

# Effect of Various Factors on Cadmium Tolerance of *Cupriavidus* sp. HMT 35

Harshada Joshi

Department of Biotechnology, Vigyan Bhawan, Block B, Mohanlal Sukhadia University, Udaipur, Rajasthan, India  
Email: [hjbiotech\[at\]gmail.com](mailto:hjbiotech[at]gmail.com)

**Abstract:** Heavy metal pollution poses a serious threat to environment disturbing the natural flora and fauna of the nearby area. These contaminated sites can provide efficient pool of microbial diversity to be used for bioremediation of contaminated sites. The present study was conducted to isolate a potential cadmium tolerant strain. *Cupriavidus* sp. HMT 35 strain was isolated from heavy metal contaminated soil collected from Zawar, Udaipur, India. *Cupriavidus* sp. HMT 35 was found cadmium tolerant and showed fairly higher MIC value (1500µg/ml). *Cupriavidus* sp. HMT 35 showed maximum cadmium tolerance in nutrient broth adjusted to pH 7.0 at 37° C after an incubation period of 48h. The isolate has a potential to tolerate high concentration of cadmium. It can be further explored for its possible use in remediation and restoration of contaminated land.

**Keywords:** *Cupriavidus* sp., Cadmium tolerance, bioremediation, heavy metal tolerance

## 1. Introduction

The mining activity has produced a huge amount of metal-rich wastes, caused serious environmental pollution and soil degradation in the world [1, 2]. Excessive concentrations of heavy metals especially cadmium acts as a significant toxic factor to biota in the environment [3]. Cadmium may decrease metabolic activity and diversity as well as affect the qualitative and quantitative structure of microbial communities [4]. Chemical methods for removal of cadmium are quite expensive [5]. Bacterial pool can be an option for effective remediation of cadmium contaminated soil. Several workers have isolated heavy metal tolerant bacteria from various contaminated sites [6, 7]. The common heavy metal tolerant bacteria as reported in previous studies were found to be member of Enterobacteriaceae, Pseudomonadaceae and Flavobacteriaceae family including *Pseudomonas* spp., *Alcaligenes* spp., *Enterobacter* spp. etc. [8, 9]. The possible reasons for the high metal tolerance ability of bacteria surviving in metal-contaminated environments is attributed to some detoxifying mechanisms developed by these strains such as accumulation and adsorption ion exchange on the cell surface, binding and complexation by exopolysaccharides, bioaccumulation and sequestration in cell compartments or heavy metal efflux [10-12].

The present study was designed to study the effect of various factors on cadmium tolerance of *Cupriavidus* sp. HMT 35 isolated from contaminated soil collected from Zawar mining site, Udaipur.

## 2. Materials and Methods

### Bacterial strain and maintenance

*Cupriavidus* sp. HMT 35 strain was isolated from heavy metal contaminated soil collected from Zawar, Udaipur, India. Isolate was identified using 16S rRNA sequencing. The strain was maintained on nutrient agar and activated by growing in nutrient agar at 37° C for 24h.

### Determination of minimal inhibitory concentration

The bacterium was streaked on nutrient agar supplemented with increasing concentrations of cadmium chloride ranging from 100µg/ml to 1500 µg/ml with a difference of 100µg/ml. The petri plates were inoculated with the test culture and incubated at 37°C for 48 h. The colonies were observed after 48 h of incubation.

### Effect of various factors on levels of bacterial tolerance to cadmium

The effect of various factors such as temperature, pH, duration and organic matter was studied on levels of bacterial tolerance to cadmium chloride.

**Effect of temperature:** The nutrient broth containing cadmium chloride was inoculated (1% inoculum) with the bacterial culture. The incubation was done at three different temperatures (10°C, 37°C, 45°C) on a rotary shaker at 175rev/min for 48h. The growth was recorded in terms of optical density at 600nm.

**Effect of pH:** The bacterial culture was inoculated (1% inoculum) in nutrient broth with different pH values (4.0, 7.0, and 9.0) and supplemented with varying concentrations of cadmium chloride. The incubation was carried out on a rotary shaker at 175rev/min at appropriate temperature for 48h. The growth was recorded in terms of optical density at 600nm.

**Effect of duration:** The bacterial culture was inoculated (1% inoculum) in nutrient broth with appropriate pH and supplemented with varying concentrations of cadmium chloride. The incubation was carried out on a rotary shaker at 175rev/min and appropriate temperature for 24h, 48h and 72h. The growth was recorded in terms of optical density at 600nm.

**Effect of tween 80 supplementation:** The bacterial culture was inoculated (1% inoculum) in nutrient broth with appropriate pH and supplemented with varying concentrations of cadmium chloride. In addition, varying concentrations (0.01, 0.1, 1mg/ml) of tween 80 was also

supplemented to nutrient broth. The incubation was carried out on a rotary shaker at 175rev/min and appropriate temperature and duration. The growth was recorded as O. D. at 600nm.

All the experiments were done in triplicates and statistical analysis was done using ANOVA.

### 3. Results

The bacterium was streaked on nutrient agar supplemented with increasing concentrations of cadmium chloride ranging from 100µg/ml to 2000 µg/ml with a difference of 100µg/ml. The petri plates were inoculated with the test culture and incubated at 37°C for 48 h. The well-defined colonies were observed after 48 h of incubation. *Cupriavidussp.* HMT 35 showed fairly higher MIC value (1500µg/ml).

The effect of various factors such as temperature, pH, duration and organic matter on the cadmium tolerance level of *Cupriavidussp.* HMT 35 was studied.

**Effect of temperature:** *Cupriavidussp.* HMT 35 was inoculated in nutrient broth supplemented with varying concentrations of cadmium chloride (0,  $8 \times 10^2$ ,  $16 \times 10^2$  and  $24 \times 10^2$  µg/ml) and incubated at different temperatures (10°C, 37°C, 45°C) to determine an optimum temperature at which highest metal tolerance can be seen. The incubation was carried out on a rotary shaker at 175rev/min for 48h. The growth was recorded in terms of optical density at 600nm.

Analysis of variance (ANOVA) revealed that when the cadmium concentration was applied at different temperature the difference between concentration, between temperature and their interaction was significant. The effect of temperature on cadmium tolerance (growth) level of *Cupriavidussp.* HMT 35 is presented in Fig.1. The isolate was able to grow well at 37°C but exhibit very little growth at 10°C and 45°C. The growth of isolate was gradually decreased with an increase in concentration of cadmium chloride ( $\text{CdCl}_2$ ) for all the three temperatures (10°C, 37°C, 45°C). No growth was observed in nutrient broth supplemented with  $24 \times 10^2$  µg/ml of  $\text{CdCl}_2$  even when incubated up to 96 h (which is indicated by the almost constant O. D. values) The isolate showed better tolerance (growth) at 37°C for all the concentrations of cadmium chloride as compared to other incubation temperatures. Therefore, 37°C temperature was found to be the best at which maximum tolerance (maximum growth) to cadmium was observed by *Cupriavidussp.* HMT 35.

**Effect of pH:** The cadmium tolerant *Cupriavidussp.* HMT 35 was inoculated in nutrient broth with different pH values (4.0, 7.0, 9.0) and supplemented with varying concentrations of cadmium chloride (0,  $8 \times 10^2$ ,  $16 \times 10^2$  and  $24 \times 10^2$  µg/ml). The incubation was carried out on a rotary shaker at 175rev/min at 37°C for 48h. The growth was recorded in terms of optical density at 600nm.

Analysis of variance (ANOVA) revealed that when the cadmium concentration was applied at different pH the

difference between concentration, between pH and their interaction was significant. The data obtained are presented in Fig.2. It is clear from figure that the isolate was able to grow well in nutrient broth with pH 7 and showed less growth in nutrient broth adjusted to pH 4 and pH 9. The growth of isolate was gradually decreased with an increase in concentration of cadmium chloride ( $\text{CdCl}_2$ ) for all the three pH values (pH 4, pH 7, pH 9). No growth was observed in nutrient broth supplemented with  $24 \times 10^2$  µg/ml of  $\text{CdCl}_2$  even when incubated up to 72 h.

*Cupriavidussp.* HMT 35 showed maximum cadmium tolerance (maximum growth) in nutrient broth adjusted to pH 7.0.

**Effect of duration:** The cadmium tolerant *Cupriavidussp.* HMT 35 was inoculated in nutrient broth with pH 7 and supplemented with varying concentrations of cadmium chloride (0,  $8 \times 10^2$ ,  $16 \times 10^2$  and  $24 \times 10^2$  µg/ml). The incubation was carried out on a rotary shaker at 175rev/min and 37°C for 24h, 48h and 72h. The growth was recorded as O. D. at 600nm. Analysis of variance (ANOVA) revealed that when the cadmium concentration was applied at different duration the difference between cadmium concentration, between duration and their interaction was significant. The data obtained for the effect of duration on cadmium tolerance (growth) level of *Cupriavidussp.* HMT 35 are presented in Fig.3. It is clear from figure that the tolerance (growth) of the isolate increased up to 48h of incubation period for all concentrations of cadmium chloride. After that the growth was gradually decreased up to 72 h of incubation period for all the concentrations of cadmium chloride. No growth of the isolate in broth supplemented with  $24 \times 10^2$  µg/ml of  $\text{CdCl}_2$  even when incubated up to 72h which is indicated by the almost constant O. D. values. Hence, the duration of 48h is considered as the best duration for the growth of this isolate at all the concentration of cadmium chloride. *Cupriavidussp.* HMT 35 showed maximum tolerance to cadmium (maximum growth) when incubated for 48h of duration.

**Effect of tween 80 supplementation:** The cadmium tolerant *Cupriavidussp.* HMT 35 was inoculated in nutrient broth with pH 7 and supplemented with varying concentrations of cadmium chloride (0,  $8 \times 10^2$ ,  $16 \times 10^2$  and  $24 \times 10^2$  µg/ml). In addition, varying concentrations (0.01, 0.1, 1mg/ml) of tween 80 was supplemented to this nutrient broth. The incubation was carried out on a rotary shaker at 175rev/min and 37°C for 48h. The growth was recorded as O. D. at 600nm.

Analysis of variance (ANOVA) revealed that when varying concentration of cadmium was applied at different concentrations of tween 80, the difference between concentration of cadmium was significant. The data obtained to study the effect of varying concentrations of tween 80 on cadmium tolerance (growth) ability of *Cupriavidussp.* HMT 35 is presented in Fig.4. It is clear from figure that there was an increase in the tolerance (growth) of the isolate as compared to control when different concentration of tween 80 (0.01, 0.1, 1mg/ml) were added to nutrient broth supplemented with varying concentrations of cadmium chloride. The fairly high tolerance (maximum growth) of the

isolate was observed on addition of 0.1mg/ml of tween 80 as compared to the other concentrations of tween 80 used in the study. No growth was observed for the isolate in nutrient broth supplemented with  $24 \times 10^2 \mu\text{g/ml}$  of  $\text{CdCl}_2$  even after addition of tween 80 which is indicated by the almost constant O. D. values.

Hence it was found that 0.1mg/ml of tween 80 increases the cadmium tolerance (growth) ability of *Cupriavidus*sp. HMT 35.

#### 4. Discussion

Microorganisms tolerant to stress due to heavy metals can bioremediate soils with metal contamination [13]. We have identified a bacterial strain that can hold high levels of tolerance to cadmium. *Cupriavidus*sp. HMT 35 exhibited MIC of  $1500 \mu\text{g/ml}$  against cadmium chloride. Many workers previously studied MIC of *Cupriavidus* and reported it variably. Zhang *et al.*, [14] reported it to be 7mM for cadmium by *Cupriavidusnecator*YR2. These results are in accordance to our results in the present study. The isolate *Cupriavidus*sp. HMT 35 showed remarkable tolerance against  $\text{CdCl}_2$  at  $37^\circ\text{C}$  in nutrient broth with pH 7 on addition of 0.1% tween 80 when incubated for 48h. The effect of various factors such as temperature, pH, incubation period, presence of certain organic compounds etc. on heavy metal tolerance capability of bacteria is strain specific. Shamim and Rehman [15] studied the effect of temperature and pH on growth of cadmium tolerant *Klebsiellapneumoniae* strain CBL isolated from waste water collected from industrial areas of Shekhupura (Pakistan) on LB agar supplemented with  $100 \mu\text{g}$  of  $\text{Cd}^{+2}$ . They found that  $30^\circ\text{C}$  temperature, pH 7 and 12h of incubation period were best conditions for maximum growth of the strain. Dogenet *al* [16] studied the effect of organic acids (citric acid, glucuronic acid, alginate acid) on growth and exopolymetric (EPS) substance production. They reported that all organic acids increased the growth and EPS production.

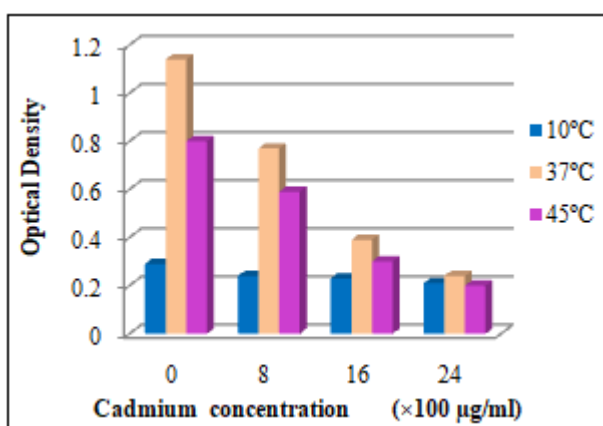


Figure 1: Effect of temperature on cadmium tolerance of *Cupriavidus*sp. HMT 35

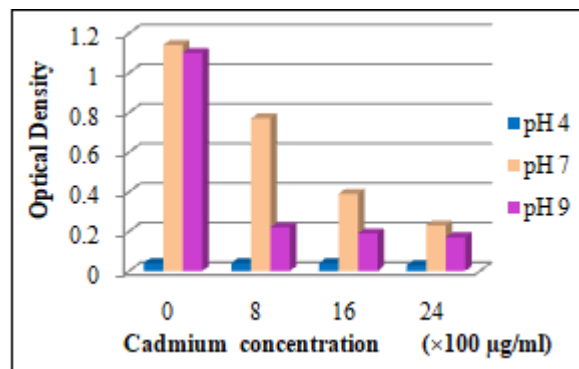


Figure 2: Effect of pH on cadmium tolerance of *Cupriavidus*sp. HMT 35

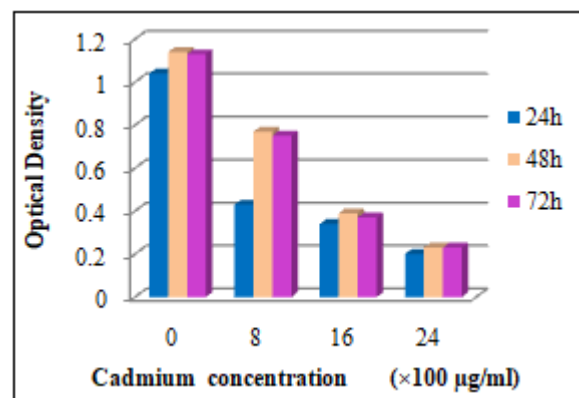


Figure 3: Effect of duration on cadmium tolerance of *Cupriavidus*sp. HMT 35

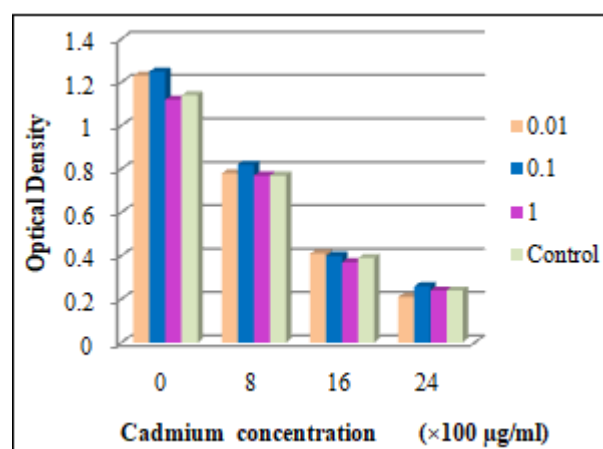


Figure 4: Effect of tween 80 supplementation on cadmium tolerance of *Cupriavidus*sp. HMT35

#### 5. Conclusion

*Cupriavidus*sp. HMT 35 is identified as a good cadmium tolerant strain. Cd detoxification will depend on the uniqueness of the bacterial strains, the biological and chemical properties of Cd formation, as well as the bacterial growth conditions. Optimization studies on isolate for effective cadmium tolerance revealed its potential to tolerate cadmium in normal environmental conditions. Further research can be carried out to use this strain in the bioremediation of contaminated agricultural soils.

## 6. Acknowledgement

SERB-DST, New Delhi is greatly acknowledged for providing financial assistance.

## References

- [1] Sheoran, A. S., and Sheoran, V.2006. Heavy metal removal mechanism of acid mine drainage in wetlands: a critical review. *Miner. Eng.*19: 105-116.
- [2] Li, H. F., Gray, C., Mico, C., Zhao, F. J. and McGrath, S. P., 2009. Phytotoxicity and bioavailability of cobalt to plants in a range of soils. *Chemosphere* 75: 979–986.
- [3] Juan, C. C., María, C. C. and Manuel, C. B., 2018. Biosorption of Cd by non-toxic extracellular polymeric substances (EPS) synthesized by bacteria from marine intertidal biofilms. *Int. J. Environ. Res. Public Health* 15: 314
- [4] Giller, K. E., Witter, E. and McGrath, S. P., 1998. Toxicity of heavy metals to microorganisms and microbial process in agricultural soils: a review. *Soil Biol. Biochem.*30: 1389–1414.
- [5] Yadav, M. M., Singh, G. and Jadeja, R. N., 2021. Physical and chemical methods for heavy metal removal. Hoboken, NJ: Wiley Online. doi: 10.1002/971119693635. ch15
- [6] Piotrowska-Seget, Z., Cycon, M. and Kozdroj, J.2005. Metal-tolerant bacteria occurring in heavily polluted soil and mine spoil. *App Soil Ecol.*28: 237-246.
- [7] Kafilzadeh, F., Moghtaderi, Y., and Johrami, A. R.2013. Isolation and identification of cadmium – resistant bacteria in Soltan Abad river sediments and determination of tolerance of bacteria through MIC and MBC. *Eur J Experimental Biol.*3 (5): 268-273
- [8] Hassen, A., Saidi, N., Cherif, M., and Boudabous, A.1998. Effects of heavy metals on *Pseudomonas aeruginosa* and *Bacillus thuringiensis*. *Biores. Technol.*65: 73-82.
- [9] Mishra, N., Gupta, G. and Jha, P. N.2012. Assessment of mineral phosphate-solubilizing properties and molecular characterization of zinc-tolerant bacteria. *J Basic Microbiol.*52: 549-558.
- [10] Gadd, G. M.1990. Heavy metal accumulation by bacteria and other microorganisms. *Experientia*, 46: 834-840.
- [11] Kanamarlapudi, S. L. R. K., Chintalapudi, K. V. and Muddada, S.2018. Application of biosorption for removal of heavy metals from waste water, biosorption, Jan Derco and Branislav Vrana. London: IntechOpen. doi: 10.5772/intechopen.77315
- [12] Zhang, J., Li, Q., Zeng, Y., Zhang, Jian, Lu, G., et al.2019. Bioaccumulation and distribution of cadmium by *Burkholderiacepacia* GYP1 under oligotrophic condition and mechanism analysis at proteome level. *Ecotoxicol. Environ. Saf.*176: 162–169.
- [13] Ojuederie, O. B. and Babalola, O. O.2017. Microbial and plant-assisted bioremediation of heavy metal polluted environments: A review. *Int. J. Environ. Res. Public Health* 14: 1504.
- [14] Zhang, Q., V. Achal, W. N. Xiang and D. Wang, 2014. Identification of heavy metal resistant bacteria isolated

from Yangtze river, China. *Int. J. Agric. Biol.*, 16: 619–623.

- [15] Shamim, S. and Rehman, A.2012. Cadmium resistance and accumulation potential of *Klebsiella pneumoniae* Strain CBL-1 isolated from industrial waste water. *Pakistan J. Zool.*, 44 (1): 203-208.
- [16] Dogan, N. M, Kanter, C. and Dogan, G.2014. Effect of chromium and organic acids on microbial growth and exopolymetric substance production by *Pseudomonas* bacteria. *Clean-Soil, Air, Water*, 42 (5): 674-681.

## Author Profile



**Dr. Harshada Joshiis** Associate Professor and Course Director, Department of Biotechnology, Mohanlal Sukhadia University, Udaipur (Raj.) India. She worked as CSIR-NET JRF & SRF and subsequently was awarded Ph. D. degree from the same University in the year 2004. She is actively involved in teaching and research in Biotechnology from last 15 years. Her research interests include Molecular and Applied Microbiology & Environmental Microbiology. Six students have been awarded Ph. D. under her supervision and five are working. Dr. Harshada has been awarded and successfully completed three research projects funded by different government agencies. She has authored six books and has published more than 60 research articles in journals of national and international repute. She has presented her research findings in several conferences and symposia in India and abroad. She has visited France, Belgium and Netherlands in connection to academic pursuance. She is member of several professional and academic bodies.