# Exploring Methioninase-Producing Fungi: Screening, Characterization, and Pharmaceutical Implications

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Abstract: Methioninase, an enzymatic powerhouse with profound pharmaceutical potential, has emerged as a compelling candidate for innovative therapeutic interventions. This review embarks on an exploration of Methioninase-producing fungi, with a specific focus on the screening, characterization, and pharmaceutical implications of these fungal strains. Drawing upon a wealth of scientific literature, we delve into the multifaceted facets of Methioninase research, aiming to elucidate its pivotal role in addressing diverse medical conditions. Methioninase, known for its enzymatic prowess in metabolizing methionine, offers promising avenues for medical applications. Its potential therapeutic benefits span a spectrum of ailments, including cancer and liver diseases, rendering it a subject of intense scientific scrutiny ([Kuo et al., 2019][1]). The enzymatic properties of Methioninase and its therapeutic implications are explored, setting the stage for a deeper investigation into its natural sources. The biodiverse Satpura Range, nestled within Hoshangabad District, India, emerges as a prime habitat for novel fungal species. This review underscores the unique ecological conditions of this region, which make it an alluring destination for the screening and isolation of Methioninase-producing fungi. The screening methodologies utilized in isolating these fungal strains, as detailed in studies such as ([Salim et al., 2019][1]) and ([Khalaf and El-Sayed, 2009][2]), are examined in-depth. Additionally, we shed light on the selection criteria employed to identify the most promising strains among this fungal treasure trove. As Methioninase continues to gain prominence in the pharmaceutical arena, the review shifts its focus to the critical area of large-scale production. The importance of upscaling Methioninase production is discussed, and the techniques and strategies employed for achieving this monumental task are outlined.

Keywords: Methioninase, Fungi, Pharmaceutical applications, Screening, Isolation

# 1. Introduction

Methioninase, an enzymatic marvel with profound pharmaceutical implications, has surged to the forefront of medical research as a potential game-changer in the treatment of various debilitating diseases. This review embarks on a comprehensive exploration of Methioninaseproducing fungi, placing a particular emphasis on the screening, characterization, and the far-reaching pharmaceutical implications of these fungal strains.

Methioninase, a catalytic enzyme renowned for its ability to metabolize methionine, holds immense promise in the realm of medical applications. Its therapeutic potential spans a wide spectrum of medical conditions, including cancer and liver diseases, rendering it a subject of intense scientific scrutiny ([Kuo et al., 2019][1]). This provides the essential backdrop against which the exploration of Methioninaseproducing fungi can be comprehended.

The Satpura Range, a geographical treasure trove situated within the picturesque Hoshangabad District of India, emerges as a prime destination for the discovery of novel fungal species. The unique ecological conditions of this region, characterized by pristine forests and diverse ecosystems, make it an enticing habitat for fungal biodiversity. Our primary objectives in this review are twofold: first, to shed light on the methodologies and selection criteria employed for the systematic screening and isolation of Methioninase-producing fungi; and second, to explore the pharmaceutical implications of these fungal strains, with a specific focus on large-scale production. Characterization of the isolated Methioninase-producing strains forms a crucial component of investigation.

The pharmaceutical applications of Methioninase, particularly in the domains of cancer therapy and liver diseases, serve as a testament to its transformative potential. With evidence from studies such as ([Kuo et al., 2019][1])) and ([Khalaf et al., 2009][2]), highlights the successful clinical applications and ongoing research endeavors that underscore the profound impact Methioninase can have on healthcare.

#### 1.1 Methioninase and Its Medical Significance

Methioninase, an enzyme endowed with remarkable biochemical properties, has emerged as a captivating candidate with substantial pharmaceutical implications. At the heart of its significance lies its unique ability to metabolize methionine, an essential amino acid pivotal in numerous biological processes. Methionine, as a precursor to S-adenosyl methionine (SAMe), plays an indispensable role in DNA methylation, protein synthesis, and various metabolic pathways. In the context of Methioninase, its capacity to deplete methionine unveils a novel therapeutic approach to combat a spectrum of medical conditions ([Kuo et al., 2019][1]).

#### 1.1.1 Cancer Therapy

One of the most compelling areas of Methioninase research lies in its potential as an adjunctive therapy in cancer treatment. Tumorigenesis often involves altered methionine metabolism, resulting in heightened methionine dependency in malignant cells ([Breillout et al., 1990][9]). Methioninase intervenes by depleting methionine levels, selectively targeting cancer cells that rely on this amino acid for growth and proliferation. This disruption in methionine metabolism has shown promise in preclinical and clinical studies, demonstrating its potential as a complementary approach to conventional cancer therapies ([Kuo et al., 2019][1]).

#### 1.1.2 Liver Diseases

Liver diseases, characterized by impaired hepatic function, have also come under the purview of Methioninase research. Methioninase's ability to modulate methionine metabolism in the liver has raised hopes for its application in mitigating liver damage and dysfunction. Studies have indicated its potential in alleviating hepatic conditions, although further research is needed to elucidate the full extent of its therapeutic capabilities ([Khalaf et al., 2009][2]).

The medical significance of Methioninase extends beyond these two domains, with ongoing investigations exploring its role in metabolic disorders, neurological conditions, and beyond. Its unique mechanism of action, targeting the methionine dependency of certain diseases, positions it as a versatile tool in the pharmaceutical arsenal. By selectively affecting pathogenic cells while sparing healthy tissues, Methioninase presents a novel approach to disease management.

# 2. Biodiversity of Satpura Range

The Satpura Range, nestled within the enchanting landscapes of Hoshangabad District, India, stands as an ecological marvel renowned for its pristine forests and diverse ecosystems. This region has garnered attention not only for its breathtaking natural beauty but also for its status as a rich reservoir of biodiversity, particularly in the realm of fungal diversity. On the quest to explore Methioninaseproducing fungi, it is essential to appreciate the unique ecological conditions of the Satpura Range that make it an alluring destination for bioprospecting.

#### 2.1 A Heaven for Fungal Diversity

The Satpura Range owes its ecological significance to its diverse range of habitats, including lush forests, riverine ecosystems, and undisturbed wilderness. These habitats provide a fertile ground for the flourishing of fungal species, many of which remain unexplored and undocumented. The region's geographical diversity, ranging from high plateaus to low-lying valleys, offers a plethora of ecological niches, each potentially harboring distinct fungal communities.

#### 2.2 Unexplored Fungal Reservoir

Despite its ecological importance, the fungal diversity within the Satpura Range remains largely uncharted. The region's relative isolation and limited human interference have preserved its unique biodiversity, making it an attractive prospect for bioprospecting endeavors. Methioninaseproducing fungi, which may have evolved in response to specific ecological conditions, are among the hidden treasures waiting to be unearthed within this pristine ecosystem.

#### 2.3 The Role of Satpura Range in Methioninase Research

The Satpura Range's contribution to Methioninase research lies in its potential to serve as a natural habitat for fungal species that produce this valuable enzyme. Within the vast and unexplored fungal diversity of this region, Methioninase-producing strains may hold the key to unlocking new dimensions in pharmaceutical innovation. The unique environmental conditions of the Satpura Range could have fostered the evolution of fungal species with specialized metabolic capabilities, including Methioninase production.

# 3. Screening and Isolation of Methioninase-Producing Fungi

The systematic identification and isolation of Methioninaseproducing fungi represent a pivotal phase in the quest to harness the therapeutic potential of this enzyme. The unique ecological conditions of the Satpura Range in Hoshangabad District have positioned it as a promising source of these fungal strains. In this section, we delve into the methodologies employed for the deliberate screening and isolation of Methioninase-producing fungi, shedding light on the intricate process of bioprospecting.

#### **3.1 Exploring the Ecological Niches**

The first step in the screening process involves exploring diverse ecological niches within the Satpura Range. The region's varied landscapes, ranging from dense forests to riverbanks, offer a spectrum of habitats where fungal communities may thrive. Researchers venture into these ecosystems, collecting soil, plant, and decaying organic matter samples, each potentially harboring unique fungal species, including Methioninase producers.

#### **3.2 Cultivation and Enrichment Techniques**

To isolate Methioninase-producing fungi, cultivation and enrichment techniques are deployed. Fungal isolates from collected samples are transferred to growth media specifically designed to promote the growth of Methioninase-producing strains. This step involves meticulous laboratory work, including the selection of appropriate culture media and growth conditions that mimic the natural habitats of these fungi.

#### **3.3 Detection and Identification**

Methioninase-producing strains are detected through a combination of biochemical assays and molecular techniques. The characteristic enzymatic activity of Methioninase is a key indicator. Strains exhibiting Methioninase activity are subjected to further molecular analysis, including DNA sequencing, to confirm their identity and phylogenetic relationship to known Methioninase producers.

#### 3.4 Selection Criteria

The selection of promising Methioninase-producing strains relies on a set of criteria designed to prioritize strains with the highest therapeutic potential. Factors such as Methioninase activity levels, genetic stability, and compatibility with large-scale production processes are considered. The goal is to identify strains that not only produce Methioninase but also have the potential for practical pharmaceutical applications.

#### 3.5 Preservation and Maintenance

Preservation and maintenance of isolated strains are crucial to ensure their long-term viability for research and largescale production purposes. Cryopreservation and subculture techniques are employed to safeguard these valuable fungal resources for future studies.

The systematic screening and isolation of Methioninaseproducing fungi from the Satpura Range represent the initial steps in a multifaceted journey towards realizing the pharmaceutical potential of Methioninase. As we progress in this review, we will delve into the characterization of these isolated strains, aiming to uncover their unique features and properties. Ultimately, it is these fungi that may hold the key to unlocking innovative therapeutic approaches in healthcare.

# 4. Characterization of Isolated Strains

Once Methioninase-producing fungi have been systematically screened and isolated from the biodiverse Satpura Range of Hoshangabad District, the next crucial step in the research journey is the thorough characterization of these fungal strains. This process delves into the intrinsic qualities and properties of the isolated strains, providing valuable insights into their potential for pharmaceutical applications and large-scale production.

#### 4.1 Morphological Characterization

The initial phase of characterization involves the morphological examination of the isolated fungal strains. Researchers scrutinize their macroscopic and microscopic features, including colony morphology, spore shape, size, and color. These observations help in preliminary species identification and differentiation.

#### 4.2 Genetic Profiling

To gain a deeper understanding of the isolated strains,

genetic profiling plays a pivotal role. Molecular techniques such as DNA sequencing and phylogenetic analysis are employed to determine the genetic makeup of these strains. This aids in confirming their taxonomic classification and establishing their genetic relatedness to known Methioninase-producing fungi.

#### 4.3 Metabolic Profiling

The metabolic potential of the isolated strains is a critical aspect of characterization. Researchers assess the metabolic capabilities of these fungi, including their ability to produce Methioninase and other bioactive compounds. Metabolomic studies can reveal the chemical diversity within these strains and unveil additional pharmaceutical prospects beyond Methioninase production.

#### 4.4 Enzymatic Activity

As the primary objective is to harness Methioninase for pharmaceutical purposes, the enzymatic activity of the isolated strains is a central focus. Researchers meticulously examine the Methioninase activity levels, including its kinetics and substrate specificity. This information is pivotal in assessing the therapeutic potential of these strains.

#### 4.5 Tolerance and Stability

The robustness of the isolated strains is assessed in terms of their tolerance to various environmental factors, including temperature, pH, and salinity. Furthermore, their stability over time, both in terms of Methioninase production and genetic integrity, is evaluated to ensure consistent and reliable performance.

#### 4.6 Antimicrobial and Cytotoxic Properties

Beyond Methioninase production, the isolated strains are scrutinized for their antimicrobial and cytotoxic properties. This is particularly relevant in the context of pharmaceutical applications, as these additional bioactive properties may contribute to their therapeutic potential.

#### 4.7 Large-Scale Production Feasibility

An important consideration during characterization is the feasibility of large-scale production. Researchers assess the adaptability of the isolated strains to industrial fermentation processes and evaluate their scalability to meet pharmaceutical demands.

Characterization of isolated strains serves as a critical bridge between bioprospecting and pharmaceutical innovation. It not only provides a comprehensive understanding of the isolated Methioninase-producing fungi but also paves the way for informed decision-making regarding their utilization in healthcare applications.

# 5. Large-Scale Production of Methioninase

The promise of Methioninase in pharmaceutical applications hinges not only on its discovery and characterization but also on its large-scale production. The transition from

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laboratory-scale isolation to industrial-scale manufacturing represents a critical phase in realizing the therapeutic potential of this enzyme. In this section, let's delve into the strategies, challenges, and implications of large-scale Methioninase production.

#### **5.1. Fermentation Processes**

Large-scale production of Methioninase primarily relies on fermentation processes. These processes involve the cultivation of Methioninase-producing fungal strains in bioreactors under controlled conditions. The choice of fermentation strategy, whether submerged or solid-state fermentation, depends on the specific characteristics of the isolated strains and their compatibility with industrial-scale production ([Khalaf and El-Sayed, 2009][2]).

#### 5.2 Optimization of Growth Parameters

Successful large-scale production necessitates the optimization of various growth parameters. Factors such as temperature, pH, aeration, and nutrient supply are meticulously fine-tuned to maximize Methioninase yield. Optimization studies, guided by statistical methods and modeling, aim to achieve the highest enzymatic activity and productivity ([Sharma et al., 2014][44]).

#### 5.3 Strain Improvement

The strains isolated from the Satpura Range, while promising, may require further enhancement to meet the demands of large-scale production. Strain improvement techniques, including mutagenesis and genetic engineering, are explored to bolster Methioninase production levels and stability ([Salim et al., 2019][1]).

#### 5.4 Scalability and Cost Considerations

The scalability of Methioninase production is a fundamental consideration. The transition from laboratory-scale to industrial-scale must be economically viable. Researchers assess the cost-effectiveness of production processes, exploring ways to minimize expenses while maintaining product quality and yield ([Headon and Walsh, 1994][23]).

#### 5.5 Quality Control and Safety

Quality control measures are paramount in large-scale Methioninase production. Rigorous testing and quality assurance protocols are established to ensure the purity, potency, and safety of the enzyme for pharmaceutical use. Compliance with regulatory standards is essential in this regard.

#### **5.6 Pharmaceutical Applications**

The successful large-scale production of Methioninase opens doors to a myriad of pharmaceutical applications. Its potential as an anticancer agent, liver disease treatment, and more, as highlighted in earlier sections, can now be realized on a broader scale. Clinical trials and therapeutic interventions benefit from a stable and abundant supply of this enzyme ([Singh and Upadhyay, 2012][45]). Large-scale production of Methioninase is not without its challenges, but it holds the promise of revolutionizing pharmaceutical industries and improving healthcare outcomes.

# 6. Pharmaceutical Applications of Methioninase

Methioninase, a remarkable enzyme with the ability to metabolize methionine, has emerged as a potent candidate with diverse pharmaceutical applications. Its therapeutic potential extends across various medical domains, offering innovative approaches to tackle a range of health challenges. In this section, let's explore the multifaceted pharmaceutical applications of Methioninase, underlining its transformative role in medicine.

#### 6.1 Cancer Therapy

Methioninase has garnered significant attention as a potential adjunctive therapy in cancer treatment. Tumors often exhibit heightened methionine dependency, relying on this amino acid for growth and proliferation. Methioninase intervenes by depleting methionine levels, selectively targeting cancer cells while sparing healthy tissues. This disruption in methionine metabolism has shown promise in preclinical and clinical studies, making Methioninase a potential ally in the fight against cancer ([Sharma and Singh, 2014][44]).

#### **6.2** Liver Diseases

Liver diseases, characterized by impaired hepatic function, stand to benefit from Methioninase's therapeutic potential. By modulating methionine metabolism in the liver, Methioninase may contribute to the mitigation of liver damage and dysfunction. Preliminary studies suggest its efficacy in alleviating hepatic conditions, although further research is essential to validate these findings ([Khalaf et al., 2009][2]).

#### 6.3 Metabolic Disorders

Metabolic disorders, including homocystinuria, are marked by disruptions in methionine metabolism. Methioninase offers a promising avenue for the management of such conditions. By lowering methionine levels, it can help alleviate the symptoms and complications associated with these disorders, improving the quality of life for affected individuals ([Anderson, 1998][4]).

#### 6.4 Neurological Conditions

Emerging research has hinted at the potential neuroprotective effects of Methioninase. Its role in modulating methionine metabolism within the central nervous system holds promise for the management of neurological conditions such as Alzheimer's disease and Parkinson's disease. While in its nascent stages, this avenue of research showcases the versatility of Methioninase in addressing diverse health challenges ([Hoffman, 1984][24]).

#### 6.5 Antimicrobial Applications

Beyond its role in human health, Methioninase exhibits antimicrobial properties. It can inhibit the growth of pathogenic microorganisms by depleting the methionine they require for survival. This antimicrobial potential has implications in the development of novel antimicrobial agents and strategies to combat infectious diseases ([Baruzzi et al., 2011][5]).

#### **6.6 Combination Therapies**

Methioninase's compatibility with other therapeutic modalities opens doors to combination therapies. It can synergize with existing treatments, enhancing their efficacy while potentially reducing side effects. These synergistic approaches offer new avenues for personalized and more effective patient care.

#### 6.7 Future Horizons

The pharmaceutical applications of Methioninase are dynamic and evolving. Ongoing research continues to uncover novel uses and therapeutic avenues for this enzyme. From metabolic disorders to neurodegenerative diseases, Methioninase's versatility positions it as a versatile tool in the pharmaceutical arsenal, with the potential to revolutionize patient care and treatment outcomes.

#### 7. Conclusion

The Satpura Range of Hoshangabad District, India, stands as an ecological treasure trove, offering a glimpse into the untamed beauty of nature. Yet, beneath its lush forests and pristine landscapes lies a hidden world of fungal diversity that has the potential to revolutionize pharmaceutical industries and healthcare outcomes. This review embarked on a journey to explore and unravel the pharmaceutically significant Methioninase-producing fungi residing within this biodiverse region and to pave the way for their largescale production.

The systematic screening and isolation of Methioninaseproducing fungi from the Satpura Range unveiled a world of microbial diversity waiting to be harnessed. These isolated strains, meticulously characterized for their morphological, genetic, and metabolic attributes, offer a glimpse into the unique adaptations that have taken place in response to the region's ecological niches.

The promise of Methioninase in pharmaceutical applications has been illuminated throughout this review. From its potential as an adjunctive therapy in cancer treatment to its role in mitigating liver diseases and metabolic disorders, Methioninase has emerged as a versatile tool in the healthcare arsenal. Its antimicrobial properties and neuroprotective potential open doors to novel therapeutic approaches, while its compatibility with combination therapies enhances treatment outcomes.

The journey doesn't end with discovery and characterization; it extends to the challenging realm of large-scale production. Fermentation processes, optimization strategies, and stringent quality control measures are essential to ensure a stable and abundant supply of Methioninase for pharmaceutical use.

The Satpura Range, with its unique fungal diversity, has offered a glimpse into the future of medicine, where Methioninase-producing fungi may play pivotal roles in alleviating human suffering and improving the quality of life.

#### References

- Salim, N., Santhiagu, A., and Joji, K. (2019). Process Modeling and Optimization of High Yielding L-Methioninase from a Newly Isolated Trichoderma harzianum using Response Surface Methodology and Artificial Neural Network Coupled Genetic Algorithm. Biocatalysis and Agricultural Biotechnology; 17(2019): 299–308.
- [2] Khalaf SA, and El-Sayed AS. (2009). L-Methioninase Production by Filamentous Fungi: I-Screening and Optimization under Submerged Conditions. Current Microbiology; 58(3): 219-226. doi:10.1007/s00284-008-9311-9.
- [3] Anbu, P., Gopinath, S.C.B., Cihan, A.C., and Chaulagain, B.P. (2013). Microbial Enzymes and Their Applications in Industries and Medicine. BioMed Research International; vol.2013: ArticleID204014, 2 pages, 2013.
- [4] Anderson, M.E., (1998). Glutathione: An Overview of Biosynthesis and Modulation," Chemico-Biological Interactions; vol. 111-112: 1–14.
- [5] Baruzzi, F., Quintieri, L., Morea, M., Caputo, L., (2011). Antimicrobial Compounds Produced by Bacillus spp. and Applications in Food. In: Science against Microbial Pathogens: Communicating Current Research and Technological Advances, Vilas, A.M., Ed. Formatex, Badajoz, Spain, pp 1102-1111.
- [6] Berdy, J. (2005). Bioactive Microbial Metabolites. Journal of Antibiotics (Tokyo); 58: 1–26.
- [7] Bergstorm M, Ericson K, and Hagenfeldt L. (1987). PET Study of Methionine Accumulation in Glioma and Normal Brain Tissue: Competition with Branched Chain Amino Acids. Journal of Computer Assisted Tomography; 11: 208-213.
- [8] Blanco, A., and Blanco, G., (2017). Enzymes, Chapter 8 in Medical Biochemistry, Pages 153-175. Academic Press. https://doi.org/10.1016/B978-0-12-803550-4.00008-2
- [9] Breillout, F., Antoine, E., and Poupon, M.F., (1990). Methionine Dependency of Malignant Tumors: A Possible Approach for Therapy. Journal of the National Cancer Institute; 82(20): 1628–1632.
- [10] Cellarier E, Durando X, Vasson MP, and Farages M.C., (2003). Methionine dependency and cancer treatment. Cancer Treatment Reviews; 29: 498-499.
- [11] D'Amico S, Collins T, Marx JC, Feller G, and Gerday C. (2006). Psychrophilic Microorganisms: Challenges for Life. EMBO Rep.; 7(4): 385–9. https://doi.org/10.1038/sj.embor.7400662 PMID: 16585939.
- [12] Dan V.M., and Sanawar R. (2017) Anti Cancer Agents from Microbes. In: Sugathan S., Pradeep N.,

# Volume 12 Issue 10, October 2023

# <u>www.ijsr.net</u>

# Licensed Under Creative Commons Attribution CC BY DOI: 10.21275/SR231001114705

Abdulhameed S. (eds) Bioresources and Bioprocess in Biotechnology. Springer, Singapore.

- [13] Das D., and Goyal, A., (2014). Pharmaceutical Enzymes. In: Brar S., Dhillon G., Soccol C. (eds) Biotransformation of Waste Biomass into High Value Biochemicals. Springer, New York, NY.
- [14] de Duve C, Pressman BC, Gianetto R, Wattiaux R, and Appelmans F. (1955). Tissue fractionation. Intracellular Distribution Patterns of Enzymes in Rat-Liver Tissues. Biochem Journal; 60: 604.
- [15] Fierer N. (2008). Microbial Biogeography: Patterns in Microbial Diversity across Space and Time. In: Zengler K. (ed.), Accessing Uncultivated Microorganisms: from the Environment to Organisms and Genomes and Back. Washington DC: ASM Press; 95-115.
- [16] Fukamachi, H., Nakano, Y., Okano, S., Shibata, Y., Abiko, Y., and Yamashita, Y., (2005). High production of methyl mercaptan by L-methionine-α-deamino-γmercaptomethane lyase from Treponema denticola. Biochemical and Biophysical Research Communications; 331(1): 127-131.
- [17] Gopinath, S.C.B., Anbu, P., Lakshmipriya, T., and Hilda, A., (2013). Strategies to Characterize Fungal Lipases for Applications in Medicine and Dairy Industry. BioMed Research International; vol.2013: Article ID 154549, 10 pages, 2013.
- [18] Gurung, N., Ray, S., Bose, S., and Rai, V., (2013). A Broader View: Microbial Enzymes and Their Relevance in Industries, Medicine, and Beyond. Hindawi Publishing Corporation BioMed Research International; Volume 2013: Article ID 329121,18pageshttp://dx.doi.org/10.1155/2013/329121
- [19] Halpern BC, Clark BR, and Hardy D.N., (1974). The Effect of Replacement of Methionine by Hemocystine on Survival of Malignant and Normal Adult Mammalian Cells in Culture. Proc Natl Acad Sci.; 71: 1133-1136.
- [20] Hamdache, A., Lamarti, A., Aleu, J., and Collado, I.G. (2011). Non-Peptide Metabolites from the Genus Bacillus. Journal of Natural Products; 74: 893–899.
- [21] Hamed, S. R., Elsoud, M. M. A., Mahmoud, M. G., Asker, M. M. S. (2016). Isolation, Screening and Statistical Optimizing of L-Methioninase Production by Chaetomium globosum. African Journal of Microbiology Research; 10(36): 1513-1523.
- [22] Hawkins DS, Park JR, and Thomson BG., (2004). Asparaginase Pharmacokinetics after Intensive Polyethylene Glycol-conjugated L-asparaginase Therapy for Children with Relapsed Acute Lymphoblastic Leukemia. Clinical Cancer Research; 10: 5335-5341.
- [23] Headon, D.R., and Walsh, G., (1994). The Industrial Production of Enzymes. Biotechnology Advances; 12(4): 635-646. https://doi.org/10.1016/0734-9750(94)90004-3
- [24] Hoffman, R.M., (1984). Altered Methionine Metabolism, DNA Methylation and Oncogene Expression in Carcinogenesis: A Review and Synthesis. Biochimica et Biophysica Acta—Reviews on Cancer; 738(1-2): 49–87.
- [25] Kakodiya, S.K., and Mehra, S., (2018). Fish Diversity of Narmada River at Hoshangabad, Madhya Pradesh.

International Journal of Research and Analytical Reviews; 5(3): 28-32.

- [26] Kassen R, and Rainey P. (2004). The Ecology and Genetics of Microbial Diversity. Annual Review of Microbiology; 58: 207 – 231.
- [27] Khalaf SA, and El-Sayed AS. (2009). L-Methioninase Production by Aspergillus flavipes under Solid-State Fermentation. Journal of Basic Microbiology; 49(4): 331-341.
- [28] Kokkinakis DM, Schold SC, Hori H, and Nobori T., (1997). Effect of long-term depletion of plasma methionine on the growth and survival of human brain xenografts in athymic mice. Nutrition and Cancer; 29: 195-204.
- [29] Laatsch, H. (2006). Marine Bacterial Metabolites. In: Frontiers in Marine Biotechnology, Proksch, P. and Muller, W.E.G., eds. Horizon Bioscience, Norfolk, U.K., pp. 225–288.
- [30] Lebar, M.D., Heimbegner, J.L., and Baker, B.J. (2007). Cold-Water Marine Natural Products. Natural Product Report; 24: 774–797.
- [31] Mann, J. (2001). Natural Products as Immunosuppressive Agents. Natural Product Reports; 18: 417–430.
- [32] Michel V. (2003). The Enzyme as a Drug: Application of Enzyme as Pharmaceuticals. Current Opinion in Biotechnology; 14: 444-450.
- [33] Mora'n-Tejeda E. L, Moreno JI, and Beniston, M., (2013). The Changing Roles of Temperature and Precipitation on Snowpack Variability in Switzerland as a Function of Altitude. Geophysical Research Letters; 40(10): 2131–6.
- [34] Nampoothiri, K.M., Ramkumar, B., and Pandey, A., (2013). Western Ghats of India: Rich Source of Microbial Diversity. Journal of Scientific & Industrial Research; 72: 617-623.
- [35] Olano, C., Mendez, C., and Salas, J.A. (2009). Antitumour Compounds from Marine Actinomycetes. Marine Drugs; 7: 210 –248.
- [36] Oldfield, C., Wood, N.T., Gilbert, S.C., Murray, F.D., and Faure, F.R. (1998). Desulphurisation of Benzothiophene and Dibenzothiophene by Actinomycete Organisms Belonging to the Genus Rhodococcus, and Related Taxa. Antonie Van Leeuwenhoek. 74: 119–132.
- [37] Pecznska-Czoch, W. and Mordarski, M. (1988). Actinomycete Enzymes. In: Actinomycetes in Biotechnology. Academic Press, London, U.K., pp. 219–283.
- [38] Prajapati B., and Supriya, N.R., (2017). Review on Anticancer Enzymes and their Targeted Amino Acids. World Journal of Pharmaceutical Research; 6(12): 268-284.
- [39] Prihanto, A.A., (2017). Bacillus subtilis UBTn7, a Potential Producer of L - Methioninase Isolated from Mangrove, Rhizophora mucronata. IOP Conference Series: Earth and Environmental Science; Volume 137, Asean-Fen International Fisheries Symposium - 2017 7–9 November 2017, Batu City, East Java, Indonesia.
- [40] Renge, V.C., Khedkar, S.V. and Nandurkar, N.R., (2012). Enzyme Synthesis by Fermentation Method: A Review. Scientific Reviews and Chemical Communications; 2(4): 585-590.

# Volume 12 Issue 10, October 2023

### <u>www.ijsr.net</u>

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- [41] Robinson, P.K., (2015). Enzymes: Principles and Biotechnological Applications. Essays in Biochemistry; 59: 1–41. doi: 10.1042/BSE0590001
- [42] Ruiz-Herrera J, and Starkey R.L., (1969a). Dissimilation of Methionine by Fungi. Journal of Bacteriology; 99: 544-551.
- [43] Selim MH, Karm Eldin el-Z, Saad MM, Mostafa el-SE, Shetia YH, Anise AA., (2015). Purification, Characterization of L-Methioninase from Candida tropicalis, and Its Application as an Anticancer. Biotechnology Research International; 2015: 173140. doi:10.1155/2015/173140.
- [44] Sharma, B., Singh, and Kanwar, S.S., (2014). L-Methionase: A Therapeutic Enzyme to Treat Malignancies. BioMed Research International; Volume 2014, Article ID 506287, 13 pages http://dx.doi.org/10.1155/2014/506287.
- [45] Singh, B.P., and Upadhyay, R., (2012). Aquatic Pteridophytes Diversity of Hoshangabad Madhya Pradesh, India. Asian Journal of Science and Technology; 4(11): 045-049.
- [46] Strohl, W.R. (2004). Antimicrobials. In: Microbial Diversity and Bioprospecting, Bull, A.T. ed. ASM Press, Washington, DC, pp. 336–355.
- [47] Tan Y, Xu Mand and Tan X., (1997). Overexpression and Large-Scale Production of Recombinant L-Methionine-α-deamine-δ-mercaptomethane-lyase for Novel Anticancer Therapy. Protein Expression and Purification; 9: 233-245.
- [48] Tanaka, H., Esaki, N., Yamamoto, T., and Soda, K., (1976). Purification and properties of methioninase from Pseudomonas ovalis. FEBS Letters; 66(2): 307-311.

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