# Activity of Silver Nanoparticle Loaded Sodium Alginate Beats for Removal of Pathogenic Bacteria from Water

#### Shweta Ashokrao Deshpande

Department of biotechnology, Govt. Institute of Science Aurangabad, 431004 Maharashtra, India dshweta4595[at[gmail.com

**Abstract:** Silver nanoparticles (AgNPs) show effective antimicrobial activity against wide range of disease demonstrated by Fusarium oxysporum fungus. I investigated the synthesis of silver nanoparticles through biological method. Then silver nanoparticle loaded with sodium alginate beads were tested for removal of pathogenic bacteria from water. Bacteria are naturally found in clinical and industrial settings in association with surfaces. E.coli, most commonly found dominant disease causing bacteria. Biologically synthesized AgNPs shows significantly reduce growth of E.coli. as a time of incubation increases. Sodium alginates beads are ecofriendly nanocomposites film.

Keywords: sodium alginates beads, silver nanoparticle, bacteria

## 1. Introduction

Nanotechnology is a interdisciplinary science dealing with smallest particle. (krukemeyer M.G. et al., 2015). Nanotechnology prominently involves use of nanoparticles. Nanoparticles, whose smallest functional organization is on nanometer scale or one billionth of meter (109). Nanotechnology has a massive range of applications. One of the functions of nanoparticles is to show antimicrobial activity against wide range of disease causing microbes. Due to their high specific surface area, AgNPs are more able to interact with membranes of bacterial cells. Several study reported that AgNPs can damage the cell membrane, leading to structural changes that makes bacteria more permeable. In this nanoparticle which is characterized by XRD, FTIR, U.V. Spectroscopy, TEM, Zeta Potential, used to test antimicrobial activity.

### 2. Materials and Methods

- 1) Potato Dextrose Agar and Broth
- 2) E. coli
- 3) F. oxysporum
- 4. Silver Nitrate, Sodium alginate 1% W/V at 25%, Calcium chloride reagent

#### Stock solution of silver Nitrate:

- Required: 0.1Mm silver nitrate solution.
- Molecular weight of silver nitrate: 169.87g
- 0.1M silver nitrate solution was prepared by adding 1.69g of silver nitrate in 100ml distilled water.
- 5) Sterile Distilled Water
- 6) Glassware's
- 7) Centrifuge, Incubator

#### 3. Procedure

#### Part A:

#### **Preparation of beads-**

50ml of 10% W/V aqueous solution of sodium alginate was introduce drop wise from a glass syringe with a size 22 needle into 100ml of an aqueous 10% calcium chloride solution being stirred at 400rpm. Calcium alginate beads were harvested by filtration, were washed and stored in (PBS) phosphate buffer saline.

#### **Collection of microorganisms:**

The microbes selected for the present study were *Escherichia coli*. The given stock of *E.coli* was used for all experiments.

#### **Preparation of inoculums:**

*E.coli* was recovered for testing by sub-culturing on fresh media. A loopful inoculum of bacterium was suspended in 5 ml of nutrient broth and incubated overnight at 37°C. These overnight culture were used as inoculums in all our experiments.

#### **Preparation of media:**

The growth media employed in the present study included LB agar and LB broth. The medium was adjusted to pH 7 and sterilized by autoclaving at 15 lbs pressure (121°C) for 15 minutes for sterilization.

#### Sub-culturing of microorganism:

The pure culture of was maintained on nutrient agar slant by frequent sub culturing. These cultures was stored at 4°C for further experiments

#### Part B:

#### **Column disinfection experiment:**

1) The prepared alginates beads were taken & repeatedly washed with sterile distilled water & were soaked in

www.ijsr.net

silver nanoparticles prepared from *F.oxysporum* into a sterile petriplate.

- 2) Soaked beads after an hour were packed in sterile chromatographic column.
- 3) The packed column was then placed U.V for an hour.
- 4) The sterile distilled water was passed through the column to remove any adsorbed nanoparticles on surface.
- 5) 0.1ml of sterile distilled water was passed though it and immediately collected in a sterile beaker, which was then immediately spread evenly on LB agar plate and kept for incubation at 37°C.
- 6) Dilutions of activated culture of *E.Coli* were prepared in sterile saline upto  $10^{-4}$  dilution.

- 7)  $10^{-4}$  dilution sample was then passed thorough nanoparticles coated column and collected at T = 1, 4, 8, 12 & 24 min
- 8) For each run three samples of effluent collected at T= 0min,
- 9) Immediately after being collected, *E.Coli* samples were homogenously spread on LB agar plates & placed into incubator at 37°C overnight to allow the growth of visible colony.
- 10) Also plated were the influent sample which was, *E.Coli* suspension without passing through any column.



# 4. Result

Number of colonies observed after exposure of E. coli to AgNPs loaded Sodium alginate

Sr. No.	Exposure time	No. of colony observed		
	(Minutes)	Set 1	Set 2	Set 3
1.	Control	00	00	00
2	1	286	281	290
3	4	231	240	228
4	8	190	196	194
5	12	180	184	177
6	24	94	99	91



Antibacterial activity results

# Volume 11 Issue 5, May 2022 www.ijsr.net

Licensed Under Creative Commons Attribution CC BY

# 5. Discussion

- Mycosynthesied AgNPs showed highest antibacterial activity at T= 24min.
- Thus AgNPs sodium alginate beads has remarkable potential for antibacterial water filter

# References

- [1] Abbasa, Q., Saleemb, M., Phulla, A., Rafiqa, M., Hassana, M., Leeb, K., and Seoa, S.(2017).Green synthesis of silver nanoparticles using bidens frondosa extract and their tyrosinase activity Iran. J Pharm. Res, 16 (2), 763-777.
- [2] AbidinAli, Z., Yahya, R., Sekaran, S., and Puteh, R. (2016). Green synthesis of silver nanoparticles using apple extract and its antibacterial properties, Adv. Mat. Sci. Engg.
- [3] Article ID 4102196, 6 pages.
- [4] Al-Thawadi, S., Rasool, A, and Youssef, K. (2017). Antimicrobial activity of biosynthesized silver nanoparticles against *E. coli and B. subtilis*. J Bioanal. Biomed. 9, 299-305.
- [5] Bhatia, S. (2016). Nanoparticles types, classification, characterization, fabrication methods and drug delivery applications. Nat. Pol.Drug Del. Sys.. DOI 10.1007/978-3319-41129-3
- [6] Brady, A., Laverty, G., Gilpin, D., Kearney, P.and Tunney, M. (2017). Antibiotic susceptibility of planktonic and biofilm grown staphylococci isolated from implant associated infections; Should MBEC and nature of biofilm formation replace MIC, J Med Microbiol 66, 461-469
- [7] Devika, R., Elumalai, S., Manikandan, E., and Eswaramoorthy, D.(2012). Biosynthesis of silver nanoparticles using the fungus *Pleurotusostreatus* & their antimicrobial activity, 1:557 doi;10.4172/scientificreports.557.
- [8] Doyle, W., (1992). Principles and applications of fourier transform infrared (FTIR) Process Analysis Technical Note AN–906 Rev. C.
- [9] Firdhouse, M., and Lalitha, P. (2015). Biosynthesis of silver nanoparticles and its applications J. Nanotechnol., Article ID 82.
- [10] Gudikandula, K., and Maringanti, S. (2016). Synthesis of silver nanoparticles by chemical and biological methods and their antimicrobial properties, J.Exptl. Nanosci., 11: 9, 714721. Guzmán, M., Dille, J., and Godet, S. (2008). Synthesis of silver nanoparticles by chemical reduction method and their antibacterial activity, Interntl. J Chem, Mol, Nuc, Mat Metall Engg 2:7.
- [11] HsuehY-H, LinK-S, KeW-J, , HsiehC-T, ChiangC-L and TzouD-Y.(2015). The antimicrobial properties of silver nanoparticles in *Bacillus subtilis are* mediated by released Ag+Ions. PLoSONE10(12):e0144306.doi:10.1371/journal.pone .0144306.
- [12] Hussain, J., Kumar, S., Hashmi, A., and Khan, Z. (2011). Silver nanoparticles: preparation, characterization, and Adv. Mat. Lett. 2:3, 188-194.
- [13] Hussain, J., Kumar, s., Hashmi, A. & Khan, Z. (2011). Silver nanoparticles: preparation,

characterization, and kinetics www.vbripress.com, www.amlett.com, DOI: 10.5185/amlett.2011.1206.

- [14] Ingle, A., Rai, M., Gade, A., and Bawaskar, M. (2009). *Fusarium solani*: a novel biological agent for the extracellular synthesis of silver nanoparticles, J Nanopart Res 11, 2079–2085.
- [15] Joanna, C., Marci, L., Ewa, K., and Grażyna, P. (2018). A nonspecific synergistic effect of biogenic silver nanoparticles and biosurfactant towards environmental bacteria and fungi. Ecotoxicology 27, 352–359.
- [16] Kohanski, M. Dwayer and Collins J.(2010). How antibiotics kill bacteria: from targets to networks, Nat Rev.. Microbiol 8(6), 423-435.
- [17] Kubyshkin, A., Chegodar, D., Katsev, A., Petrosyan, A., Krivorutchenko, Y., and Postnikova, O.(2016).Antimicrobial effects of silver nanoparticles stabilized in solution by sodium alginate. Biochem Mol Biol J.; 2(2): doi:10.21767/2471-8084.100022.
- [18] Kumar R., and Lal, S. (2014) Synthesis of organic nanoparticles and their applications in drug delivery and food nanotechnology: a review, J.Nanomat Mol Nanotechnol,.
- [19] Kumar, M., Bansal, K., Gondil, V., Sharma, S., Jain, D. V. S., Chhibber, S., Sharma, R., and Wangoo, N. (2018). Synthesis, characterization mechanistic studies and antimicrobial efficacy of biomolecule capped and ph modulated silver nanoparticles. J. Mol Liq 249, 1145–1150.
- [20] Mandal, S., Kumar, S., Krishnamoorthy, and B., Basu, S.(2010). Development and evaluation of calcium alginate beads prepared by sequential and simultaneous methods, Braz. J Pharma.Sci 46: 4.
- [21] Munita, J., and Arias, C. (2016). Mechanism of antibiotic resistance, Microbiol Spectr. doi: 10.1128/microbiolspec.VMBF-0016-2015.
- [22] Nguyen, T., Dang, V., Nguyen, N., Le, A., and Nguyen, Q. (2013). Synthesis of silver nanoparticles deposited on silica by  $\gamma$ -irradiation and preparation of PE/Ag nano compound master batches, Advances in natural sciences: Nanosci and Nanotechnol, 4 : 045004 4pp.
- [23] Nomcebo H.mthombemi, Onyangoa, M.S., Momba M.N.B. (2012). Break through analysis for water disinfection using silver nanoparticles resin beads in fixed bed column, J Hazard Mat 217218.
- [24] Nurani, S., Saha, C., Khan, A., Hossain, S., and Sunny. (2015). Silver nanoparticles synthesis, properties, applications and future perspectives: a short review. IOSR J Elect. Electronics Engg (iosrjeee), 10(6)
- [25] Ortiz, E., P. Roque-Ruiz, J., H. Hinojos-Márquez, A., E. Esparza, J., L. Cornejo, D., A.González, J., C. Cristóbal, E., F.and López, S., Y. R. (2017). Dosedependent antimicrobial activity of silver nanoparticles on polycaprolactone fibers against Grampositive and Gram-negative bacteria. Hindawi J Nanomat (2017). Article ID 4752314, 9 pages https://doi.org/10.1155/2017/4752314
- [26] Ottoni, C., Simões, Fernandes, M., Santos, Silva, E.Rodrigo Fernando Souza, B., and Maiorano, A.(2017) Screening of filamentous fungi for antimicrobial silver nanoparticles synthesis, Ottoni et

# Volume 11 Issue 5, May 2022

# <u>www.ijsr.net</u>

# Licensed Under Creative Commons Attribution CC BY

al. AMB Expr (2017) 7:31 DOI 10.1186/s13568017-0332-2.

- [27] Parveen, K., Banse, V., and Ledwani, L. (2016).Green synthesis of nanoparticles: their advantages and disadvantages, 1724, 020048; doi: 10.1063/1.4945168.
- [28] Paulkumar, Gnanajobitha1, G., Vanaja1, Manickam, M., Pavunraj, and Annadurai, G.(2017).Green synthesis of silver nanoparticle and silver based chitosan bionanocomposite using stem extract of Saccharum officinarum and assessment of its antibacterial activity, Adv. Nat. Sci.: Nanosci. Nanotechnol. 8.
- [29] Ramya, M., and Subapriya, M. (2012). Green synthesis of silver nanoparticles, Intntl. J. Pharma Med. Biol. Sci. 1: No.1.

DOI: 10.21275/SR22518085907