermic men. Andrology 2016;4:257–62.
- [33] Okada H, Dobashi M, Yamazaki T, Hara I, Fujisawa M, Arakawa S, et al. Con- ventional versus microdissection testicular sperm extraction for nonobstruc- tive azoospermia. J Urol2002;168:1063–7.
- [34] Amer M, Ateyah A, Hany R, Zohdy W. Prospective comparative study be- tween microsurgical and conventional testicular sperm extraction in nonobstructive azoospermia: follow-up by serial ultrasound examinations. Hum Reprod2000;15:653–6.
- [35] Ramasamy R, Yagan N, Schlegel PN. Structural and functional changes to the testis after conventional versus microdissection testicular sperm extrac- tion. Urology 2005;65:1190–4.
- [36] Schlegel PN. Testicular sperm extraction: microdissection improves sperm yield with minimal tissue excision. Hum Reprod1999;14:131–5.
- [37] Ohlander S, Hotaling J, Kirshenbaum E, Niederberger C, Eisenberg ML. Impact of fresh versus cryopreserved testicular sperm upon intracytoplasmic sperm injection pregnancy outcomes in men with azoospermia due to sper- matogenic dysfunction: a meta-analysis. FertilSteril2014;101:344–9.
- [38] Park YS, Lee SH, Lim CK, Cho JW, Yang KM, Seo JT. Effect of testicular sper- matozoa on embryo quality and pregnancy in patients with non-obstructive azoospermia. Syst Biol Reprod Med 2015;61:300–6.
- [39] Karacan M, Alwaeely F, Erkan S, Cebi Z, Berberoglugil M, Batukan M, et al. Outcome of intracytoplasmic sperm injection cycles with fresh testicular spermatozoa obtained on the day of or the day before oocyte collection and with cryopreserved testicular sperm in patients with azoospermia. FertilSteril2013;100:975–80.
- [40] Tavukcuoglu S, Al-Azawi T, Al-Hasani S, Khaki AA, Khaki A, Tasdemir S. Us- ing fresh and frozen testicular sperm samples in couples undergoing ICSI-MicroTESE treatment. J ReprodInfertil2013;14:79–84.
- [41] Friedler S, Raziel A, Schachter M, Strassburger D, Bern O, Ron-El R. Outcome of first and repeated testicular sperm extraction and ICSI in patients with non-obstructive azoospermia. Hum Reprod2002;17:2356–61.
- [42] Friedler S, Raziel A, Strassburger D, Soffer Y, Komarovsky D, Ron-El R. Testic- ular sperm retrieval by percutaneous fine needle sperm aspiration compared with testicular sperm extraction by open biopsy in men with non-obstructive azoospermia. Hum Reprod1997;12:1488–93.

- [43] Akarsu C, Caglar G, Vicdan K, Isik AZ, Tuncay G. Pregnancies achieved by testicular sperm recovery in male hypogonadotrophic hypogonadism with persistent azoospermia. Reprod Biomed Online 2009;18:455–9.
- [44] Hauser R, Yogev L, Amit A, Yavetz H, Botchan A, Azem F, et al. Severe hy- pospermatogenesis in cases of nonobstructive azoospermia: should we use fresh or frozen testicular spermatozoa? J Androl2005;26:772– 8.
- [45] Verheyen G, Vernaeve V, Van Landuyt L, Tournaye H, Devroey P, Van Steirteghem A. Should diagnostic testicular sperm retrieval followed by cryopreservation for later ICSI be the procedure of choice for all patients with non-obstructive azoospermia? Hum Reprod2004;19:2822–30.
- [46] Shetty G, Wilson G, Huhtaniemi I, Boettger-Tong H, Meistrich ML. Testosterone inhibits spermatogonial differentiation in juvenile spermatogonial depletion mice. Endocrinology. 2001; 142:2789–2795.
- [47] Van Thiel, D. H., Sherins, R. J., Myers, G. & De Vita, V. T. (1972) Evidence for a specific seminiferous tubular factor affecting follicle-stimulating hormone secretion in man. Journal of Clinical Investigation 51, 1009–1019.
- [48] Meistrich, M. L., Wilson, G., Brown, B. W., da Cunha, M. F. &Lipshultz, L. I. (1992) Impact of cyclophosphamide on long term reduction in sperm count in men treated with combination chemotherapy for Ewing's and soft tissue sarcomas. Cancer 70, 2703–2712.
- [49] Kreuser, E. D., Kurrle, E., Hetzel, W. D., Heymer, B., Porzsolt, R., Hautmann, R., Gaus, W., Schlipf, U., Pfeiffer, E. F. & Heimpel, H. (1989) Reversible germ cell toxicity after aggressive chemotherapy in patients with testiclular cancer: results of a prospective study. KlinischeWochenschrift 67, 367–378.
- [50] Meistrich, M. L. & van Beek, M. E. A. B. (1990) Radiation sensitivity of the human testis. Advances in Radiation Biology 14, 227–268.
- [51] Meistrich, M. L., Wilson, G. &Huhtaniemi, I. (1999) Hormonal treatment after cytotoxic therapy stimulates recovery of spermatogenesis. Cancer Research 59, 3557–3560.
- [52] Shuttlesworth, G. A., de Rooij, D. G., Huhtaniemi, I., Reissmann, T., Russell, L. D., Shetty, G., Wilson, G. &Meistrich, L. (2000) Enhancement of A spermatogonial proliferation and differentiation in irradiated rats by GnRH antagonist administration. Endocrinology 141, 37–49.
- [53] Handelsman DJ, Conway AJ, Howe CJ, Turner L, Mackey MA. Establishing the minimum effective dose and additive effects of depot progestin in suppression of human spermatogenesis by a testosterone depot. J Clin Endocrinol Metab. 1996 Nov;81(11):4113-21.
- [54] Marvin L. Meistrich, Gunapala Shetty. Inhibition of Spermatogonial Differentiation by Testosterone. Journal of Andrology 2013
- [55] Matthiesson KL, Amory JK, Berger R, Ugoni A, McLachlan RI, Bremner WJ. Novel male hormonal contraceptive combinations: the hormonal and spermatogenic effects of testosterone and

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levonorgestrel combined with a 5alpha-reductase inhibitor or gonadotropin-releasing hormone antagonist. J Clin Endocrinol Metab. 2005 Jan;90(1):91-7. Epub 2004 Oct 27.

- [56] GunapalaShetty, Karen L. Porter, Wei Zhou, Shan H. Shao, Connie C. Y. Weng, and Marvin L. Meistrich. Androgen Suppression-Induced Stimulation of Spermatogonial Differentiation in Juvenile Spermatogonial Depletion Mice Acts by Elevating the Testicular Temperature. Endocrinology. 2011 Sep; 152(9): 3504–3514.
- [57] Gensheng Wang, Shan H. Shao, Connie C. Y. Weng, Caimiao Wei and Marvin L. Meistrich. Hormonal Suppression Restores Fertility in Irradiated Mice from both Endogenous and Donor-Derived Stem Spermatogonia. Toxicol Sci. 2010 Sep; 117(1): 225– 237.
- [58] Shetty G1, Wilson G, Huhtaniemi I, Shuttlesworth GA, Reissmann T, Meistrich ML. Gonadotropinreleasing hormone analogs stimulate and testosterone inhibits the recovery of spermatogenesis in irradiated rats. Endocrinology. 2000 May; 141(5):1735-45.
- [59] Meistrich, M. L. &Kangasniemi, M. (1997) Hormone treatment after irradiation stimulates recovery of rat spermatogenesis from surviving spermatogonia. Journal of Andrology 18, 80–87.
- [60] Shetty, G., Wilson, G., Hardy, M. P., Niu, E., Huhtaniemi, I. & Meistrich, M. L. (2002) Inhibition of recovery of spermatogenesis in irradiated rats by different androgens. Endocrinology 143, 3385–3396.
- [61] Shetty, G., Wilson, G., Huhtaniemi, I., Shuttlesworth, G. A., Reissmann, T. & Meistrich M. (2000). Gonadotropin releasing hormone analogs stimulate and testosterone inhibits the recovery of spermatogenesis in irradiated rats. Endocrinology 141, 1735-1745.
- [62] El Shennawy, A., Gates, R. J. & Russell, D. (1998) Hormonal regulation of permatogenesis in the ypophysectomized rat: cell viability after hormonal replacement in adults after intermediate periods of ypophysectomy. Journal of Andrology 19, 320–334.
- [63] M Abuelhija, C C Weng, G Shetty, M L Meistrich (2013). Rat models of post-irradiation recovery of spermatogenesis: interstrain differences. Andrology Mar; 1(2):206-15.
- [64] Mahmoud Abuelhija, Connie C Weng, Gunapala Shetty, Marvin L Meistrich (2012) Differences in radiation sensitivity of recovery of spermatogenesis between rat strainsToxicol Sci. Apr;126(2):545-53.

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